ASSESSMENT OF THE ANTIMICROBIAL ACTIVITY OF Chlorella vulgaris EXTRACTS ON BETALACTAMASE PRODUCING BACTERIAL **ISOLATES**

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Abstract: In the present study, the solvent extract of *Chlorella vulgaris* have tested with the agar well diffusion method for their antibacterial against betalactamase producing gram negative and positive bacteria (E.coli, E.faecalis, K.pneumoniae, Salmonella spp, Proteus spp, Shigella spp, P.aeruginosa and S.aureus). The results indicated that both extracts of Chlorella vulgaris were efficient against the betalactamse bacterial isolates. The zone of inhibition ranges from 10 to 24 mm and chloroform extract was high active against to *Proteus* spp. whereas these extracts were ineffective against P. aeruginosa and S. aureus. As a result, it may be concluded that extracts of C. vulgaris, have the capabilities to be explored for antibiotic production. Additional, efforts must be made to identify the compounds straight responsible for antibacterial properties.

Index Terms: Chlorella vulgaris, Betalactamase, Antibacterial activity

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I. INTRODUCTION

Antimicrobial resistance is one of the most important public health issues particularly in developing countries wherever comparatively easily availability and over utilization of medicines have cause to a disproportionately higher incidence of inappropriate use of antibiotics and greater levels of resistance compared to developed countries. In India, the infectious disease trouble is among the highest in the world and a recent report showed the inappropriate and unreasonable use of antimicrobial agents against these diseases, which led to increasing in development of antimicrobial resistance (Gansesh et al., 2013). The crude infectious disease mortality rate in India today is 416.75 per 100,000 persons (Laxminarayan and Ranjit, 2016).

Studies from WHO file have proven very high rates of resistance in micro organism such as Escherichia coli against antibiotics as cephalosporin and fluoroquinolones, Klebsiella pneumoniae in opposition to cephalosporin and carbapenems, Staphylococcus aureus against methicillin. These isolates inflicting infection was untreatable due to the fact its causative agent has been observed be resistant to cephalosporin as well as carbapenems due to extended spectrum of β-lactamases (ESBL) mediated mechanism (Bennett et al., 2010).

The repeated exposure of bacterial isolates to a β -lactams has triggered production of β -lactamases in those bacteria, this example, increasing their pastime even in opposition to the newly advanced β-lactam antibiotics, this circumstance, extending their movement even against the recently created β -lactam antibiotics. In this situation urgently we need a choice solution of eradication of bacterial isolates. The natural antimicrobial substances have been recorded in marine environments. Among the marine organisms, the microalgae (seaweeds) occupy a special site as a source of beneficial substances which are active against to bacteria, fungi, virus and cancer (Maria et al., 2016).

Many bioactive and pharmacologically active components have been isolated from algae. Algae based antimicrobials substance has enormous therapeutic potential as they can serve the motive with lesser side effects that are often associated with synthetic antimicrobials (Justella et al., 2011). Among the various microalgae, Chlorella vulgaris have been explored for their suitability for commercial potential, chlorella species are the major type that has been used successfully to produced high concentration of valuable compounds. (Priya et al., 2012).

Several authors studied the antimicrobial activities of *Chlorella algae* in different parts of our country. According to previous literature, no one study was the antimicrobial activity of Chlorella vulgaris against beta-lactamase producing isolates. Therefore, the present study was to investigate the antimicrobial effect of Chlorella vulgaris against betalactamase producing food pathogens.

II. **METHODS**

a. Food pathogens

For antimicrobial activity of algal extracts against microbial pathogens including Gram positive and Gram negative bacteria were isolates from food samples. Different types of meat samples were collected from the local market. Samples were homogenized using a meat grinder under aseptic conditions and were inoculated on following media such as chromogenic and SS agar. After 24 hrs observed the morphological character. Bacterial identification was conducted by morphological and gram staining method (Collee et al., 1996).

2.2 Isolation of betalactamase producing isolates by Tube Method

Penicillin solution was dispensed in 0.5ml volume in small test tubes. Test bacteria were took with a loop from an overnight culture on solid medium and suspended in the Penicillin solution to give a density of at least 10⁴ CFU/ml. After one hour at room temperature, two drops of the starch indicator was added to the suspension, followed by one drop of Iodine reagent. The positive reaction was indicated by the disappearance of blue color immediately. Persistence of blue color for longer than 10 minutes constituted a negative test (Christensen et al.1990).

The sample microalgae species Chlorella vulgaris was obtained from Royal research centre, Chennai. The algae was cultured on using Bold Basal Medium(BBM). The cultures were grown at 24±1°C in a thermostatically controlled room with cool white in fluorescent lamps at 2-3weeks. After incubation, algal growth was measured by using UV-VIS spectrophotometer at 680nm. Ten ml from cultures were filtered under vacuum using filter membrane (0.45 ym) and washed several times with distilled water. Then, the algae cells were dried at 80°C for 30min and weighed.

2.4 Preparation of algae extracts

Extraction was performed using the solvents of different polarity: chloroform and Methanol. Five grams of the algal dry powder was suspended in the solvents at the ratio of 6:1 (v/w), left at room temperature for 48 h and then, were homogenized for 30 min. Finally, the suspensions were centrifuged at 3000 g for 10 min and the supernatants were filtered and dried. Then dried material was dissolved in DMSO and extracts were kept in dark at 4 °C until use (Ahmad et al., 2017).

2.5 Determination antibacterial activity

This test was carried out according to the method of Jahir et al., 2011. The twenty ml of sterilized Mueller hinton agar (MHA) was poured into each Petri plate (90 mm diameter) and allowed to solidify. The plates were incubated with freshly prepared inoculums which were swabbed over the entire surface of the medium, rotating the plate 60 degrees after each application by using a sterile cotton swab, to ensure the spread of the tested microbes on the surface of the plate completely. Inoculums were 10⁸ CFU/ml of bacteria. One well of 6mm diameter was bored with the medium of each plate with the help of sterile cork-borer. Different concentration of extract was filled in each well with the help of micropipette. Ampicillin (5µg/ml) was used as positive control.

RESULT AND DISCUSSION III.

In the past few years, ESBL-producing E. coli have been increasingly isolated from food-producing animals raising global concerns for veterinary and public health. In the past few years, ESBL-producing E. coli have been increasingly isolated from food-producing animals raising global concerns for veterinary and public health (Seiffert et al., 2013). In the past few years, ESBL-producing E. coli have been increasingly isolated from food-producing animals raising global concerns for veterinary and public health (Seiffert et al., 2013). According to chromogenic and SS agar media, totally 15 isolates from 8 types of bacterial genera were obtained from meat namely as E.coli, E.faecalis, K.pneumoniae, Salmonella spp, Proteus spp, Shigella spp, Pseudomonas aeruginosa and S.aureus.

In the past few years, betalactamase producing isolates have been increasingly isolated from food-producing animals raising global concerns for veterinary and public health (Seiffert et al., 2013). The current study reports on the higher occurrence 67% of betalactamase producing isolates. This report concerns more often the occurrence of the mentioned microorganisms in animals at the farm or the slaughter-house. Previous studies showed a high prevalence of betalactamase producing Enterobacteriaceae in poultry production in India (Charles et al., 2017).

For the last ten years, there have been studies on ESBL in India. In 2015, Kar et al has conducted the first systematic report on multidrug resistance ESBL producing isolates in meat samples in India. In Hyderabad, India, ESBL producing isolates were observed from various meat samples (Rasheed et al. 2014).

ESBL-producing Enterobacteriaceae are highly resistant to a number of drugs, can make a contribution to obtained resistance via horizontal gene transfer throughout a wide range of bacterial species, which are associated with long hospital stay and spend the much money. Recently a number of researches were developed on secondary metabolites on bioassay-guided screening of prokaryotic and eukaryotic microalgae. However, so far these potentials haven't been exploited to any great extent. In the prevailing look at, we targeted to overcome the betalactamase producing Gram-negative and positive isolates causing infection. Therefore, algae of Chlorella vulgaris was collected and subjected to inhibition of betalactamase producing bacterial isolates.

The antimicrobial activity of extracts of C. vulgaris prepared by using methanol, chloroform solvents was recorded in Figure 1 and 2. Data clearly show the great potency of C. vulgaris extracts to inhibit the bacterial growth. Both extracts were able to inhibit the growth of one or more pathogens and the zone of inhibition ranges from 10 to 24 mm whereas methanol extract was ineffective against *P. aeruginosa* and *S. aureus* spp. Currently, none of the isolates were suppressed when using standard antibiotic and DMSO.

In the current study, chloroform extract showed better activity but there is no much more different when using the methanol extract. These results were similar to previous studies of Entesar (2016). This result was showed antimicrobial activity of C. vulgaris against various bacterial isolates. On the other hand, C. vulgaris crude extract showed antibacterial activity against S. aureus and St. pyogenes, K. pneumoniae and E. coli with inhibition zones (19.3 \pm 2.1, 21.1 \pm 0.63, 20.1 \pm 0.58 and 22.4 \pm 0.63 mm respectively). In 2016, Shakeel et al determined the antibacterial activity of lake isolates C. vulgaris, they also report that chloroform extract was activity against wide range of bacterial isolates.

According to literature reports, this is the first study of antibacterial activity of C. vulgaris extracts against betalactamase producing food borne isolates. The present results provide an illustration to the presence of promising the antimicrobial compounds in solvents extract of C. vulgaris under study. Further phytochemical studies are needed to elucidate the components responsible for antimicrobial activity of these extracts against bacteria.

REFERENCE

- [1] Ahmad SI, ed. Reactive oxygen species in biology and human health. UK: Nottingham Trent University; 2016.
- [2] Bennett, J.W., Robertson, J.L., Hospenthal, D.R., Wolf, S.E., Chung, K.K., Mende, K., Murray, C.K., 2010. Impact of extended spectrum beta-lactamase producing Klebsiella pneumoniae infections in severely burned patients. J. Am. Coll. Surg, 211(3): 391-399.
- [3] Charles H. Brower, Siddhartha Mandal Shivdeep Hayer, Mandeep Sran, Asima Zehra, Sunny J. Patel, Ravneet Kaur, Leena Chatterjee, Savita Mishra, B.R. Das, Parminder Singh, Randhir Singh, J.P.S. Gill, and Ramanan Laxminarayan. Prevalence of Extended-Spectrum Beta-Lactamase-Producing 2017. The Resistant Escherichia Coli in Poultry Chickens and Variation According to Farming Practices in Punjab, India. Environ Health Perspect. 125(7): 077015.
- [4] Christensen JJ, Keiding J, Bruun B. 1990. Antimicrobial susceptibility and beta-lactamase characterization of Branhamella catarrhalis isolates from 1983/1984 and 1988. APMIS, 98(11):1039–1044.
- [5] Collee, J.G., Miles, R.S., Watt, B. 1996. Test for the identification of bacteria. In: Collee, J.G., Faser, A.G., Marmion, B.P., Simmons, A. (Eds). Mackie and McCartney practical medical microbiology, 14th edn. Churchill Livingstone, London, Pp. 131 145
- [6] Entesar A. Ahmed. 2010. Antimicrobial Activity of Microalgal Extracts Isolated From Baharia Oasis, Egypt. Global Advanced Research Journal of Microbiology, 5(3):033-041.
- [7] Ganesh Kumar, C. Adithan, B. N. Harish, S. Sujatha, Gautam Roy, and A. Malini. 2013. Antimicrobial resistance in India: A review. J Nat Sci Biol Med, 4(2): 286-291.
- [8] Jahir Alam Khan and Saurabh Tewari. 2011. A Study on Antibacterial Properties of Rosa indica against Various Pathogens. Asian Journal of Plant Science and Research, 1 (1): 22-30.
- [9] Justella Wilfred Prakash, Solomon Jeeva. 2011. Antimicrobial activity of certain fresh water microalgae from Thamirabarani River, Tamil Nadu, South India. Asian Pacific Journal of Tropical Biomedicine, 1(2):170-173
- [10]Kar D, BandyopadhyayS, Bhattacharyya D, SamantaI, Mahanti A, Nanda PK. 2015. Molecular and phylogenetic characterization of multidrug resistant extended spectrum beta-lactamase producing Escherichiacoli isolated from poultry and cattle in Odisha, India. InfectGenet, 29:82-90,
- [11] Maria Jose Pérez, Elena Falque, and Herminia Domínguez. 2016. Antimicrobial Action of Compounds from Marine Seaweed. Mar Drugs, 14(3): 52.-58.
- [12] Priya S. 2012. Analysis of value-added biochemical compounds and antimicrobial activity of green algae Chlorella vulgaris. J Chem Pharm Res, 4(5):2577-2579.
- [13]Ramanan Laxminarayan and Ranjit Roy Chaudhury. 2016. Antibiotic Resistance in India: Drivers and Opportunities for Action. PLoS Med, 13(3): e1001974.
- [14]Rasheed MU, Thajuddin N, Ahamed P, Teklemariam Z, Jamil K. 2014. Antimicrobial drug resistance in strains of Escherichia coli isolated from food sources. Rev Inst Med Trop Sao Paulo, 56(4):341–346.
- [15]Seiffert, S. N., Hilty, M., Perreten, V. & Endimiani, A. 2013. Extended-spectrum cephalosporin resistant Gramnegative organisms in livestock: an emerging problem for human health? Drug Resistance Updates, 16, 22-45.
- [16] Shakeel Ahmed Adhoni , Shivasharana Chandrabanda Thimmappa , Basappa Basawanneppa Kaliwal Shakeel Ahmed Adhoni , Shivasharana Chandrabanda Thimmappa , Basappa Basawanneppa Kaliwal Shakeel Ahmed Adhoni, Shivasharana

Chandrabanda Thimmappa, Basappa Basawanneppa Kaliwal. 2016. Phytochemical analysis and antimicrobial activity of Chorella vulgaris isolated from Unkal Lake. Journal of Coastal Life Medicine, 4(5): 368.

Figure 1 Antibacterial activity of methanol extract of C. vulgaris

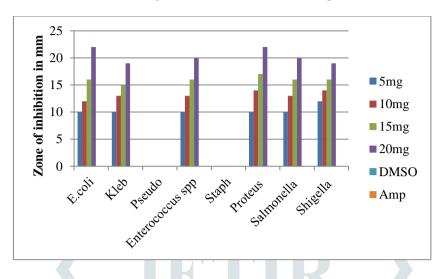


Figure 1 Antibacterial activity of Chloroform extract of C. vulgaris

