

REVIEW ON CARDIO PROTECTIVE AND ANTI- OXIDANT ACTIVITY OF PAULOWNIA TOMENTOSA PLANT

Sudarshan Singh^{1*}, Arun Dev Pokharel¹, Mrs. Kusu Susan Cyriac²

Karnataka College of pharmacy #33/2, Thirumenahalli, Bangalore – 560064, India. Department of pharmacology.

Corresponding author: sudarshan singh

ABSTRACT

Paulownia tomentosa is one of the most useful trees in China. Various parts (leaves, flowers, fruits, wood, bark, roots and seeds) of Paulownia have been used for treating a variety of ailments and diseases. Each of these parts has been shown to contain one or more bioactive components, such as ursolic acid and mattheucinol in the leaves; Paulownia and dsesaminin in the wood/xylem; syringin and catalpinoside in the bark. The fruits contain fatty oils, alkaloids, flavonones as well as flavonoids with antioxidant properties. In vitro grown Paulownia fortunei Hemsl. Seedlings, inoculated with Agrobacterium rhizogenes have a potential to produce hairy roots and synthesize bioactive compounds such as acteosides. With various new studies describing isolation of therapeutic compounds and their probable application in human health, it is an opportune moment to revisit medicinal potential of this tree.

KEYWORDS

Paulownia tomentosa, cardioprotective activity, antioxidant activity, 5-Fluorouracil induced cardio toxic rats.

INTRODUCTION

Cardiovascular disease (CVD) is a group of disorders/diseases of the heart and blood vessels, including heart attack and stroke. Cardiovascular diseases include: coronary heart disease (heart attacks), cerebrovascular disease, raised blood pressure (hypertension), peripheral artery disease, rheumatic heart disease, congenital heart disease, and heart failure^[1].

Cardiovascular diseases (CVDs) continue to be a leading cause of morbidity and mortality among adults around the world. Risk factors have included blood pressure, cigarette smoking, Total Cholesterol (TC), Low Density Lipoprotein Cholesterol (LDL-C), High Density Lipoprotein Cholesterol (HDL-C), and Diabetes Mellitus. Factors such as obesity, left ventricular hypertrophy, family history of premature CVD, and Estrogen Replacement Therapy (ERT) have also been considered in defining CVD risk. Data from population studies enabled prediction of Coronary Heart Disease (CHD) during a follow-interval of several years, based on blood pressure, smoking history, TC, LDL-C and HDL-C levels, Diabetes Mellitus, and left ventricular hypertrophy on the electrocardiogram ECG^[2].

Cardiovascular disease is considered as the leading cause of death among high-income countries and is projected to be the leading cause of death worldwide by 2030. Much of the current research efforts have been aimed toward the identification, modification and treatment of individual-level risk factors^[3].

Cardio toxicity is the occurrence of heart electrophysiology dysfunction or muscle damage. The heart becomes weaker and is not as efficient in pumping and therefore circulating blood^[4].

Paulownia tomentosa belongs to the family Scrophulariaceae, it is commonly known as princess tree [5]. *Paulownia tomentosa* is a genus that is composed of a number of species. They are all native to China except *P. fortune* which extends into Vietnam and Laos. *Paulownia tomentosa*. Also name a princess or empress tree [6]. *P. tomentosa* is a rich source of Geranylated flavanones, furanoquinones, iridoides and flavonoids were also present. *Paulownia tomentosa* is a large indeciduous tree planted mostly for its fast growing wood and decoratative purposes. The tree is also used in traditional Chinese medicine. As a part of our study of natural polyphenols, the fruits of *Paulownia tomentosa* although with the development of chemical and pharmacological analysis methods we can study natural compounds more thoroughly, a great number of compounds and plants still remain unexplored. One such plant is *Paulownia tomentosa*. In this plant only a few compounds have been identified, mostly of polar character and divided into five groups: phenolic glycosides, furofuran lignanes, furanoquinones, iridoides and flavonoids. A large group of essential oil substances has also been identified in the flowers. These compounds and especially the flavonoids were identified from different species, where they probably serve as UV irradiation protectors. The increase in free radical species (corresponding with a high UV irradiation) is nowadays considered to be the true cause and effect of many metabolic disorders connected to such diseases as neurodegeneration, cancer and diabetes mellitus. A screening assay of *P. tomentosa* fruit extracts showed an antiradical effect [7].

Glycosides were the most abundant compounds isolated from the secretions of glandular hairs on the leaves and flowers of *P. tomentosa* these compounds were sticky but non-toxic to several insects surface of immature fruit were also rich in flavonoids. Oncidinol, a glyceride structurally related the glycerides of the glandular hairs, was isolated from the floral oil of *Ornithophora radicans* and may act as a reward for pollinating bees, also suggesting that the glycerides in the glandular hairs of *P. tomentosa* are not toxic. Therefore, the glandular hairs on the leaves and flowers may only physically obstruct herbivores. In contrast, the secretions of the glandular hairs [8].

BOTANIC DESCRIPTION

Paulownia is a deciduous tree and can reach a height of 20 – 30 m under natural conditions and up to 50 m recorded in China, its origin land [9]. Its diameter can reach 2 m [10]. *Paulownia* plant has a tendency to form many branches if it is grown in open space, whereas in the forest it tends to form a straight trunk. *Paulownia* bark is brown to black, smooth but with visible lenticels in the young tree then gradually are developed vertical cracks together with its growth. Often all the parts except of the old branches are covered with glandular mucigel hair, thick hair and branched hairs or stellate (star shaped). Most of the *Paulownia* species have pseudo-dikomite ramification which are dried after the wither period. The rare leaves create a cylindrical crown or an umbrella shape. Leaves at the matured tree reach the length of 15 – 30 cm and width of 10 – 12 cm, with smooth and weaved sides. The new plants have big leaves and long stem, with trowel sides, placed in front of each other or in spiral shape [11]. *Paulownia* trunk is light, strong, dries quickly, aesthetically pleasant, with light colour that do not change, easily workable and suitable for carvings and isolations [12].

PLANT PARTS AND THEIR MEDICINAL USE

Paulownia is one of the most exploited medicinal plants in terms of the plant parts that have been used in traditional medicine. In traditional Chinese medicine, the bark, fruit, xylem, and leaves of *P. tomentosa* var. *tomentosa* have been applied to treat or prevent a variety of diseases, such as hemorrhoid, carbuncle, inflammatory bronchitis, gonorrhoea, upper respiratory tract infection, parotitis, asthma, traumatic bleeding, erysipelas, bacteriological diarrhea, swelling, bronchopneumonia, enteritis, conjunctivitis, hypertension, and tonsillitis [13,14]. The leaves, wood, and fruits of *P. tomentosa* have been traditionally used for the treatment of tonsillitis, bronchitis, asthmatic attack, and bacterial infections such as enteritis or dysentery. *Paulownia* may also have wound-healing properties, as the leaves have been used for the treatment of frostbite and leg ulcers [15]. Leaves, fruits, and flower are the most important plant parts

employed in folk herbal medicine. Folk remedies in China use mashed *Paulownia* flowers to treat acne vulgaris and the decoction to treat fungal infection on the sole of the foot and the skin between toes [16]. Flowers are also used in treatment of first to second degree empyrosis [17].

P. TOMENTOSA FLAVONOIDS AND THEIR BIOLOGICAL ACTIVITY

Flavonoids represent the most numerous group of secondary metabolites isolated from *P. tomentosa*. Some of the flavonoid compounds found in *P. tomentosa* have been categorized as dietary flavonoids. Consuming this compound is believed to deliver health benefits. The activities of these flavonoids are frequently been reviewed [18]. *P. tomentosa* are rich in flavonoids, with concentrations over 1,000 times greater than those on the surfaces of the young leaves [19].

LEAVES

Flavonoids isolated from *P. tomentosa* leaves have antiradical and cell protective effects [20]. Aqueous extracts of fresh *P. elongata* leaves and silage show *in vitro* antimicrobial activity against *Salmonella enterica*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Paenibacillus alvei*, and *Candida albicans*. The inhibitory effect is more pronounced against gram-negative bacteria [21]. The leaves from *P. tomentosa* (misidentified as *P. coreana*) contain isoatriplicolide tiglate (PCAC) that induced apoptosis *in vitro* studies on cervical and Breast cancer cell lines [22].

FLOWERS

Among various parts of *Paulownia* tree, the flowers seem to be the most used plant part with multiple usages in folk herbal medicine [23]. Extracts of *P. tomentosa* flowers have been of particular research interest due to the presence of flavonoids, specifically Apigenin. Apigenin has been shown to have hypotensive [24], anti-inflammatory [25], antispasmodic [26], antioxidant [27], and vasorelaxant properties [28]. Additionally, Apigenin has been reported to exert its anti-tumorigenic effect *in vitro* as well as *in vivo* not only via the inhibition of tumor cell proliferation, but also via the impairment of the invasive potential of tumor cells [29]. *Paulownia* flower extracts inhibit the growth of certain bacteria the strongest effect was seen on *Staphylococcus aureus*, whereas the effect on *Aspergillus niger*, *Saccharomyces cerevisiae*, and *Penicillium chrysogenum* were not so significant [30]. Methanol extracts from dried flowers of *P. tomentosa* have shown potential antiviral activity against enterovirus 71 (EV 71) and Coxsackie virus A16 (CAV 16), the two main pathogens causing hand, foot and mouth disease (HFMD) [31]. Reported broad spectrum antimicrobial activity of the essential oils derived from *P. tomentosa* flowers the flavonoids extracted from *Paulownia* flowers have been shown to suppress asthmatic tracheal inflammation, while the essential oil from the flowers also alleviated the allergic airway inflammation in mice [32].

FRUITS

It has been noted that certain compounds of *Paulownia* fruits may have high inhibitory activity against *Staphylococcus epidermidis*, a pathogenic gram-positive bacteria [33]. A study reported that the *P. tomentosa* fruits can be a feasible source of natural antioxidant substances [34]. Purification of the methanol extract of *P. tomentosa* fruits yielded potent acetyl cholinesterase (hAChE) and butyrylcholinesterase (BChE) inhibitory flavonoids which have been linked to amelioration of Alzheimer's symptoms in addition to being a rich source of various geranylated flavonoids [35].

WOOD AND BARK

Paulownia wood serves for making paper pulp, musical instruments, furniture, and is also used in construction [36]. Antioxidant activity of extracts from *P. tomentosa* var. *tomentosa* bark has also been demonstrated that might lead to medicinal applications. The latter study showed that isocampneoside II plays a critical role in neuroprotection by acting as a free radical scavenger and antioxidant. [37]

ROOT

Roots have been used to treat chronic retrograde inflammation of the shoulder joint capsule and

Surrounding ligaments, muscles, tendons and bursa mucosa, also known as scapulohumeralperiarthritis in medical terms^[38].

SEED

Paulownia seeds can also be used as a non-traditional material for the production of oil which is rich in bioactive compounds such as sterols and tocopherols for nutritive purposes^[39].

SYRINGIN

Syringin is a Phenylpropanoid glycoside pertaining to eleutheroside derivative. The pharmacological properties of syringin include free radicals scavenging, neuronal cell damage prevention, apoptosis inhibition, antidiabetic effect, anti-inflammatory potential, antinociceptive effect, and anti-allergic actions^[40].

FLAVONOIDS

The four main groups of flavonoids are flavones, flavanones, catechins, and anthocyanins^[41]. The geranylated flavanones from *P. tomentosa* was shown to inhibit levels of nitrite oxide in LPS stimulated rat macrophages^[42].

GLYCOSIDE

Glycosides are a large and very significant class of carbohydrate derivatives that are characterized by replacement of the anomeric hydroxyl group by some other substituent^[43]. With the advancement in chromatographic methods and modern spectroscopic techniques, studies on natural products from the genus *Paulownia* continue to reveal new compounds. The most notable ones are iridoid glycosides, phenylpropanoid, lignin glycosides, flavonoids, sesquiterpene and triterpenes. Many of these compounds have been proven to contain a certain degree of bioactivity^[44]. Phenylethanoid derivatives which are incorporated by ether or ester bond to iridoid glycosides have recently been isolated^[45]. Phenylethanoid glycosides Campneosid I extracted from *P. tomentosa* have a high biological activity. Campneosid I was found to have significant antibacterial activity against several pathogenic strains of *Streptococcus* and *Staphylococcus* including *S. aureus*^[46].

CHEMICAL COMPOUND OF THE PAULOWNIA LEAF^[47]

COMPOUND	IN %
Organic matter	91.4
Proteins	22.6
Nitrogen	2.8-3.0
Kappa	0.4
Potassium	0.6
Calcium	2.1
Iron	0.6
Zinc	0.9
Metabolisation energy	15-18mg /kg

CULTIVATION OF PAULOWNIA TOMENTOSA

Paulownia consumes about 2000 liters of water per tree to reach a production of 4,3 t/ha during the first cut. *Paulownia* has a high adaptability with land climatic conditions. In the Mediterranean climate conditions the *Paulownia* production is negatively affected by the high evapotranspiration but not from the rainfalls and temperatures. This species is very adaptable and widely dispersed. It has a natural distribution from the tropical zones till the ones with moderated climate. And to the zones where the annual rainfalls are between 500 to 2000 mm and a high from the sea level up to 2400 m. The adequate conditions for the *Paulownia* cultivation are attained in a height of 200 – 1300 m above the sea level with an average of the

annual temperature 15 – 23°C and annual rainfalls 1400 – 2800 mm. Paulownia is not affected from pests and diseases; the plant is very flexible and usually not affected by diseases^[48]

Cultivation requirement of Paulownia tomentosa^[49]

Temperature	25-47°C (optimum 27°C)
Water	500-2000 mm (700 mm during vegetative growth or over 150 mm/ month)
Sea level height	2400 m (preferred 750-800m)
Soil temperature	15-16°C
Soil PH	5-8.9
Clay	<25-30%
Total porosity	≥ 50%
Salinity	<1%

MATERIALS AND METHODS

Plant material

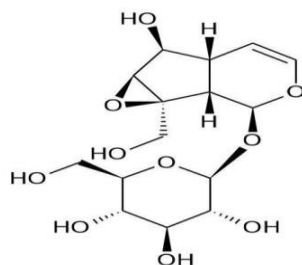
Fresh leaves and fruits of *Paulownia tomentosa*. Were collected from Agricultural Museum, Dokki, Giza, Egypt in July and September respectively. The plant materials were identified by Mrs. Teras Labib, head of the taxonomy at El-Orman Botanical Garden. A voucher specimen (No.06-06-03-16) was kept at the Herbarium of El-Orman Botanical Garden.

Extraction & Isolation

Mature fresh fruits of *P. tomentosa*. Were subjected to complete extraction by cold maceration in absolute methanol. The extract was collected, filtered and evaporated under vacuum. The residue was mixed with distilled water and successively extracted on cold with chloroform, ethyl acetate, methanol respectively. The different extracts were evaporated separately under vacuum till dryness. The methanol extract were subjected to silica gel column chromatography (CC) by (Chloroform – Methanol – Water) to give catalpol (1), Aucubin (2) and paulownioside (3) and new hydrocarbon.

Compound 1

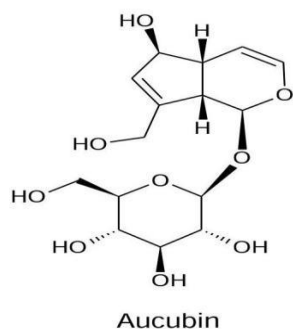
It was isolated as white crystalline powder by preparative TLC from fractions (1-15) CHCl₃-MeOH-H₂O (80:20:2) then purified and crystallized from methanol its m.p.= 203-205, It is highly soluble in water and methanol also soluble in ethanol, acetone, but almost insoluble in lipophilic organic solvents such as chloroform, benzene, and petroleum ether & R_f value is (0.21) in solvent system *n*-butanol-H₂O (9:1).



Catalpol

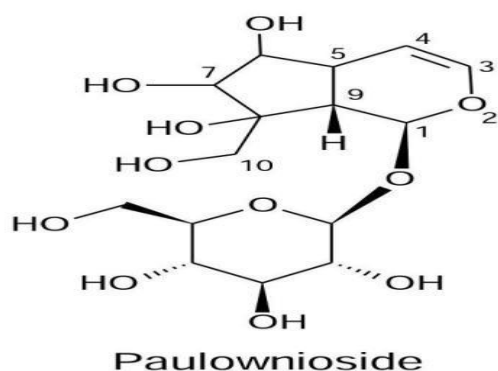
Compound 2

It was isolated as white crystalline powder by preparative TLC from fractions (31-45) CHCl₃-MeOH-H₂O (80:20:2) then purified and crystallized from methanol, its m.p. =181°C, soluble in water, insoluble in chloroform, ether, petroleum ether and R_f values (0.5 & 0.8) in two different solvent systems *n*-butanol-H₂O (9:1) & *n*-butanol-MeOH-H₂O (7:2:1).



Compound 3

It was isolated as white crystalline powder by preparative TLC from fractions (31-45) CHCl₃-MeOH-H₂O (80:20:2) then purified and crystallized from methanol its m.p. is 308°C (decomposed) soluble in water, methanol and insoluble in chloroform & R_f values (0.38 & 0.68) in two different solvent systems *n*-butanol-H₂O (9:1) & *n*-butanol-MeOH H₂O (7:2:1) as shown.



Compound 4

It was isolated as white amorphous powder from the chloroformic fractions (20 fractions). Crystallized by methanol, its m.p. = 201, soluble in chloroform, insoluble in methanol and R_f value (0.75) in solvent system *n*-butanol-H₂O (9:1).

Chemicals and biochemical kits

All chemicals used in the present study were of high analytical grade, products of Sigma (USA), Merck (Germany), BDH (England) was used for the induction of diabetes in rats. Glibenclamide (Daonil) (Sanofi Aventis) was used as standard antidiabetic drug. Kits used for the quantitative determination of different parameters were purchased from Bio-diagnostic (Egypt).

Animals

Male albino rats weighting (120-150g), supplied from the animal house of National Research Centre (Dokki, Giza, Egypt) were used for experimental investigation. The rats were kept in our laboratory under controlled environmental conditions. Anesthetic procedures complied with the legal ethical guidelines approved by the Ethical Committee of the Federal Legislation and National Institutes of Health Guidelines in USA and were approved by the ethical committee of the National Research Centre in Egypt with registration No. 13-015.^[50]

Experimental Design

Wistar Albino rats of either sex, weighing (150-250 g) will be used in this study. Rats will be housed at constant temperature (20 ± 1.8°C) and relative humidity (50 ± 10%) in standard polypropylene cages, six per cage, under a 12-h light/dark cycle, and allowed food and water freely.

Animals are divided into 5 groups each group consist of 6 rats.

Group-1: Control (normal saline for 7 days).

Group-2: Normal saline for 7 days +5 fluorouracil (150mg/kg/ I.P) on 5th day (48 hr. before scarification).

Group-3: Vitamin-E (for 7 days) +5-fluorouracil.

Group-4: Low dose Ethanolic extract of *Paulownia tomentosa*, for 7 days +5 fluorouracil.

Group-5: High dose of Ethanolic extract of *Paulownia tomentosa*, for 7 days +5 fluorouracil.

Forty-eight hours after the 5-fluorouracil administration, blood will be collected from the retro orbital route under ether anesthesia, the rats will be sacrificed and the parameters going to be studied are general appearance, gross morphology, heart weight, histopathology, hematological and biochemical parameters and tissue antioxidant markers.^[51]

Preparation of serum and tissue homogenate

After The experimental regimen, the animals were scarified by cervical dislocation under mild chloroform anaesthesia. Blood was collected on decapitation. The heart were excised immediately and thoroughly washed with ice cold physiological saline. The collected blood was centrifuged at 2500 rpm for 10 min and collects the serum. The serum was used for various biochemical experiments. 1 g of heart was taken and Homogenized with 0.1M cold buffer (pH-7.4) in a potter Homogenizer fitted with Teflon plunger at 600 revolution per for 3min. the homogenate was used for various biochemical assays. The protein Urea, Uric acid and Creatinine were estimated by colorimetric method. Lipid profile in serum Triglycerides, Cholesterol, HDL LDL and Phospholipids were assayed in serum using standard kits. Cardiac markers enzymes such as AST, ALT, LDH, ACP, and ALP. Enzymatic antioxidants such as SOD, Catalase, Lipid peroxidation^[52]

HISTOPATHOLOGICAL STUDIES

Heart was washed in saline and a small portion of it was quickly fixed in 10% formalin. Then the tissue were proceed by standard histopathological technique (i.e.) dehydration through graded isopropyl alcohol, cleaning through xylene and impregnated in paraffin wax for 2 hours. Then wax blocks were made, sections were used for cutting microtone and stained by haematoxylin eosin method and photographed.

STATISTICAL ANALYSIS

The results of cardio protective and antioxidant activities are expressed as mean \pm SD from six animals in each group. The results were statistically analysed using one way ANOVA followed by Tukey-Kramer post test for version 3.00 of graph Pad software, Inc. (San Deigo CA), was used for statistical analysis.

ACUTE TOXICITY STUDIES

The acute toxicity study will be performed by using up and down procedure (OECD-425, guidelines).

ANTIOXIDANT STUDY

Estimation of superoxide dismutase:

Add 2.78 ml sodium carbonate buffer (0.05 mM, pH 10.2), 100 μ L of EDTA (1 mM, 0.0037 g in 10 ml distilled water). Add 20 μ L supernant / sucrose for blank and Incubate at 30°C for 45 min. Thereafter, the reaction will be initiated by adding 100 μ L of adrenaline. The change in the absorbance will be recorded at 480nm for 3 min.

Catalase estimation

Pipette out 100 μ L of supernatant to 1.9 ml phosphate buffer (pH 7), add 1 ml H₂O₂ and measure the changes at the 240 nm for three min.

Lipid peroxidation

1 ml homogenate will be combined with 2 ml (TBA- TCA-HCl). Solution will be heated for 15 min in a boiling water bath, keep it for cooling at room temp, centrifuge at 4000 rpm for 10 min. Take supernatant and measure at 532 nm.

Estimation of Glutathione Peroxidase

To 0.2 ml of tris buffer, 0.2 ml of EDTA, 0.1 ml of sodium azide and 0.5 ml of tissue homogenate will be added. To this mixture, 0.2 ml of glutathione and 0.1 ml of hydrogen peroxide will be added. The contents will be mixed well and incubated at 37°C for 10 minutes along with a tube containing all the reagents except sample. After 10 minutes the reaction will be arrested with the addition of 0.5 ml of 10 % TCA, centrifuged and the supernatant will be assayed for glutathione by Ellman's method. To 2.0 ml of the supernatant, 3.0 ml disodium hydrogen phosphate solution and 1.0 ml of DTNB reagent will be added. The color developed will be read at 412 nm. Standards in the range of 200-1000 μ g will be taken and treated in the similar manner. The activity will be expressed in term of μ g of glutathione consumed/min/mg protein.^[53]

RESULT

Results of cardiac biomarkers were showed a significant increase in serum concentrations of CK-MB in FU-treated group in comparison to control group. This elevation was significantly decreased in LS-treated group, indicating the cardioprotective role of the plant. 5-FU treatment significantly increased the serum TAG and TC levels in FU-treated group in comparison to control group indicating hypertriglyceridemia and hypercholesterolemia. Serum LDL-c and VLDL-c concentrations are significantly increased while the serum HDL-c concentration is significantly decreased in FU-treated group in comparison to control group. Pre-co-post-treatment with LS significantly improved the tested parameters. Results showed a significant increase in this ratio in the FU-treated group which was significantly decreased by LS treatment. It showed a significant increase in inflammatory markers such as myocardial IL-1 β and MPO activity and a significant decrease in GSH concentration in the FU-treated group when compared to control rats. LS treatment reversed the results of these tested parameters. However, cardiac MDA and NO concentrations were non-significantly increased in the FU-treated group in comparison to control group and were non-significantly decreased in the LS-treated group.^[54]

DISCUSSION

The mechanisms of 5-FU-induced cardio toxicity are hemorrhagic infarction, myocardial inflammatory reaction with interstitial fibrosis; arterial endothelial injury followed by thrombosis increased metabolism leading to depletion of ATP, increased levels of superoxide anion and a decreased antioxidant capacity, arterial vasoconstriction and altered plasma levels of substances involved in coagulation and fibrinolysis. Oxidative stress causes cellular damage, coronary artery spasm and the decreased affinity of RBCs to transfer oxygen, resulting in myocardial ischemia, cardiac arrest and sudden death. Myocardium contains high concentrations of diagnostic markers for myocardial infarction and once metabolically damaged, it releases its content into the extracellular fluid. Serum Creatinine kinase (CK) and troponins are some of these markers. The increased activity of serum CK-MB isoenzyme reflects the alterations in the plasma membrane integrity and permeability. In the present study, 5-FU treated rats showed a significant elevation in the activity of serum CK-MB and cTnI level. Which indicated 5-FU induced myocardial necrotic damage and the leakiness of the plasma membrane. LS treatment resulted in lower activity of the CKMB and cTnI level in serum. It was demonstrated that LS could maintain membrane integrity, thereby limiting the leakage of these biomarkers. Hyperlipidemia plays an important role in cardiovascular diseases and the development of atherosclerosis. A significant elevation in the serum TC, TAG, VLDL-c and LDL-c fractions along with a

decrease in HDL-c were observed in 5-FU treated rats compared to control rats. These observed changes concerning lipid profile come in agreement and could be attributed to the enhanced lipid synthesis via cardiac cyclic^[55]

CONCLUSION

Our results confirmed the existence of cardio toxicity due to 5-FU therapy, which was indicated by an elevation of serum cardiac cTnI, CK-MB, altered lipid profile and atherogenic index with enhanced oxidative stress and the release of some inflammatory markers. It can be concluded that LS seed exerts cardioprotective activity that could be partly contributed by its antioxidant and anti-inflammatory activities. So, *Lepidium sativum* can be considered a candidate to protect against cardio toxicity commonly encountered with 5-FU treatment.^[56]

REFERENCES

- [1] World Health Organization. Cardiovascular Disease. Fact sheet N°317. Updated January 2015. Accessed at: <http://www.who.int/mediacentre/factsheets/fs317/en/index.html>.
- [2] Donald M Lloyd-Jones, Peter WF Wilson, Martin G. Larson, Alexa Beiser, Eric P Leip, Ralph B D Agostino, Daniel Levy. Framingham risk score and prediction of lifetime risk for coronary heart disease. *The American Journal of Cardiology*. 2004;94(1):20–4.
- [3] Catherine Kreatsoulas, Sonia S Anand. The impact of social determinants on cardiovascular disease. *Can J Cardiol*. 2010; 26(C):8C–13C.
- [4] Huang, C.; Zhang, X.; Ramil, J. M.; Rikka, S.; Kim, L.; Lee, Y.; Gude, N. A.; Thistlethwaite, P. A.; Sussman, M. A. “Juvenile Exposure to Anthracyclines Impairs Cardiac Progenitor Cell Function and Vascularization Resulting in Greater Susceptibility to Stress-Induced Myocardial Injury in Adult Mice”. *Circulation*. (2010). 121 (5): 675–83.
- [5] Hu Sh.-Y. A Monograph of the Genus *Paulownia*. *Quarterly J. Taiwan Museum*. 1959; 12(1-2):1-54.
- [6] Stringer J. Forest health. Invasive plant hit list *Paulownia*. *Kentucky Woodlands Magazine*. 2009; 4(3):10-11.
- [7] Oprea E, Radulescu V, Chiliment S. The analysis of the volatile and semi volatile compounds of the *Paulownia tomentosa* flowers by gas chromatography coupled with mass spectrometry. *Revista de chimii*. 2004; 55:410-412.
- [8] Geranylated flavanones from the secretion on the surface of the immature fruits of *Paulownia tomentosa*. *Phytochemistry*. 2008; 69(5):1234-41.
- [9] Innes, Robin J. 2009. “*Paulownia tomentosa*. In: Fire Effects Information System” [Online]. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fire Sciences Laboratory (Producer).
- [10] Navroodi Iraj Hassanzad “Comparison of growth and wood production of *Populus deltoides* and *Paulownia fortunei* in Guilan province (Iran)” *Indian Journal of Science and Technology* Vol: 6 Issue: 2 February 2013.

- [11] Zhao-Hua Zhu, Ching-Ju Chao, Xin-Yu Lu, Xiong Yao Gao "Paulownia in China: cultivation and utilization" Published by Asian network for biological sciences and International development research centre 1986.
- [12] Yadav Niraj Kumarmangalam, Brajesh Nanda Vaidya, Kyle Henderson, Jennifer Frost Lee, Whitley Marshay Stewart, Sadanand Arun Dhekney, Nirmal Joshee. "A Review of *Paulownia* Biotechnology: A Short Rotation, Fast Growing Multipurpose Bioenergy Tree" *American Journal of Plant Sciences*, 2013, 4, 2070-2082 Published Online November 2013.
- [13] Jiang JP. Plantation of *Paulownia*. Chinese Forestry Press, Beijing; 2003. 14 Jiangsu New Medical College. Chin. Med. Dict. Shanghai People's Publishing House, Shanghai; 1977.
- [14] Jiang JP. Plantation of *Paulownia*. Chinese Forestry Press, Beijing; 2003. 14 Jiangsu New Medical College. Chin. Med. Dict. Shanghai People's Publishing House, Shanghai; 1977.
- [15] Zhao LZ. Clinical applications of *Paulownia*. *Mag Tradit Chin Ext Med*. 2003; 12(2):48.
- [16] Guo JH, Du G, Shen LL. The research progress of the chemical compositions and Pharmacological actions of *Paulownia* flowers. *Herald Med*. 2011; 30(2):234-235.
- [17] Luo JH, Li K, Entomack B. Research progress on *Paulownia fortunei*. *Guizhou Agri Sci*. 2010; 38(1):200-202.
- [18] Romano B, Pagano E et al (2013) Novel insights into the pharmacology of flavonoids. *Phytother Res* 27(11):1588–1596.
- [19] Asai T, Hara N, Kobayashi S et al (2008) Geranylated flavanones from the secretion on the surface of the immature fruits of *Paulownia tomentosa*. *Phytochemistry* 69:1234–1241.
- [20] Zima A, Hosek J, Tremel J. Antiradical and cytoprotective activities of several Cgeranyl Substituted flavanones from *Paulownia tomentosa* fruit. *Mol*. 2010; 15(9):6035-6049.
- [21] Popova TP, Baykov BD. Antimicrobial activity of aqueous extracts of leaves and silage from *Paulownia elongata*. *Am J Biol Chem Pharm Sci*. 2013; 1(2):8-15.
- [22] Jung S, Moon HI, Ohk J, Lee S, Li C, Kim SK, Lee M-S. Inhibitory effect and mechanism on antiproliferation of Isoatriplicolide Tiglate (PCAC) from *Paulownia coreana*. *Mol*. 2012; 17:5945-5951.
- [23] Kang KH, Huh H, Kim BK, Lee CK. An antiviral furanoquinone from *Paulownia tomentosa* Steud. *Phytother Res*. 1999; 13(7):624-626.
- [24] Gerritsen ME, Carley WW, Ranges GE, Shen CP, Phan SA, Ligon GF, Perry CA. Flavonoids inhibit cytokine-induced endothelial cell adhesion protein gene expression. *Am J Pathol*. 1995; 147(2):278-292.
- [25] Ko HH, Weng JR, Tsao LT, Yen MH, Wang JP, Lin CN. Anti-inflammatory flavonoids and pterocarpanoid from *Crotalaria pallida* and *C. assamica*. *Bioorg Med Chem Lett*. 2004; 14(4):1011-1014.
- [26] Capasso A, Pinto A, Sorrentino R, Capasso F. Inhibitory effects of quercetin and other flavonoids on electrically induced contractions of guinea pig isolated ileum. *J Ethnopharmacol*. 1991; 34(2-3):279-281.

- [27] Cos P, Ying L, Calomme M, Hu JP, Cimanga K, Poel BV, Pieters L, Vlietinck AJ, Berghe DV. Structure-activity relationship and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. *J Nat Prod.* 1998;61(1):71-76.
- [28] Zhang YH, Park YS, Kim TJ, Fang LH, Ahn HY, Hong JT, Kim Y, Lee CK, Yun YP. Endothelium-dependent vasorelaxant and antiproliferative effects of apigenin. *Gen Pharmacol.* 2000;35(6):341-347.
- [29] Czyz J, Madeja Z, Irmer U, Korohoda W, Hülser DF. Flavonoid apigenin inhibits motility and invasiveness of carcinoma cells *in vitro*. *Int J Cancer.* 2005;114(1):12-18.
- [30] Parajuli P, Joshee N, Rimando AM, Mittal S, Yadav AK. *In vitro* anti-tumor mechanisms of various *Scutellaria* extracts and their constituent flavonoids. *Planta Medica.* 2009;75(1):41-48.
- [31] Ji P, Chen C, Hu Y, Zhan Z, Pan W, Li R, Li E, Ge HM, Yang G. Antiviral activity of *Paulownia tomentosa* against enterovirus 71 of hand, foot, and mouth disease. *Biol Pharm Bull.* 2015;38(1):1-6.
- [32] Ibrahim HA, El-Hawary SS, Mohammed MMD, Faraid MA, Nayera AMA, Refaat ES. Chemical composition, antimicrobial activity of the essential oil of the flowers of *Paulownia tomentosa* (Thunb.) Steud. Growing in Egypt. *J Appl Sci Res.* 2013;9(4):3228-3232.
- [33] Chen BH, Li YC. The experimental study of the essential oil of *Paulownia* flower on asthmatic airway inflammation. *Res Tradit Chin Med.* 2007;20(10):16-18.
- [34] Si CS, Wu L, Xu J. The chemical compounds of *Paulownia tomentosa*. *SciTech Cellul.* 2009;17(4):47-5.
- [35] Zima A, Hosek J, Treml J. Antiradical and cytoprotective activities of several Cgeranyl substituted flavanones from *Paulownia tomentosa* fruit. *Mol.* 2010;15(9):6035-6049.
- [36] Cho JK, Ryu YB, Curtis-Long MJ, Ryu HW, Yuk HJ, Kim DW, Kim HJ, Lee WS, Park KH. Cholinesterase inhibitory effects of geranylated flavonoids from *Paulownia tomentosa* fruits. *Bioorg Med Chem.* 2012;20(8):2595-2602.
- [37] Park YM, Jang SK, Kim YS, Kim BK. The constituents of *Paulownia tomentosa* Stem. *Yakhak Hoeji.* 1991;35:301-307.
- [38] Radev R, Sokolova K, Tsokeva Z, Pyrovski L. Antioxidant activity on total extract of *Haberlea rhodopensis*. VI National Congress of Pharm; 2009.
- [39] Zhao LZ. Clinical applications of *Paulownia*. *Mag Tradit Chin Ext Med.* 2003;12(2):48.
- [40] Angelova-Romova Koleva MA, Antova G, Zlatanov M, Stoyanova M, Dobreva K, Denev P, Damianova S, Angelov B, Stoyanova A. Lipid composition of *Paulownia* seeds grown in Bulgaria. *Trakya Uni. J. Sci.* 2011;13(2):101-111.
- [41] Rao USM, Zin T, Abdurrazak M, Ahmad BA. Chemistry and pharmacology of syringin, a novel bioglycoside: A review. *Asian J Pharma Clinical Res.* 2015;8(3),20-25.
- [42] Nešuta O, López-Abán J, Shariati F, Yepes E, Navrátilová A, Šmejkal K, Žemlička M, Muro A. Polyphenols as inhibitors of NO production in LPS-stimulated rat macrophages. *J Nat Pharma.* 2011;2:15-19.

- [43] Jiang JP. Plantation of Paulownia. Chinese Forestry Press, Beijing; 2003.
- [44] Carey FA. Organic chemistry. 7th Ed. McGraw-Hill Companies, Inc., USA; 2007.
- [45] Cao YC, Shao CJ, He Y. The overview of the chemical compositions and biological activities of the genus *Paulownia*. Chin. J. Med. 2008;8(4):308-3010.
- [46] Radev R. Pharmacological effects of phenylethanoid glycosides. J Clin Med. 2010;3(2):20-23
- [47] Yadav Niraj Kumarmangalam, Brajesh Nanda Vaidya, Kyle Henderson, Jennifer Frost Lee, Whitley Marshay Stewart, Sadanand Arun Dhekney, Nirmal Joshee. "A Review of Paulownia Biotechnology: A Short Rotation, Fast Growing Multipurpose Bioenergy Tree" American Journal of Plant Sciences, 2013, 4, 2070-2082 Published Online November 2013.
- [48] Francisco Antonio García-Morote, Francisco Ramón López-Serrano, Eduardo Martínez-García, Manuela Andrés-Abellán, Tarek Dadi, David Candel, Eva Rubio and Manuel Esteban Lucas-Borja. "Stem Biomass Production of *Paulownia elongata* × *P. fortune* under Low Irrigation in a Semi-Arid Environment" *Forests* 2014, 5, 2505-2520.
- [49] Paulownia Bulletin ≠ 3 advices and instructions "Paulownia for biomass production" BIO TREE LTDBulgaria
- [50] Nabaweya A. Ibrahim¹, Seham S. El-Hawary², Sanaa A. Ali^{3*}, Magdy M.D. Mohammed¹, Esraa E. Refaa chemical constituents of *paulownia tomentosa* (thunb) steud. fam. scrophulariaceae and its role against hyperglycemia World Journal of Pharmaceutical Research Volume 4, Issue 8, 2445-2466. ISSN 2277- 7105.
- [51] Kosmas C, Kallistratos M, Kopterides P, Syrios J, Skopelitis H, Mylonakis N, Karabelis A, Tsavaris N. Cardiotoxicity of fluoropyrimidines in different schedules of administration. A prospective study. J Cancer Res Clin Oncol. 2008;134:75-82.
- [52] Ragavan Balliah*, Suganya, Monisha Sudhakar, cardioprotective and antioxidant potential of 50% hydroethanolic crude extract of *theobroma cocoa* in isoproterenol – induced myocardial rats european journal of pharmaceutical and medical research ;2015,2(7), 220-226.
- [53] Rotruck JT, Pope A.L, Ganther HE, Swanson AB, Hafema DG, Hoekstra WG. Science. 1973;179:588-90.
- [54] Zhang R, Brennan ML, Shen Z, MacPherson JC, Schmitt D, Molenda CE, et al. Myeloperoxidase functions as a major enzymatic catalyst for initiation of lipid peroxidation at sites of inflammation. J Biol Chem 2002;277:46116-22.
- [55] Dechant C, Baur M, Böck R, Czejka M, Podcizek-Schweighofer A, Dittrich C, et al. Acute reversible heart failure caused by coronary vasoconstriction due to continuous 5-fluorouracil Combination chemotherapy. Case Rep Oncol 2012;5:296-301.
- [56] Zhao TD, Etherton KR, Martin PJ, Gillies SG. Dietary linolenic acid inhibits proinflammatory cytokine production by peripheral blood mononuclear cells in hypercholesterolemic subjects. Am J Clin Nutr 2007;85:385-91.