

# Review on: QUANTITATIVE ANALYSIS OF POTENT ANTIARRTHMATIC ACTIVE ENTITIES BY RP-HPLC.

<sup>1</sup>Jay Patel , <sup>2</sup>Krunal Detholia , <sup>3</sup>Pinak Patel

<sup>1</sup>Research Scholar , <sup>2</sup>Assistant Professor (Pharmaceutics) , <sup>3</sup>Associate Professor (Pharmaceutical Quality Assurance)

<sup>1</sup>Pharmaceutical Quality Assurance

<sup>1</sup>Smt. S.M. Shah Pharmacy College, Amsaran , Gujarat-387130.

## ABSTRACT

Analytical method development and its validation is a predominant feature in drug discovery process and Reverse Phase-High Performance Liquid Chromatography (RP-HPLC) is the most accepted analytical method utilized for analysis of drug due to its accuracy, selectivity and sensitivity. Development and validation of analytical method are providing accurate and precise data to ensure drug for its quality and safety. Sundry methods of analysis are reported/available for estimation of Anti-arrthmatic agent including RP-HPLC. This review article concisely discusses about analytical methods obtainable for the estimation of Anti-arrthmatic drugs substantially focusing on RP-HPLC.

**KEYWORDS:** - Anti-arrthmatic, Analytical method development, RP-HPLC.

## INTRODUCTION

The technique of RP -HPLC is so called because of its improved performance when compared to other chromatographic techniques. Since high pressure is used when compared to other chromatography, it is so called as high pressure liquid chromatography. In reverse phase technique, stationary phase is a non-polar in nature and polar mobile phase is used. Hence, polar components get eluted first and non-polar compounds are retained for longer time. Due to the polar nature of the most of the drugs and pharmaceuticals, they are eluted faster and not retained for a longer time, which is advantageous. Different columns used are ODS (Octadecyl silane), C18, C8, C4, etc<sup>[1]</sup>. Table 1 represents the classification of Anti-arrthmatic drugs based on their chemical structure<sup>[2]</sup>.

**Table 1**  
Chemical classification of Anti-arrthmatic agents<sup>[2]</sup>.

Class	Action	Drugs
1.	Membrane stabilizing agents(Na <sup>+</sup> channel blockers)	
	A. Moderately decrease dv/dt of 0 phase	Quinidine, Procainamide, Disopyramide
	B. Little decrease in dv/dt of 0 phase	Lidocaine, Mexiletine
	C. Marked decrease in dv/dt of 0 phase	Propafenone, Flecainide
2.	Antiadrenergic agents	Propranolol, Esmolol, Sotalol
3.	Agents widening AP	Amiodarone, Bretylium, Dofetilide , Ibutilidine
4.	Calcium channel blocker	Verapamil, Diltiazem
IN Addition		
5.	For PSTV	Adenosine, Digitalis
6.	For A-V block	Symathomimetics-Isoprenaline Anticholinergics-Atropine
7.	Digitalis in used in AF,AFI, and PSTV to control ventricular rate	

Table 2

Chromatographic condition for Anti-arrhythmic class 1 drugs: membrane stabilizing agent( Na<sup>+</sup> channel blocker ) A:  
Moderately decrease dv/dt of 0 phase

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Quinidine <sup>[3]</sup>	Bulk and tablet dosage form	Phenomenex C-8, (250mm x 4.6mm, 5 µm)	Methanol:sodium dihydrogen ortho phosphate buffer(70:30v/v)	UV Detection At 250nm
Quinidine <sup>[4]</sup>	Bulk and capsule dosage form	ODS C18 (250mm X 4.6 mm i.d., 5 µm particle size)	Phosphate buffer (pH 4.5): Methanol (55: 45 v/v)	UV detection at 234 nm
Quinidine <sup>[5]</sup>	Pharmaceutical dosage form	C-4 column,(250 mm x 4.6mm, 5 µm)	ammonium acetate buffer (0.02 M, pH 3.5) and acetonitrile (30:70 v/v)	UV Detection At 250nm
Procainamide <sup>[6]</sup>	Plasma	10cmx4 mm Novapak reversedphase (5 µm) equipped with a 2 cm guard column for RP column (37 to 53 P>	Water:methanol:acetic acid:triethylamine (74:25:1:0.03 v/v/v/v)	UV detection at 280 nm
Procainamide <sup>[6]</sup>	Plasma and drugs	25 cm x 4.6 mm of ultrasphere ODS (5 µm) with a guard column (3 cm x 4 mm) of pBondapak C18 - Corasil (37-50 µm)	0.05M KH <sub>2</sub> P0 <sub>4</sub> (pH 2.5) - acetonitrile	220 nm to 280 nm
Procainamide <sup>[6]</sup>	Bulk and Pharmaceutical dosage form	12.5 cm x 4.9 mm of Sherisorb s5w	Methanolic 10 mM Ammonium perchlorate and methanolic 0.1 M-sodium hydroxide (pH 6)	UV detection at 254 nm
Procainamide <sup>[6]</sup>	Human serum	15 cm x 3.9 mm of Novapak clg FLm)	Acetic acid;water:acetonitrile (2:50:5v/v/v) containing 3.5% of sodium acetate	UV detection at 254 nm
Procainamide <sup>[6]</sup>	Serum and plasma	10cmx5 mm of Novapak cyano HP.	5mM acetate buffer (pH 6) containing 0.05% triethylamine and 10% acetonitrile	UV detection at 280nm
Procainamide <sup>[6]</sup>	Bulk and Pharmaceutical dosage form	10 cm x 4.6 mm Brownlee RP-8 (5 µm)	100% methanol:aqueous phosphate buffer (pH 3) (1:1 v/v)	UV detection at 460nm
Procainamide <sup>[6]</sup>	Bulk and Pharmaceutical dosage form	5cmx4 mm of Nucleosil 5SA (5 µm)	0.1 Mphosphate buffer (pH 3) containing 30% of acetonitrile.	UV detection at 280 nm
Procainamide <sup>[6]</sup>	Serum and urine	15 cm x 4.5 mL IBM phenyl(5 elm.).	75 mM Acetate buffer(pH4.3) acetonitrile (20:3 v/v)	UV detection at 270nm
Procainamide <sup>[6]</sup>	Human blood plasma	15 cm x 4.8 mm stainless steel, packed with Nucleosil CIS (5 µm.1.	Methanol : acetic acid : triethylamine : water (40:2:1:157 v/v/v/v)	UV detection at 335 nm(extraction at 290 nm)
Procainamide <sup>[6]</sup>	Serum	25 cm x 4.6 mm of CNtype (5µm) material operated at 40° c	Acetonitrile:methanol : buffer (60: 7 : 3 v/v/v), buffer is 10 mM phosphate - 0.5 mMtriethylamine, pH was adjusted to 7.1	UV detection at 205 nm
Procainamide <sup>[6]</sup>	Human serum	pBondapak ODS	Aqueous 35% methanol : 0.06M acetate buffer (pH 4.5).	UV detection at 220 nm
Procainamide <sup>[6]</sup>	Serum	CS Reversed phase column.	80% Phosphate buffer (25 mmol/L, pH3.4)20% organic (acetonitrile: methanol, 2:3 v/v) and then changed to 30% phosphate 70% organic at 20 min after injection.	UV detection at 212 nm
Procainamide <sup>[6]</sup>	Biological fluid	30 cm x 4 mm i.d.	Acetonitrile phosphate buffer	UV detection at

		of alkylphenyl column	pH 6.6 column (60:40 v/v)	280 nm
Procainamide <sup>[6]</sup>	Serum	15 cm x 4.6 mm i.d. of S ynchropak SCD-100 (5 Pm)	25 mM- Ammonium sulfate (pH 7.3) containing 24% of acetonitrile	UV detection at 230 nm
Procainamide <sup>[6]</sup>	Serum	0.5 m x 2.6 mm i.d. of ODS-SIL-X	Acetonitrile: phosphate buffer (1 0:90 v/v).	UV detection at 205 nm
Disopyramide <sup>[7]</sup>	Human plasma	5- $\mu$ m Supelcosil CN 25 cm X 4.6 mm column	Gradient: Mobile phase I, acetonitrile 25 mM :sodium phosphate, pH 6.8 (50:50v/v) Mobile phase II, acetonitrile-25 mM :sodium phosphate, pH 6.8 (38:62v/v)	UV Detection At: 215nm
Disopyramide <sup>[8]</sup>	Pharmaceutical dosage form	10- $\mu$ m Lichrosorb RP-18 250 mm X 4 mm column	Acetonitrile : methanol:phosphate buffer (PH 3.2)(6:1:3,v/v/v)	UV Detection At: 269nm

**Table 3**

Chromatographic condition for Anti-arrhythmic class 1 drugs: membrane stabilizing agent( Na<sup>+</sup> channel blocker ) B: Little decrease in dv/dt of 0 phase

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Lidocaine <sup>[9]</sup>	Topical formulation	HiQSiHSC18column (250 mm $\times$ 4.6 mm, 5 $\mu$ m).	acetonitrile: 0.01 M diethylamine solution (pH adjusts to 6.8 with orthophosphoric acid) (60:40 v/v)	UV Detection At: 225nm
Lidocaine <sup>[10]</sup>	Human serum	Alltima C18-column ( 150 $\times$ 4.6 mm) i.d., 3.5 $\mu$ m	acetonitrile and phosphate solution (25 mM KH <sub>2</sub> PO <sub>4</sub> -3 mM sulfuric acid-3.6 mM triethylamine) in a ratio of 12:88 (v/v)	UV Detection At: 210nm
Lidocaine <sup>[11]</sup>	Pharmaceutical dosage form	ODS 4.6 mm x15 cm, 5 $\mu$ m	Water and acetonitrile(72:28v/v) pH adjusted to 2.0 with 85% orthophosphoric acid	UV Detection At: 208nm
Lidocaine <sup>[12]</sup>	Mouthwash formulauion	Waters Symmetry C18 HPLC column (150 mm $\times$ 4.6 mm, 5 $\mu$ m)	methanol - 0.1M Sodium dihydrogen phoshpate with a pH that was previously adjusted to 4.5 with dilute phosphoric acid with gradient elution.	UV Detection At: 230nm
Lidocaine <sup>[13]</sup>	Pharmaceutical dosage form	C18 Hypersil BDS column	Briton-Robinson buffer, pH 7: methanol:acetonitrile (40: 45: 15 v/v/v)	UV Detection At: 225nm
Lidocaine <sup>[14]</sup>	Pharmaceutical gel	5 mm LichroCART RP-18 column (125 mm x 4 mm i.d.)	acetonitrile: 0.05 M sodium phosphate buffer, pH 6.0 (35:65 v/v) and 0.05% of diethylamine	UV Detection At: 210nm
Lidocaine <sup>[15]</sup>	Tablet and Injection	Spherisorb ODS (5 microm) (4.6 mm x 250 mm(2))	5.5% triethylamine in acetonitrile/water(70/30 v/v)	UV Detection At: 254nm
Mexiletine <sup>[16]</sup>	Capsule	5 $\mu$ m LiChrospher 60, RP-Select B column (250mm $\times$ 4 mm ID)	1.28 g dodecyl sodium sulfate and 4.00 g of anhydrous sodium dihydrogen phosphate in 600 mL of water (pH 4.5) acetonitrile (60 : 42, v/v).	UV detection at 262 nm

**Table 4**

Chromatographic condition for Anti-arrhythmic class 1 drugs: membrane stabilizing agent( Na<sup>+</sup> channel blocker ) C:  
Marked decrease in dv/dt of 0 phase

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Propafenone <sup>[17]</sup>	Blood sample	Spherisorb SCN column(4.6 mm x 120mm)3 μm	Acetonitrile:buffer(7.7 gm ammonium acetate 900 ml water then it is acidified with acetic acid pH 6.0 that it will diluted till 1000 ml.(50/50 v/v)	UV detection at 248 nm
Propafenone <sup>[18]</sup>	Tablet	Inerstil ODS-3 Vs column	Gradient; Solvent A: Acetonitrile(75) Solvent B : methanol:water(90:10)	UV detection at 245 nm
Flecainide <sup>[19]</sup>	Bulk powder and dosage form.	RP-C18 column	phosphate buffer pH 3.3:acetonitrile:triethylamine (53:47:0.03v/v/v)	UV detection at 292 nm
Flecainide <sup>[20]</sup>	Biological specimen	HYPERSIL BDS Phenyl column (53mm X 7.0 mm),3 μm	Gradient: Solvent A: Water:Methanol:1.5 M ammonium acetate (87:10:3 V/V/V) Solvent B: Water:Methanol:1.5 M ammonium acetate (17:80:3 V/V/V)	UV detection at 230nm

**Table 5**

Chromatographic condition for Anti-arrhythmic class 2 drugs: Antiadrenergic agents

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Propranolol <sup>[21]</sup>	Tablet	XTerra RP18(150mm X 4.6 mm,5 μm)	Pyrrolidine(pH 11.5):acetonitrile(50:50 v/v)	UV detection at 214 nm
Propranolol <sup>[22]</sup>	Bulk and pharmaceutical dosage form	Thermo-hypersil c8 column (250mm x 4.6 mm,5 μm)	Acetonitrile: 0.01 M ammonium phosphate buffer (pH:3 adjusted by orthophosphoric acid ) (50:50 v/v)	UV detection at 238nm
Propranolol <sup>[23]</sup>	Pharmaceutical dosage form	C18 column(Inerstil c18,250mm x 4.6 mm, 5 μm)	Methanol:acetonitrile:water(50:30:20 v/v/v)	UV detection at 232 nm
Propranolol <sup>[24]</sup>	Tablet dosage form	Agilent C18 (250 4.6 mm, 5 μm)	0.02 M sodium Phosphate buffer: acetonitrile(65:35 v/v)	UV detection at 240 nm
Propranolol <sup>[25]</sup>	Bulk and pharmaceutical dosage form	Agilent xdb C18(150 mm X 4.6 mm, 5 μm)	Methanol:acetonitrile:potassium dihydrogen phosphate(adjusted pH 3.0 using orthophosphoric acid),(20:20:60,v/v/v)	UV detection at 264 nm
Esmolol <sup>[26]</sup>	Bulk and pharmaceutical dosage form	C 18 column(250mm x 4.6 mm, 5 μm)	Acetonitrile:0.05 M sodium acetate buffer : glacial acetic acid(35:65:3 v/v/v)	UV detection at 275 nm
Sotalol <sup>[27]</sup>	Tablet	C 18 column(100mm x 4.6 mm, 5 μm)	Methanol:Acetonitrile: 20 mM ammonium acetate (pH-6)(15:10:75 v/v/v)	UV detection at 229 nm
Sotalol <sup>[28]</sup>	Oral liquid	Ascentic express c18(100 mm x4.6 mm, 2.7 μm)	Gradient phase :sodium dihydrogen phosphate dehydrate: acetonitrile	UV detection at 237 nm

**Table 6**  
Chromatographic condition for Anti-arrhythmic class 3 drugs: Agents widening AP

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Amiodarone <sup>[29]</sup>	Bulk and pharmaceutical dosage form	Hypersil BDS C 18 column(100mm x 4.6 mm, 5 µm)	0.5% v/v TEA buffer pH 6.5 and acetonitrile (25:75 v/v)	UV detection at 240 nm
Amiodarone <sup>[30]</sup>	Bulk and pharmaceutical dosage form	Hi Q SIL C 18(4.6mm x 520 mm, 5 µm)	Acetonitrile:0.5% formic acid(80:20 v/v)	UV detection at 242 nm
Amiodarone <sup>[31]</sup>	Tablet	C 18 column(150mm x 4.6 mm, 5 µm)	Buffer solution pH 5.0(prepared by 3.0ml acetic acid in 1000 ml water and adjusted pH 5.0 with dilute ammonia solution):methanol:acetonitrile(30:30:40 v/v/v)	UV detection at 240nm
Bretylum <sup>[32]</sup>	Pharmaceutical dosage forms	Inersil ods 2(150mm x 4.6 mm,5 µm)	Gradient: Eluent A: 1 ml TFA in 1000 ml water Eluent B: 1 ml TFA in 1000ml acetonitrile	UV detection at 215 nm
Bretylum <sup>[33]</sup>	Pharmaceutical dosage forms	XBridge C-18 Column(150mm x 4.6mm,5 µm)	3 mol KOH buffer solution pH 10.50:methanol(60:40v/v)	UV detection at 215 nm
Dofetilide <sup>[34]</sup>	Plasma	Ultrasphere ods c18 (250x4.6mm,5 µm)	Gradient: Buffer A:methanol/ammonium phosphate, 1-heptane-sulfonate:acetonitrile(19:70:11 v/v/v) containing 0.08% triethylamine and adjusted pH7.1 with phosphoric acid Buffer B:methanol/ammonium phosphate, 1-heptane-sulfonate:acetonitrile(38:4:20 v/v/v) containing 0.08% triethylamine and adjusted pH7.1 phosphoric acid	UV detection at 238nm
Dofetilide <sup>[35]</sup>	Bulk and pharmaceutical dosage form	Phenomenex c18 (150x4.6 mm,1.8 µm)	Potassium dihydrogen phosphate buffer pH 6.8:acetonitrile:methanol(50:30:20 v/v/v)	UV detection at 231 nm
Ibutilide <sup>[36]</sup>	Pharmaceutical manufacturing environment	Inersil c8 250x 4.6 mm,5 µm	Gradient phase Solvent A:0.01 M potassium dihydrogen phosphate Solvent B:acetonitrile	UV detection at 227 nm
Ibutilide <sup>[37]</sup>	Bulk and pharmaceutical dosage form	Hypersil c18 (250x4.6 mm,5 µm)	Acetonitrile:0.1 M ammonium dihydrogen phosphate buffer pH-6.0 (25:75 v/v)	UV detection at 250 nm
Ibutilide <sup>[38]</sup>	Pharmaceutical dosage form	Hypersil c18 (250x4.6 mm,5 µm)	Methanol:water(55:45 v/v)	UV detection at 228 nm
Ibutilide <sup>[39]</sup>	Injection	Vp-ODS column(250mm x 4.6 mm,5 µm)	0.06 M 1-1 potassium dihydrogen phosphate pH 5.8:methanol:triethylamine(45:55:0.01v/v/v)	UV detection at 228 nm

**Table 7**  
Chromatographic condition for Anti-arrhythmic class 4 drugs : Calcium channel blocker

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Verapamil <sup>[38]</sup>	Pharmaceutical dosage form	Hypersil c18 (250x4.6 mm,5 µm)	Methanol:water(55:45 v/v)	UV detection at 228 nm
Verapamil <sup>[40]</sup>	Human plasma	Lichrosphere 60 RP-select Bcolumn(250 mm x 4mm,5 µm)	Acetonitrile:0.025 mol/l potassium dihydrogen phosphate (40:60 v/v)	UV detection at 200 nm
Verapamil <sup>[41]</sup>	Human plasma	Cyanopropylsilane column	Acetonitrile:0.02 M acetate buffer pH 7.0(65:35 v/v)	UV detection at 228 nm
Verapamil <sup>[42]</sup>	Capsule	Lichrosorb RP18 column	Acetonitrile:methanol:phosphate buffer pH 2.7(40:40:20 v/v/v)	UV detection at 220 nm
Verapamil <sup>[43]</sup>	Pharmaceutical dosage form	C18 reverse phase column	Methanol:water:acetic acid:triethylamine(55:44:1:0.1v/v/v/v)	UV detection at 280nm
Verapamil <sup>[44]</sup>	Plasma	C18 reverse phase	Gradient:	Fluorescence

		column	Mobile phase A: 50 mM ammonium phosphate, pH 4.5) Mobile phase B: 50mM ammonium phosphate:acetonitrile(70:30 v/v)	detector excitation 313 nm and emission 280nm
Verapamil <sup>[45]</sup>	Residue	Hypersil ODS (125 x 4.0 mm, 5 µm)	Methanol:water:triethylamine(70:30:0.2 v/v/v)	UV detection at 278 nm
Diltiazem <sup>[46]</sup>	Tablet	Hypersil bds c18(150 mm x 4.6 mm, 5.0 µm)	Gradient : Mobile phase A:0.2%triethylamine pH adjusted to 4.5 with orthophosphoric acid Mobile phase B:acetonitrile	UV detection at 240 nm
Diltiazem <sup>[47]</sup>	Raw and pharmaceutical formulation	Hypersil ODS C 18(150mm x4.6 mm, 5 µm)	Methanol:water(80:20 v/v)	UV detection at 236 nm
Diltiazem <sup>[48]</sup>	Plasma	Spherisorb c18	Methanol:water containing 2.8 mM triethylamine(80:20 v/v)	UV detection at 239 nm
Diltiazem <sup>[49]</sup>	Drug metal interaction	Shim pack CLC – ODS(6.0mm x150 mm)	Acetonitrile:water(40:60 v/v), pH adjusted to 2.8 with phosphoric acid	UV detection at 235 nm
Diltiazem <sup>[50]</sup>	Plasma	Zorbax SB c18(250mm x4.6 mm, 5 µm)	0.2 M ammonium dihydrogen phosphate :acetonitrile:isopropyl alcohol:triethylamine(55:43:1.7:0.3 v/v/v/v)	UV detection at 240 nm
Diltiazem <sup>[51]</sup>	Raw and pharmaceutical dosage form	Rp c-18(250mm x 4.6 mm, 5 µm)	0.01 M ammonium acetate in water:methanol:acetonitrile(700:240:60 v/v/v)	UV detection at 295 nm
Diltiazem <sup>[52]</sup>	Pharmaceutical and human serum	Hiber rp c-18(250mm x 4.6 , 5 µm)	Acetonitrile:water(85:15 v/v)	UV detection at 230 nm
Diltiazem <sup>[53]</sup>	Raw material	Microbonapack c 18 column(250mm x 4.6 mm, 5 µm)	0.1 M sodium acetate buffer pH 6.3:acetonitrile(65:35 v/v)	UV detection at 240 nm
Diltiazem <sup>[54]</sup>	Plasma	Agilent TC c18 column(250mm x 4.6 mm, 5 µm)	Methanol:water(90:10 v/v)	UV detection at 237 nm

Table 8

Chromatographic condition for Anti-arrhythmic class 5 drugs : For PSTV

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Adenosine <sup>[55]</sup>	Kinase	C 18 column	Water:acetonitrile(93:7 v/v)	UV detection at 260 nm
Adenosine <sup>[56]</sup>	Intracellular	Luna c18(250mm x 4.6 mm, 5 µm)	Acetonitrile:50 mM monobasic potassium phosphate pH 4.6(0.5:99.5 v/v)	UV detection at 254 nm
Adenosine <sup>[57]</sup>	Human synovial fluid	ODS (250mm X 4.6 mm, 5 µm)	Phosphate buffer and acetonitrile(90:10 v/v)	UV detection at 260 nm
Digitalis <sup>[58]</sup>	Leaves	Octylsilyl bonded silica column	Acetonitrile:methanol:water(10:15:18 v/v/v)	UV detection at 220 nm
Digitalis <sup>[59]</sup>	Tablet (digoxin)	Symmetry c 18 (75mm x 4.6 mm, 3.5 µm)	Water:acetonitrile(72:28 v/v)	UV detection at 220nm

Table 9

Chromatographic condition for Anti-arrhythmic class 6 drugs:For A-V block A:Symathomimetics

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Isoprenaline <sup>[60]</sup>	Injection	Eclipse XDB C-18(250mm x 4.6 mm, 5 µm)	0.14% sodium heptan esulfonate with adjusted pH 3.5 with orthophosphoric acid:methanol (65:35v/v)	UV detection at 280 nm

**Table 10**

Chromatographic condition for Anti-arrhythmic class 6 drugs: For A-V block B:Anticholinergics

Name of Drug	Sample Matrix	Chromatographic Condition	Mobile Phase	Detection
Atropine <sup>[61]</sup>	Ophthalmic formulation	ODS –BP hyperchrome c 18 column(250mm x 4.6 mm,5 µm)	0.05 M potassium dihydrogen phosphate buffer pH 4.0:acetonitrile(60:40 v/v)	UV detection at 266 nm
Atropine <sup>[62]</sup>	Raw Material	C 18 column	Methanol :5 mM potassium dihydrogen phosphate buffer(50:50v/v)	UV detection at 264 nm
Atropine <sup>[63]</sup>	Eye drops	c-8 column	Acetonitrile:dilute acetic acid (80:20 v/v)	UV detection at 260nm

**Conclusion**

This review briefs about the analytical methods development and validation of anti-arrhythmic drugs in various pharmaceutical formulations, bulk and biological samples alone or in combination with other drugs by using RP-HPLC.

Purpose of using RP-HPLC method is for estimation of anti-arrhythmic drugs and validated that method as per ICH guideline. Beneficial properties of RP-HPLC method for estimation of anti-arrhythmic drugs is its high sensitivity, accuracy and reproducibility.

Also the article will be very beneficial for many researchers working in the area of estimation of anti-arrhythmic drugs as they can refer most of the RP-HPLC methods of estimation of anti-arrhythmic drugs by referring this single article.

**Acknowledgement**

The purpose of this review would not have been possible without help and guidance of Dr.Pinak Patel. I am very much thankful to my project guide Dr. Pinak Patel (HOD, Department of Pharmaceutical Quality Assurance) Smt. S.M. Shah pharmacy college, Mahemdavad as he is ever ready to help me in achieving my task, Mr. Krunal Detholia (Department of Phareaceutics), Smt. S.M. Shah pharmacy college, Mahemdavad) for providing resource, guidance and support.

**References**

- Shankar, R. 2001.International methods of chemical analysis: 18.2-18.4.
- Tripathi, KD.2013. Essential of medical pharmacology,7<sup>TH</sup> Edition:526-538.
- Poornima, K. and Madhusudan ,Y. et al. 2017. Development and validation of analytical method for simultaneous estimation of Dextromethorphan and Quinidine by RP-HPLC and UV-Spectrometry. International Journal of Pharmaceutical Sciences and Research, 8(3): 1301-1313.
- Chaudhary , Z. and Derasari, J.2014.RP-HPLC method development and validation for simultaneous estimation of Dextromethorphan hydrochloride and Quinidine Sulphate . International Bulletin of Drug Research, 4(7): 66-83.
- Venkatesh ,G. and Ramanathan, S . et al.2007. Development and validation of RP-HPLC-UV method for simultaneous determination of Buparvaquone, Atenolol, Propranolol, Quinidine and Verapamil: A tool for the standardization of rat in situ intestinal permeability studies . Journal of Pharmaceutical and Biomedical Analysis,43(4):1546-1551.
- Brittan, H.2001. Analytical Profiles of Drug Substances and Excipients.28:295-304.
- Angelo, H. and Bonde, J. et al.1986.A HPLC method for the simultaneous determination of Disopyramide, Lidocaine and their monodealkylated metabolites. Scand J Clin Lab Invest , 46: 623-627.
- Witek , A. and Zawisza, P.et al.1994.Determination of Disopyramide in plasma by high performance liquid chromatography. Journal of Pharmaceutical and Biomedical Analysis.12: 425-427.
- Goweskar, N. and Wadher, S.2017.Development and validation of HPLC method for simultaneous determination of Lidocaine and Prilocaine in topical formulation. Asian J Pharm Clin Res,10(10): 179-182.
- Nebaihi, H. and Primrose, M. et al.2017. A High-Performance Liquid Chromatography Assay Method for the Determination of Lidocaine in Human Serum. Pharmaceutics , 9(4):52.
- Malenovic, A. and Markovik, S.2005. Development and validation of RP-HPLC method for Cetrimonium bromide and Lidocaine determination. Journal of Farmaco Volime, 60(2):157-161.
- Pendela, A. et al. 2011. Simultaneous determination of Lidocaine hydrochloride, Hydrocortisone and Nystatin in a pharmaceutical preparation by RP-LC. Journal of Pharmaceutical and Biomedical Analysis ,56(3):641-644.
- Mohammad, M. et al .2009.LC determination of Lidocaine and Prilocaine containing potential risky impurities and application pharmaceuticals.Chromatographia ; 70 (3-4) : 563-568.
- Junior, E. and Bentley ,M. et al.2002.HPLC assay of Lidocaine in vitro dissolution test of the Poloxamer 407 gels. Revista Brasileira De Ciencias Farmaceuticas ,38 (1): 107-111.
- Liawruangrath, S.2001.Simultaneous determination of Tolperisone and Lidocaine by high performance liquid chromatography. Journal of Pharmaceutical and Biomedical analysis,26(5-6):865-872.

16. Zdravljje, A .2010. Development and application of a validated HPLC method for the analysis of dissolution samples of Mexiletine Hydrochloride capsules. *Journal of the Serbian Chemical Society*, 75 (7): 975–985 .
17. Alihodjaeva, M. and Atahanov , A . et al.2015.Development of methodology for the isolation and determination of Propafenone in Blood sample via HPLC. *Journal of US-China Medical Science*,12(2015):163-171.
18. Rao, N. and Reddy,S. et al .2018. New method development and validation for the determination of Propafenone HCL in pure drug and its tablet dosage form by RP-HPLC. *International Journal of Research in Pharmaceutical Sciences*,9(3):656-662.
19. El-Ragehy, NA. and Hassan, NY. et al. 2016. Stability-indicating chromatographic methods for determination of Flecainide acetate in the presence of its degradation products; isolation and identification of two of its impurities.*Biomed Chromatogr*,30(10):1541-1548.
20. Benijit, T. and Borrey, D. et al.2003.Analysis of Flecainamide and two metabolites in biological specimens by HPLC. *Journal of Analytical Toxicology*,27:47-52.
21. Shabir, G .2011. Development and validation of RP-HPLC method for the determination of Methamphetamine and Propranolol in tablet dosage form.*Indian J Pharm Sci*,73(4):430-435.
22. Lahari,K. and Kumari, S. et al. 2012. Development and validation of RP-HPLC method for simultaneous estimation of Clonazepam and Propranolol Hydrochloride in bulk and pharmaceutical dosage forms. *Inventi Rapid :Pharm Analysis & Quality Assurance* ,2012(4):1-4.
23. Bendapudi, P. and Rao , V. et al. 2012. Development of RP-HPLC method for the simultaneous estimation of Propranolol hydrochlorideand hydrochlorothiazide in combined dosage form. *International Journal of Biological & Pharmaceutical Research* ,3(7):899-903.
24. Umamaheshwari , D. and Jayakar, B .2015.RP-HPLC method for the simultaneous determination of Etizolam and Propranolol in pure and its tablet dosage form.*International Journal of Pharmaceutical , Chemical and Biological Science*.5(1):213-216.
25. Shalini,Y. and Sai, A. et al.2012.Method development and validation for simultaneous estimation of Propranolol hydrochlorothiazide and Flunarizine dihydrochloride in bulk and pharmaceutical dosae form.*International Research Journal of Pharmaceutical and Applied Science*,2(5):143-148.
26. Somasekhar V. and , Gowrisankar, D.2010. Development and validation of rapid RP-HPLC method for the estimation of Esmolol hydrochloride in bulk and pharmaceutical dosage forms.*Journal of Chemistry*,7(3):807-812.
27. Abirami,G. and Kumar, A. et al.2013.Analytical method development and validation of estimation method for Sotalol hydrochloride tablet by using RP-HPLC.*Indian Journal of Research in Pharmacy and Biotechnology*,1(5):736-740.
28. Ludmila, M. and Oxana, Z.et al. 2015.Development of gradient HPLC method for simultaneous determination of Sotalol and Sorbate in oral liquid preparation using solid core stationary phase.*Journal of Analytical Method in Chemistry*,2015:1-6.
29. Vemugunta, R. and Baratam, A. et al. 2015. Development and validation of stability indicating RP-HPLC method for the estimation of Amiodarone in bulk and pharmaceutical dosage forms. *PHARMANEST-an International Journal of Advance in Pharmaceutical Sciences*,6(2):2743-2749.
30. Gandhi , S. and Alli, P. et al. 2018.Development and validation of stability indicating HPLC method for estimation of Amiodarone hydrochloride.*European Journal of Pharmaceutical and Medical Research*,5(5):461-469.
31. Fuad, A.2010. Validation of an HPLC-UV methodfor the determination of Amiodarone impurities in tablet formulation. *Pharmaceutical Analytical Acta*,1(1);1-4.
32. Westlake, J.1991. The analysis of basic drugs by HPLC. 127-141.
33. Wiczling,P. and Nasal,P. et al. 2012.A New PH/ organic modifier gradient RP-HPLC method for convenient determination of lipophilicity and acidity of drugs as applied to established Imidazoline agents.*European Journal of Pharmaceutical Sciences*,47(1):1-5.
34. Nisrin, K. And Syvie, P.et al. 2003.Simultaneous determination of Dofetilide and Amlodipine in plasma by HPLC.*Journal of Chromatography Separation Techniques*,4(6):1-6.
35. Adepu , R. and Kumar, A .2018.Method development and method validation for the estimation of Dofetilide in bulk and pharmaceutical dosage preparation by RP-HPLC. *Indo American Journal of Pharmaceutical Research*,8(12):1336-1343.
36. Satya, B. and Anjaneyulu, Y. et al.2009. High performance liquid chromatography method for determination of trace amount of Ibutilide Fumarate in pharmaceutical manufacturing environment. *Journal of Chemistry*,6(1):53-60.
37. Rao, V. and Handrasekhar, K. et al. 2012.Estimation of Ibutilidine Fumarate in bulk and pharmaceutical formulation by simple and validated RP-HPLC. *International Journal of Pharmacy*,2(1):156-158.
38. Sangeetha, R.et al. 2017.Drug-Drug interaction studies of Ibutilidine Fumarate with Verapamil and stability indicating method by chromatographic technique. *International Journal for Research in Applied Science & Engineering Technology*,5(5):2170-2177.
39. Hong, L. and Wang, L. et al. 2009.HPLC determination of content and related substances of Ibutilide Fumarate injection. *Chinese Journal of Pharmaceutical Analysis*,2009(8):1-4.
40. Ivanova, V.2008.HPLC method for determination of Verapamil in human plasma after solid phase extraction.*Journal of Biochemical and Biophysical Method*,70(6):1297-1303.



41. Scott, J. and Shoukry, K.2006.An HPLC method for the determination of Verapamil and Norverapamil in human plasma. *Journal of Liquid Chromatography*,10(6):1187-1201.
42. Gumieniczek, A. and Hopkala, H.2007.Development and validation of liquid chromatographic method for the determination of Trandolapril and Verapamil in capsule. *Journal of Liquid Chromatography & Related Technologies*,24(3):393-400.
43. Dimitri ,C. and Tsilifonis. et al.1958(2006).High performance liquid chromatographic assay of Verapamil Hydrochloride in dosage formsk. *Journal of Liquid Chromatography*,8(3):499-511.
44. Jhee , O.and, Hong, J. et al.2005.Direct determination of Verapamil in rat plasma by coupled column microbore HPLC method. *Journal of Pharmaceutical and Biomedical Analysis*,37(2005):405-410.
45. Dragan, M. and Snezana ,P.et al. 2013.Development and validation of HPLC method for the determination of Verapamil residues in supports of cleaning procedure. *Journal of Analytical Chemistry*,68(6):545-551.
46. Chatpalliwar, V. and Upmanyu, N.2012.Validated gradient stability indicating hplc method for determining Diltiazem hydrochloride and related substance in bulk drug and novel tablet formulation. *Journal of Pharmaceutical Analysis*,2(3):226-237.
47. Sultan ,N. and Arayne, M. et al. 2007. A validated method for the anlysis of Diltiazem in raw material and pharmaceutical formulation by RP-HPLC. *Pak J Pharm Sci*,20(4):284-90.
48. Zhang, L. and Zhao, F.2003.HPLC determination of Diltiazem in human plasma and its application to pharmacokinetics in humans. *Biomed Chromatogr*,17(8):522-525.
49. Sultana, N. and Muhammad, A. et al.2009.RP-HPLC method for analysis of Diltiazem:application to drug metal interaction.*J Chem Soc Pak* ,31(2):273-278.
50. Mosab, A.2014.Simple HPLC validated method for the determination of Diltiazem Hydrochloride in human plasma.*Int J Pharm Pharm sci* ,6(9):213-216.
51. Kumar, B. and Rajkamal, B. et al. B. 2018.Validated RP-HPLC method for the determination of Diltiazem in raw material and pharmaceutical dosage form. *International Journal of Pharmaceutical Science and Drug research*,10(6):487-491.
52. Sultana, N. and Saeed, A. et al. 2010.Development of a RP-HPLC method for the simultaneous analysis of Diltiazem and strain: application in pharmaceutical and human serum. *Journal : Analytical Methods*,2(10):1571-1576.
53. Khalid, H.2014.HPLC method development for the simultaneous determination and validation of Diltiazem Hydrochloride and its major metabolite Desacetyl Diltiazem Hydrochloride.*Der Pharma Chemica*,6(6):358-365.
54. Rabiarazzaq and Mohammad, R. et al.2018.Determination of Diltiazem hcl by reverse phase high performance liquid chromatography in rabbit plasma. *Current Pharmaceutical Analysis*,14(2).153-156.
55. Rodrigo, M. and Kleber, G. et al.2007. Analysis of Adenosine by RP-HPLC method and its application to the study of Adenosine kinase kinetics. *Journal of Separation Science*,30(15):2473-2479.
56. Akhova, A. and Alexander, G.2017.HPLC- UV method for simultaneous determination o Adenosine triphosphate and its metabolites in mycobacterium smegmatis. *Acta Chromatographia* , 31(2019)1:45-48.
57. Sottofattori, E. and Anzaldi , M. et al. 2001.HPLC determination of Adenosine in human synovial fluid.*J Pharm Biomed Anal*,24(5-6):1143-1146.
58. Yukari, L. and Youichib, F. et al. 1995.Quantitative HPLC analysis of cardiac glycosides in *Digitalis purpurea* leaves. *J Nat Prod*,58(6):897-901.
59. Miroslav, Z. and Valentiana, Marinkovik, D. et al.2010.An HPLC method for the determination of Digoxin in dissolution samples. *Journal of Serbian Chemical Society*,75(11):1583-1594.
60. Zhaohong, T.2009.determination of Isoprenaline Hydrochloride injection by HPLC.*China Pharmaceutical*,2009(06):1-4.
61. Chauhan, P. and Patel,B. et al. 2016.Sensitive RP-HPLC method for estimation of Atropine Sulphate and Dexamethasone Sodium Phosphate in ophthalmic formulation. *Journal of Current Pharma Research*,6(2):1770-1776.
62. Raghuwanshi, A. and Jain, U.2009.RP- HPLC method development for estimation of Atropine Sulphate in bulk drug. *Oriental Journal of Chemistry*,25(3):621-624.
63. Santtoni,G. and Tonsini, A. et al. 1993.Determination of Atropine Sulphate and Benzalkonium Chloride in eye drops by HPLC.*International Journal of Pharmaceutics*,93(1-3):239-243.