HISTONE DEACETYLASES (HDACs) CLASS II GENES AS POTENTIAL DRUG TARGETS IN CANCER

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Abstract: This review has been undertaken to investigate the role of histone deacetylases (HDACs) class II genes in various cancers. Efforts in cancer research mainly aims to identify the major drug targets, develop and improve treatment regimens which eventually helps into better prognosis. One of the major challenges is the determination and mapping of involvement of molecular pathways. So, there has been large interest in developing pathways and also determining driver genes and regulators. Bioinformatics helps to evaluate huge data from wetlab researches to explore biologically relevant interactions and pathways. Emerging evidence shows that many HDACs exists as biomarkers in cancers. The objective of this study is to present a review on the major and latest findings on class II HDACs and its association in cancers. Here, we summarize the current knowledge of class II HDACs, including their features and classification, biological mechanism as well as their function and reported clinical studies in cancers. In addition, we discuss various trials and clinically available inhibitors targeting them. These results provide valuable insights which helps to identify and develop cancer specific therapeutic options.

Keywords: HDACs, epigenetics, cancer pathways, bioinformatics, drug target identification.

I. INTRODUCTION

Cancer is a is a slow progressing disease eventually involving the uncontrolled division of cells. In normal cells, many genes control the cell growth and division process. During cancer development, cells attains the mode of rapid division, invasion and metastasis. Generally, a tumor forms with the influence of mutation of driver gene's and acts as passengers. These driver genes controls pathways and regulatory mechanisms in the cells. The basis of cancer research is the understanding of the pathways and its underlying mechanism. Several studies shows that, most cancer cells show multiple mutations in its genetic level (1–3). Therefore, studying mutations at domain level studies helps researchers to pin point the exact mechanisms relates to cancer. These Protein domain boundaries and architecture knowledge is important aspect for mainly understanding the protein function. In the present study, we provide a comprehensive review on the role of class II HDACs in cancer. The review summarizes the recent studies of the roles of class II HDACs and we mainly focus on recent discoveries related to molecular mechanisms of HDACs. Overall, the current available information will open new perspectives for a better understanding of HDACs in cancers.

II. EPIGENETICS AND ROLE OF HDACS IN CANCER

Epigenetics is the field which studies the nature of heritable alterations in gene expression (4). These genetic abnormalities are one of the major causes in cancer. In eukaryotic DNA, chromatin is tightly bound with histone protein octamers, which restricts its accessibility to factors involved in DNA replication and transcription. Histone protein octamers or nucleosome contains or 146 bp DNA wrapping around protein complex with eight subunits of individual proteins namely H2A, H2B, H3 and H4 are situated at its center (5), Fig.1.

In 1964, it is found that, post-translational modifications (PTMs) of histones are one of the main cause of gene regulation process in RNA (6) and later it has been shown that the acetylation of lysine residue is regulated by two enzymes namely Histone acyltransferases (HATs) and histone deacetylases (HDACs) (7). These two enzymes function simultaneously and removes acetyl groups from chromatin which is known as chromatin remodeling, which regulates the gene expression (8–10). Histone acetylation and deacetylation, as one of the epigenetic mechanisms, occurs during the development of human cancer.

There are a large number of different histone PTMs such as phosphorylation, acetylation, ubiquitination and methylation in euchromatin. The dynamics of chromatin are regulated by the above processes. Euchromatin is a loosely compact structure (DNA, RNA and protein) which helps transcription factors to bind to regulatory sites on DNA, resulting transcriptional activation (11). In 1997, the crystal structure of the nucleosome was elucidated and proved PTMs causes the inter-nucleosomal interactions which influences transcription and DNA processes such as repair, replication and recombination (12). Further investigations revealed that, epigenetic modifications as well as alterations results in the regulation of gene expression by chromatin remodeling associated with reversible histone acetylation (13–15). HDACs reverse acetylation of chromatin and

altering transcription of oncogenes and tumor suppressor genes (16). Abnormal histone acetylation leads to cancer development which proves histone modifications plays a crucial role in influencing many fundamental biological processes (13). Histone acetylation modulates transcription in many ways. Acetylation of ε -amino groups of lysine residues in histone tails neutralizes their positive charge, thereby relaxing chromatin structure. This helps the transcription occur in a normal manner. The action between these enzymes serves as a key regulatory mechanism for gene expression (17). By removal of acetyl groups from histones, HDACs makes a non-permissive chromatin conformation which prevents the transcription of genes to proteins in cancer development (18). Chromatin assembly plays a major role in transcription (19,20). PTMs of histone proteins have crucial role in cancer development and progression by modulating gene transcription and chromatin remodeling. A lysine residue becomes acetylated by the action of these two enzymes namely, histone acetyltransferase enzymes and is removed by HDACs. Thus, histone lysine acetylation is highly reversible.

Since, the acetylation process is a key mechanism in cancer, studying the role of HDACs is a subject of interest and attracting a great attention from cancer researchers across the globe. Many research groups are working on the drug binding mechanisms and inhibitors search for HDACs. The integral approach of finding inhibitors lies in determining PTMs, identifying protein- protein interactions therefore, of considerable interest.



Fig (1): The nucleosome or histone core shows octamers (H2A, H2B, H3, and H4) with the linker histone (H1). Histone tails are modified through phosphorylation, acetylation, ubiquitination and methylation (13).

III. HDACS CLASSIFICATION

In humans, HDACs enzymes are classified into four classes of 18 individual members based on sequence homology to yeast (21,22). Class I comprises of 1, 2, 3, and 8. Class II divided into two subcategories such as class IIa (4, 5, 7, and 9) and class IIb (6 and 10) as shown in Table 1. Class III known as sirtuins comprises of seven members (1-7) and class IV consists of 11. These have the cofactor Zn^{2+} at the catalytic site and thus regarded as classical zinc-dependent HDACs. These HDACs present in the nucleus and cytoplasm and located in specific tissues such as the brain, heart, and muscle. Expression of class II HDACs has been found in many human cancers, including gastric cancer, lung cancer, liver cancer, prostate cancer and leukemia. In this review we will discuss about class II HDACs and evaluate the important finding and scientific interest in recent years. We searched recent research articles using keywords such as 'hdacs cancer and process', 'cancer and therapeutic options of hdacs' in pubmed and other literatures. Subsequently, the articles that matched such word criteria were completely reviewed and their findings noted.

Class II member	No. of	UniprotKb Id	PDB Id	Cancer type
	Residues			
HDAC4	1084	P56524	2VQJ	Gastric cancer, Lung cancer, Liver cancer, Prostate cancer
HDAC5	1122	Q9UQL6	5UWI	Breast cancer Hepatocellular carcinoma Ovarian cancer
HDAC7	952	Q8WUI4	3C10	Lung cancer Colorectal cancer Leukemia
HDAC9	1011	Q9UKV0	No crystal structure available yet	Breast cancer, Oral cancer Leukemia
HDAC6	1215	Q9UBN7	3PHD	Gastric cancer Breast cancer, Cervical cancer
HDAC10	669	Q969S8	5TD7	Lung cancer Leukemia, Gastric cancer

Table 1: Overview of Class II HDACs

3.1 Class II a member, histone deacetylase 4 (HDAC4)

It is found that, HDAC4 promotes gastric cancer cell progression via p21 repression (23) and involvement of the MIAT/miR-29a-3p/HDAC4axis (24). Cancer upregulated gene 2 (CUG2) via HDAC4 signalling has been reported in lung cancer (25) and HDAC4 regulates HIF1 α protein acetylation is shown in liver cancer (26) (27). Both HDAC4 and HDAC6 together with the TGF- β /Smad pathway involved in progression of glioblastoma, a type of tumor in nervous system (28). Class IIa HDACs are overexpressed in 22% of in leiomyosarcomas (LMS), where high levels of the myocyte enhancer factor 2 (MEF2), HDAC4 and HDAC9 expression is shown (29). MEF2 is a signalling pathway which control cell differentiation and organogenesis. Strong correlation has been proved in the expression levels of HDAC4 in acute myeloid leukemia (AML) (30).

3.2 Class II a member, histone deacetylase 5 (HDAC5)

HDAC6 has been identified as a promising biomarker for early detection of breast cancer by evaluating its higher expression levels in inferior prognosis (31) and in lung cancer cells (32). Several studies implicates its role in colorectal cancer cell lines (33), in hepatocellular carcinoma (34) and in breast cancer (35). In a study by yano et al, immunohistochemical analyses of major HDACs were performed to determine the expression using tissue microarrays data in prognosis of ovarian cancer (36).

3.3 Class II a member, histone deacetylase 7 (HDAC7)

HDAC7 has been experimentally studied using tube formation assay, immunofluorescence, microarray and western blot analysis to demonstrate angiogenesis process of endothelial progenitor cells (EPCs) in lung cancer (37). It is found that, HDAC7 promotes lung cancer by deacetylating stat3 leads to its inhibition (38). Studies confirms that, HDAC7 is overexpressed cancer stem cells (CSCs) (39). Studies confirms that, HDAC7 is overexpressed cancer stem cells (CSCs) (39). MiR-489 play a key role as suppressor of tumor growth and invasion in colorectal cancer by targeting HDAC7 (40). Isobaric labeling and mass spectrometry proteomics on chronic lymphocytic leukemia (CLL) samples have provided evidence linking the overexpression of HDAC7 (41) (42) and its deregulation contribute to the pathogenesis of the diseases leukemia and lymphoma (43).

3.4 Class II a member, histone deacetylase 9 (HDAC9)

HDAC9 is identified as an oncogene and its elevated expression level was evaluated in breast cancer patients (44) and in triple negative breast cancer (TNBC) HDAC9 acts as a mediator of cell invasion and angiogenesis (45). HDAC9 is found to be as a target of miR-377 and its target, NR4A1/Nur77 in Oral Squamous Cell Carcinoma (OSCS) (46). HDAC9 is found as overexpressed in acute lymphoblastic leukaemia (ALL) (47), acute myeloid leukaemia (AML) (48) and in chronic lymphocytic leukemia (CLL) (49).

3.5 Class II b member, histone deacetylase 6 (HDAC6)

The aberrant regulation of HDAC6 results the progression of malignancy in various cancers. HDAC6 has been identified as a promising biomarker for early detection of gastric cancer (50). The expression levels of HDAC6 and androgen receptor have strong correlation in breast cancer however its role is not clearly identified (51). G-protein-coupled receptor kinase 2 (GRK2), a key player in breast cancer acts as a regulator in HDAC6 mediated breast cancer. Its increased functionality and phosphorylation activates HDAC6 which leads to tumor progression (52). HDAC6 possess redundant effect on HDAC8 in cervical cancer cells (53).

3.6 Class II b member, histone deacetylase 10 (HDAC10)

HDAC10 was identified from a human cDNA library (20,54). In recent years, a number of studies have shown the mechanisms and functions of HDAC10 protein in cancer cells, such as autophagy (55), cell cycle (56) immunoregulation and specially, HDAC10 serve as oncogenic stimuli or tumor suppressors in cancer. Recent study by Shinksy et al shows that, HDAC10 is an emerging biomarker in cancers and considered as a target for the isozyme-selective inhibitors for design of autophagy mediated responses to cancer treatment (57). Studies shows that increased HDAC10 expression leads to increased phosphorylation of AKT in lung cancer (56), leukemia and lymphoma (58). HDAC10 regulates cyclin A2 expression and represses transcription which reveals its potential as targets in diseases associated with cell cycle dysregulation (59).

IV. TRIALS AND COMPOUNDS SCREENING STUDIES TOWARDS CLASS II HDACs

Li et al investigated the effects of two synthetic curcumin derivatives to restore Nrf2 activity in TRAMP C1 cells, which are hypermethylated in prostrate cancer (60). Enzyme inhibitory activity against HDAC isoforms revealed nonhydroxamate compounds shown strong inhibitory activity against class IIa HDACs especially HDAC4 (61) and strong inhibitory activity against HDAC6 (62). Two ebselen analogs namely, ebselen oxide and ebsulfur inhibited HDAC4, HDAC5, HDAC6, HDAC7 and HDAC9 (63). A flavone component namely, 3,4'-dimethoxy-3',5,7-trihydroxyflavone induces cell cycle arrest of MCF-7 in HDAC4 mediated breast cancer Cells (64). (-)-Epigallocatechin gallate (EGCG) is a component from a green tea catechin which changes the levels of acetylation of histones in and suppresses the activity of HDAC4, 5 and 6 in lung cancer (65). Screening of epigenetic modifying drugs showed, sulforaphane, downregulated HDAC5 transcription by blocking USF1 activity in breast cancer (66). Apicidin selectively reduced HDAC7 siRNAs expression in salivary mucoepidermoid carcinoma (MEC) cells (67). Antiproliferative effect of A452 was found effective for colorectal cancer treatment (68). ACY-1215 (ricolinostat) which is a known HDAC6 inhibitor, which is undergoing clinical trials for hematological malignancies (69). A series of synthetic compounds were tested and TC24 which showed strong anti-proliferative ability towards HDAC6 mediated gastric cancer cells (70). Similarly, a compound named B-R2B was identified through in silico method and its possible HDAC6 inhibitory action specifically anti proliferative activity on cervical cancer and leukemia cells were evaluated based on a reference compound namely tubacin. According to molecular dynamics (MD) simulations, the compound showed a non-competitive effect in a similar manner that tubacin does (71).

V. DISCUSSION

In this paper, we describe HDACs, which plays an important role in cancer by deacetylation process of lysine residue. Extensive research over the past years indicates their role in cell cycle progression, apoptosis, and autophagy. From our review results, it is clear that cancer development is a complex and heterogenous process which involves different steps. Researchers have identified several signalling pathways has been reported to promote the cancer. These signalling mechanisms are predominantly activated by modulation through various pathways. This creates a complexity within cancer cells. Tissue type and specificity in signalling mechanism increases the level of complexity in cancers, thus there is high demand for identification of targets and its inhibition mechanism.

VI. CONCLUSION

In the recent years, there has been an increased interest in the field of HDACs research in cancer treatments. This review article focuses relatively on the latest findings towards class II HDACs in cancer research and majority of studies shows that pathway analysis contributed significantly. There are more publications related to class II mediated HDACs and their role in cancers. The latest evidences of clinical information that have discussed in this study may provide ample observations for further studies on its molecular mechanisms. Like nearly all enzymes that are involved in critical cellular functions, the activities of HDACs are highly regulated.

VII. ACKNOWLEDGEMENTS

The authors acknowledge JNIAS, Telangana, India for providing the necessary support and facilities.

VIII. CONFLICT OF INTEREST None

REFERENCES

- 1. Tomlinson I, Sasieni P, Bodmer W. How Many Mutations in a Cancer? Am J Pathol. 2002;160(3):755–8.
- 2. Loeb KR, Loeb LA. Significance of multiple mutations in cancer. Carcinogenesis. 2000;21(3):379–85.
- 3. Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA, Kinzler KW. Cancer Genome Landscapes. Science.

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2013;339(6127):1546-58.

- 4. Lakshmaiah KC, Jacob LA, Aparna S, Lokanatha D, Saldanha SC. Epigenetic therapy of cancer with histone deacetylase inhibitors. J Cancer Res Ther. 2014;10(3):469–78.
- 5. Sun W-J, Zhou X, Zheng J-H, Lu M-D, Nie J-Y, Yang X-J, et al. Histone acetyltransferases and deacetylases: molecular and clinical implications to gastrointestinal carcinogenesis. Acta Biochim Biophys Sin (Shanghai). 2012;44(1):80–91.
- 6. Allfrey VG, Mirsky AE. Structural Modifications of Histones and their Possible Role in the Regulation of RNA Synthesis. Science. 1964;144(3618):559.
- 7. Yang X-J, Seto E. HATs and HDACs: from structure, function and regulation to novel strategies for therapy and prevention. Oncogene. 2007;26:5310.
- 8. Rundlett SE, Carmen AA, Kobayashi R, Bavykin S, Turner BM, Grunstein M. HDA1 and RPD3 are members of distinct yeast histone deacetylase complexes that regulate silencing and transcription. Proc Natl Acad Sci U S A. 1996;93(25):14503–8.
- 9. Mizzen CA, Yang XJ, Kokubo T, Brownell JE, Bannister AJ, Owen-Hughes T, et al. The TAF(II)250 subunit of TFIID has histone acetyltransferase activity. Cell. 1996;87(7):1261–70.
- 10. Okamoto M, Takemori H, Katoh Y. Salt-inducible kinase in steroidogenesis and adipogenesis. Trends Endocrinol Metab. 2004;15(1):21–6.
- 11. Fedorova E, Zink D. Nuclear architecture and gene regulation. Biochim Biophys Acta Mol Cell Res. 2008;1783(11):2174–84.
- 12. Luger K, Mader AW, Richmond RK, Sargent DF, Richmond TJ. Crystal structure of the nucleosome core particle at 2.8 A resolution. Nature. 1997;389(6648):251–60.
- 13. Ramakrishnan S, Pili R. Histone Deacetylase Inhibitors and Epigenetic Modifications as a Novel Strategy in Renal Cell Carcinoma. Cancer J. 2013;19(4):333–40.
- 14. Bolger TA, Cohen T, Yao T-P. HATs and HDACs BT Gene Expression and Regulation. In 2006. p. 111–33.
- 15. Yamada T, Mizuno K, Hirota K, Kon N, Wahls WP, Hartsuiker E, et al. Roles of histone acetylation and chromatin remodeling factor in a meiotic recombination hotspot. EMBO J. 2004;23(8):1792–803.
- 16. Li Y, Seto E. HDACs and HDAC Inhibitors in Cancer Development and Therapy. Cold Spring Harb Perspect Med. 2016;6(10).
- 17. Haberland M, Montgomery RL, Olson EN. The many roles of histone deacetylases in development and physiology: implications for disease and therapy. Nat Rev Genet. 2009 Jan;10(1):32–42.
- 18. Glozak MA, Seto E. Histone deacetylases and cancer. Oncogene. 2007;26(37):5420–32.
- 19. Krude T, Keller C. Chromatin assembly during S phase: contributions from histone deposition, DNA replication and the cell division cycle. Cell Mol Life Sci. 2001;58(5–6):665–72.
- 20. Fischer DD, Cai R, Bhatia U, Asselbergs FAM, Song C, Terry R, et al. Isolation and characterization of a novel class II histone deacetylase, HDAC10. J Biol Chem. 2002;277(8):6656–66.
- 21. Seto E, Yoshida M. Erasers of histone acetylation: the histone deacetylase enzymes. Cold Spring Harb Perspect Biol. 2014;6(4):a018713.
- 22. Cho Y, Griswold A, Campbell C, Min K-T. Individual histone deacetylases in Drosophila modulate transcription of distinct genes. Genomics. 2005;86(5):606–17.
- 23. Kang Z-H, Wang C-Y, Zhang W-L, Zhang J-T, Yuan C-H, Zhao P-W, et al. Histone deacetylase HDAC4 promotes gastric cancer SGC-7901 cells progression via p21 repression. PLoS One. 2014;9(6):e98894.
- 24. Li Y, Wang K, Wei Y, Yao Q, Zhang Q, Qu H, et al. lncRNA-MIAT regulates cell biological behaviors in gastric cancer through a mechanism involving the miR-29a-3p/HDAC4 axis. Oncol Rep. 2017;38(6):3465–72.
- 25. Kaowinn S, Jun SW, Kim CS, Shin D-M, Hwang Y-H, Kim K, et al. Increased EGFR expression induced by a novel oncogene, CUG2, confers resistance to doxorubicin through Stat1-HDAC4 signaling. Cell Oncol. 2017;40(6):549–61.
- 26. Tsai C-L, Liu W-L, Hsu F-M, Yang P-S, Yen R-F, Tzen K-Y, et al. Targeting histone deacetylase 4/ubiquitin-conjugating enzyme 9 impairs DNA repair for radiosensitization of hepatocellular carcinoma cells in mice. Hepatology. United States; 2017 Jun;
- 27. Geng H, Harvey CT, Pittsenbarger J, Liu Q, Beer TM, Xue C, et al. HDAC4 Protein Regulates HIF1α Protein Lysine Acetylation and Cancer Cell Response to Hypoxia. J Biol Chem. 2011;286(44):38095–102.
- 28. Sferra R, Pompili S, Festuccia C, Marampon F, Gravina GL, Ventura L, et al. The possible prognostic role of histone deacetylase and transforming growth factor beta/Smad signaling in high grade gliomas treated by radio-chemotherapy: a preliminary immunohistochemical study. Eur J Histochem. 2017;61(2):2732.
- 29. Di Giorgio E, Franforte E, Cefalu S, Rossi S, Dei Tos AP, Brenca M, et al. The co-existence of transcriptional activator and transcriptional repressor MEF2 complexes influences tumor aggressiveness. PLoS Genet. 2017;13(4):e1006752.
- 30. Gaal Z, Olah E, Rejto L, Erdodi F, Csernoch L. Strong Correlation between the Expression Levels of HDAC4 and SIRT6 in Hematological Malignancies of the Adults. Pathol Oncol Res. 2017;23(3):493–504.
- 31. Li A, Liu Z, Li M, Zhou S, Xu Y, Xiao Y, et al. HDAC5, a potential therapeutic target and prognostic biomarker, promotes proliferation, invasion and migration in human breast cancer. Oncotarget. 2016;7(25):37966–78.
- 32. Zhao M, Li L, Zhou J, Cui X, Tian Q, Jin Y, et al. MiR-2861 Behaves as a Biomarker of Lung Cancer Stem Cells and Regulates the HDAC5-ERK System Genes. Cell Reprogram. 2018;20(2):99–106.
- 33. He P, Liang J, Shao T, Guo Y, Hou Y, Li Y. HDAC5 promotes colorectal cancer cell proliferation by up-regulating DLL4 expression. Int J Clin Exp Med. 2015;8(4):6510–6.
- 34. Gu H, Fang Z, Cai X, Song R, Lin M, Ye J, et al. Highly expressed histone deacetylase 5 promotes the growth of

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hepatocellular carcinoma cells by inhibiting the TAp63-maspin pathway. Am J Cancer Res. 2018;8(3):462–75.

- 35. Cao C, Vasilatos SN, Bhargava R, Fine JL, Oesterreich S, Davidson NE, et al. Functional interaction of histone deacetylase 5 (HDAC5) and lysine-specific demethylase 1 (LSD1) promotes breast cancer progression. Oncogene. 2017;36(1):133–45.
- 36. Yano M, Yasuda M, Sakaki M, Nagata K, Fujino T, Arai E, et al. Association of histone deacetylase expression with histology and prognosis of ovarian cancer. Oncol Lett. 2018;15(3):3524–31.
- Wei Y, Zhou F, Zhou H, Huang J, Yu D, Wu G. Endothelial progenitor cells contribute to neovascularization of nonsmall cell lung cancer via histone deacetylase 7-mediated cytoskeleton regulation and angiogenic genes transcription. Int J cancer. 2018;
- 38. Lei Y, Liu L, Zhang S, Guo S, Li X, Wang J, et al. Hdac7 promotes lung tumorigenesis by inhibiting Stat3 activation. Mol Cancer. 2017;16(1):170.
- 39. Witt AE, Lee C-W, Lee TI, Azzam DJ, Wang B, Caslini C, et al. Identification of a cancer stem cell-specific function for the histone deacetylases, HDAC1 and HDAC7, in breast and ovarian cancer. Oncogene. 2017;36(12):1707–20.
- 40. Gao S, Liu H, Hou S, Wu L, Yang Z, Shen J, et al. MiR-489 suppresses tumor growth and invasion by targeting HDAC7 in colorectal cancer. Clin Transl Oncol. 2018;20(6):703–12.
- 41. Johnston HE, Carter MJ, Larrayoz M, Clarke J, Garbis SD, Oscier D, et al. Proteomics Profiling of CLL Versus Healthy B-cells Identifies Putative Therapeutic Targets and a Subtype-independent Signature of Spliceosome Dysregulation. Mol Cell Proteomics. 2018;17(4):776–91.
- 42. Zhou K, Zhang Q, Liu Y, Xiong Y, Wu S, Yang J, et al. Aberrant histone modification in CD19(+) B cells of patients with chronic lymphocytic leukemia. Onco Targets Ther. 2017;10:1173–9.
- 43. Barneda-Zahonero B, Collazo O, Azagra A, Fernandez-Duran I, Serra-Musach J, Islam ABMMK, et al. The transcriptional repressor HDAC7 promotes apoptosis and c-Myc downregulation in particular types of leukemia and lymphoma. Cell Death Dis. 2015;6:e1635.
- 44. Huang Y, Jian W, Zhao J, Wang G. Overexpression of HDAC9 is associated with poor prognosis and tumor progression of breast cancer in Chinese females. Onco Targets Ther. 2018;11:2177–84.
- 45. Salgado E, Bian X, Feng A, Shim H, Liang Z. HDAC9 overexpression confers invasive and angiogenic potential to triple negative breast cancer cells via modulating microRNA-206. Biochem Biophys Res Commun. 2018;
- 46. Rastogi B, Kumar A, Raut SK, Panda NK, Rattan V, Joshi N, et al. Downregulation of miR-377 Promotes Oral Squamous Cell Carcinoma Growth and Migration by Targeting HDAC9. Cancer Invest. 2017;35(3):152–62.
- 47. Moreno DA, Scrideli CA, Cortez MAA, de Paula Queiroz R, Valera ET, da Silva Silveira V, et al. Differential expression of HDAC3, HDAC7 and HDAC9 is associated with prognosis and survival in childhood acute lymphoblastic leukaemia. Br J Haematol. 2010;150(6):665–73.
- 48. Almamun M, Levinson BT, van Swaay AC, Johnson NT, McKay SD, Arthur GL, et al. Integrated methylome and transcriptome analysis reveals novel regulatory elements in pediatric acute lymphoblastic leukemia. Epigenetics. 2015;10(9):882–90.
- 49. Cahill N, Bergh A-C, Kanduri M, Goransson-Kultima H, Mansouri L, Isaksson A, et al. 450K-array analysis of chronic lymphocytic leukemia cells reveals global DNA methylation to be relatively stable over time and similar in resting and proliferative compartments. Leukemia. 2013;27(1):150–8.
- 50. He Q, Li G, Wang X, Wang S, Hu J, Yang L, et al. A Decrease of Histone Deacetylase 6 Expression Caused by Helicobacter Pylori Infection is Associated with Oncogenic Transformation in Gastric Cancer. Cell Physiol Biochem. 2017;42(4):1326–35.
- 51. Li C, Cao L, Xu C, Liu F, Xiang G, Liu X, et al. The immunohistochemical expression and potential prognostic value of HDAC6 and AR in invasive breast cancer. Hum Pathol. 2018;75:16–25.
- 52. Nogues L, Reglero C, Rivas V, Salcedo A, Lafarga V, Neves M, et al. G Protein-coupled Receptor Kinase 2 (GRK2) Promotes Breast Tumorigenesis Through a HDAC6-Pin1 Axis. EBioMedicine. 2016;13:132–45.
- 53. Vanaja GR, Ramulu HG, Kalle AM. Overexpressed HDAC8 in cervical cancer cells shows functional redundancy of tubulin deacetylation with HDAC6. Cell Commun Signal. 2018;16(1):20.
- 54. Verdin E, Dequiedt F, Kasler HG. Class II histone deacetylases: versatile regulators. Trends Genet. 2003;19(5):286–93.
- 55. Oehme I, Lodrini M, Brady NR, Witt O. Histone deacetylase 10-promoted autophagy as a druggable point of interference to improve the treatment response of advanced neuroblastomas. Autophagy. 2013.
- 56. Yang Y, Huang Y, Wang Z, Wang H, Duan B, Ye D, et al. HDAC10 promotes lung cancer proliferation via AKT phosphorylation. Oncotarget. 2016;7(37):59388–401.
- 57. Shinsky SA, Christianson DW. Polyamine Deacetylase Structure and Catalysis: Prokaryotic Acetylpolyamine Amidohydrolase and Eukaryotic HDAC10. Biochemistry. 2018;57(22):3105–14.
- 58. Powers J, Lienlaf M, Perez-Villarroel P, Deng S, Knox T, Villagra A, et al. Expression and Function of Histone Deacetylase 10 (HDAC10) in B Cell Malignancies. Methods Mol Biol. 2016;1436:129–45.
- 59. Li Y, Peng L, Seto E. Histone Deacetylase 10 Regulates the Cell Cycle G2/M Phase Transition via a Novel Let-7-HMGA2-Cyclin A2 Pathway. Mol Cell Biol. 2015;35(20):3547–65.
- 60. Li W, Su Z-Y, Guo Y, Zhang C, Wu R, Gao L, et al. Curcumin Derivative Epigenetically Reactivates Nrf2 Antioxidative Stress Signaling in Mouse Prostate Cancer TRAMP C1 Cells. Chem Res Toxicol. 2018;31(2):88–96.
- 61. Hsu K-C, Liu C-Y, Lin TE, Hsieh J-H, Sung T-Y, Tseng H-J, et al. Novel Class IIa-Selective Histone Deacetylase Inhibitors Discovered Using an in Silico Virtual Screening Approach. Sci Rep. 2017;7(1):3228.
- 62. Stenzel K, Hamacher A, Hansen FK, Gertzen CGW, Senger J, Marquardt V, et al. Alkoxyurea-Based Histone

Deacetylase Inhibitors Increase Cisplatin Potency in Chemoresistant Cancer Cell Lines. J Med Chem. 2017;60(13):5334–48.

- 63. Wang Y, Wallach J, Duane S, Wang Y, Wu J, Wang J, et al. Developing selective histone deacetylases (HDACs) inhibitors through ebselen and analogs. Drug Des Devel Ther. 2017;11:1369–82.
- 64. Weng J-R, Bai L-Y, Lin W-Y, Chiu C-F, Chen Y-C, Chao S-W, et al. A Flavone Constituent from Myoporum bontioides Induces M-Phase Cell Cycle Arrest of MCF-7 Breast Cancer Cells. Molecules. 2017;22(3).
- 65. Oya Y, Mondal A, Rawangkan A, Umsumarng S, Iida K, Watanabe T, et al. Down-regulation of histone deacetylase 4, -5 and -6 as a mechanism of synergistic enhancement of apoptosis in human lung cancer cells treated with the combination of a synthetic retinoid, Am80 and green tea catechin. J Nutr Biochem. 2017;42:7–16.
- 66. Cao C, Wu H, Vasilatos SN, Chandran U, Qin Y, Wan Y, et al. HDAC5-LSD1 axis regulates antineoplastic effect of natural HDAC inhibitor sulforaphane in human breast cancer cells. Int J cancer. 2018;
- 67. Ahn M-Y, Yoon J-H. Histone deacetylase 7 silencing induces apoptosis and autophagy in salivary mucoepidermoid carcinoma cells. J Oral Pathol Med. 2017;46(4):276–83.
- 68. Won H-R, Ryu H-W, Shin D-H, Yeon S-K, Lee DH, Kwon SH. A452, an HDAC6-selective inhibitor, synergistically enhances the anticancer activity of chemotherapeutic agents in colorectal cancer cells. Mol Carcinog. 2018;
- 69. Lee DH, Won H-R, Ryu H-W, Han JM, Kwon SH. The HDAC6 inhibitor ACY1215 enhances the anticancer activity of oxaliplatin in colorectal cancer cells. Int J Oncol. 2018;
- 70. Dong J, Zheng N, Wang X, Tang C, Yan P, Zhou H-B, et al. A novel HDAC6 inhibitor exerts an anti-cancer effect by triggering cell cycle arrest and apoptosis in gastric cancer. Eur J Pharmacol. 2018;828:67–79.
- 71. Alberto R-FR, Yudibeth S-L, Jonathan F-VM, Raul F-M, Cristina C-PL, Ismael V-M, et al. Design, Synthesis and Biological Evaluation of a Phenyl Butyric Acid Derivative, N-(4-chlorophenyl)-4-phenylbutanamide: A HDAC6 Inhibitor with Anti-proliferative Activity on Cervix Cancer and Leukemia Cells. Anticancer Agents Med Chem. 2017;17(10):1441–54.

