

Study of AM Fungi Associated with some plants of Acanthaceae

R.Chandra Sagar*¹, B.Bhadraiah¹

1. Applied Mycology and Molecular Plant Pathology Laboratory, Department of Botany, Osmania University, Hyderabad, Telangana, India.

Abstract

Some Medicinal, Ornamental and Natural weed plants are belonging to the family of Acanthaceae investigated for AM Fungal association. The plants from Acanthaceae family, *Andrographis echiodes*(L.)Nees., *Andrographis paniculata* Nees, *Barleria prionitis* L. *Crossandra infundibuliformis* (L.)Nees, *Depterocanthus prostratus* (Poir), *Hygrophila auriculata*(Schumach.&Thonn.)Heine, *Justicia glauca* Rottler, *Lepidagathis cristata* willd, *Peristrophe bicalyculata*(Retz.)Nees., *Ruellia tuberosa* L. were studied in this investigation 81 AM Fungal spores Associated with these plants were identified up to species level.

Key words: Arbuscular mycorrhiza, Acanthaceae, Fungal spores

Introduction

Mycorrhizae are non-pathogenic symbiotic soil growths which invade the root arrangement of plants. Arbuscular Mycorrhizal Fungi (AMF) are associated with about 80% of the plant families on the planet (Giovannetti and Sbrana 1998). The event of Arbuscular Mycorrhizal Fungi has been accounted for in many plant communities such as woods (Raman et al. 1993; Sengupta et al. 1998), grasslands, steppes and prairies (Sanders and Fitter 1992), deserts (Khaliel 1988) and mangroves (Sengupta and Chaudhuri 1989). The primary preferred position of mycorrhiza is its more prominent soil investigation and expanding take-up of P, N, K, Zn, Cu, S, Fe, Mg, Ca and Mn and the supply of these nutrients to the host roots (Sundar et al. 2010; Javot et al. 2007). Arbuscular Mycorrhizal Fungi (AMF) can likewise incite changes in the accumulation of secondary metabolites, including phenolics, in host plant roots (Vierheilig et al. 2000; Devi and Reddy 2002; Yao et al. 2003). Relatively little is thought about the impacts of AM colonization on the accumulation of dynamic phytochemicals in shoots of medicinal plants, which are frequently the harvest items. In any case, it was as of

late detailed that *Glomus mosseae* straightforwardly expands the essential oil content in shoots of *Origanum* sp. (Khaosaad et al. 2006) just as sweet basil (Copetta et al. 2006).

Material and Methods

Rhizosphere soil samples were collected from the family members of Acanthaceae from Warangal, Khammam, Hyderabad and Rangareddy districts of Telangana State. AM Fungal spores were isolated from the soil samples by Wet-sieving and decanting method (Gerdemann and Nicolson 1963). The identification was done based on Morphotaxonomic criteria (Schenck and Perez 1990).

Results and Discussion:

In the present study reveals that the Acanthaceae was associated with 81 AM fungal species belonging to 8 genera viz., *Acaulospora*, *Ambispora*, *Archaeospora*, *Entrophospora*, *Gigaspora*, *Glomus*, *Intraspora*, and *Scutellospora*. Of which 18 species belongs to *Acaulospora*, 2 belong to *Ambispora* 1 belong to *Archaeospora*, 3 belong to *Entrophospora*, 7 belong to *Gigaspora* 42 belong to *Glomus*, 1 belong to *Intraspora*, 7 to *Scutellospora*.

The highest AM fungal spore count per 100gm of rhizosphere soil was recorded in Hyderabad with 272 AM fungal spores with 95.12 % root colonization followed by with 154 AM fungal spores with 97 % root colonization and Warangal with 207 AM fungal spores with 96.45 % root colonization. The lowest AM fungal spore count per 100gm of rhizosphere soil was recorded in Khammam with 168 AM fungal spores with 82.67 % root colonization. The lowest root colonization 73% was recorded in Khammam (Schenck N.C. and Perez Y. 1990). The AM spore number of Acanthaceae family was higher in Hyderabad, Rangareddy and Warangal samples compared to that of Khammam sample which might be due to the presence of sandy loam soil in Hyderabad, Rangareddy, Warangal and clay loam soil in the Khammam. Rachel et al (1993) reported more AM fungal infection in sandy loam soil followed by other soil types.

The AM fungal association with Acanthaceae of all the 4 Districts investigated in the present study (Table 1). From the research, we could conclude that the biodiversity of AM fungi was abundant, though *Glomus* was the dominant genus. The AM fungal spore density and root colonization varied markedly among 4 districts. Considering the potential application of AM fungi on Acanthaceae, it seems that more attention should be paid

to the predominant AM fungi during the process of their cultivation, especially mycorrhizal performance i.e., improving growth, increasing secondary metabolite production.

Table 1: The AM fungal association with Acanthaceae in 4 District

slno	AM Fungal species	Hyderabad	Rangareddy	Warangal	Khammam
1.	<i>Acaulospora bireticulata</i>	+	-	+	+
2.	<i>Acaulospora dilicata</i>	+	+	-	+
3.	<i>Acaulospora delatata</i>	-	+	+	-
4.	<i>Acaulospora elegans</i>	+	-	+	+
5.	<i>Acaulospora foveata</i>	+	+	+	-
6.	<i>Acaulospora gardemannii</i>	-	+	+	+
7.	<i>Acaulospora laevis</i>	+	-	+	-
8.	<i>Acaulospora mellea</i>	-	+	+	+
9.	<i>Acaulospora myriocarpa</i>	+	+	-	+
10.	<i>Acaulospora nicolsoni</i>	-	+	+	+
11.	<i>Acaulospora rehmi</i>	+	+	-	-
12.	<i>Acaulospora rugosa</i>	+	+	+	+
13.	<i>Acaulospora scrobiculata</i>	+	-	+	-
14.	<i>Acaulospora spinosa</i>	+	+	+	+
15.	<i>Acaulospora splendida</i>	+	+	+	-
16.	<i>Acaulospora sporocarpia</i>	-	+	+	+
17.	<i>Acaulospora tuberculata</i>	+	+	+	+
18.	<i>Acaulospora undulate</i>	-	+	-	+
19.	<i>Ambiospora fecundispora</i>	-	+	-	+
20.	<i>Ambiospora leptoticha</i>	+	+	+	-
21.	<i>Archaeospora trappei</i>	+	+	-	+

22.	<i>Entrophospora columbiana</i>	-	+	+	+
23.	<i>Entrophospora infrequens</i>	+	+	-	+
24.	<i>Entrophospora Schenckli</i>	-	+	+	+
25.	<i>Gigaspora albida</i>	+	+	+	-
26.	<i>Gigaspora candida</i>	-	+	+	+
27.	<i>Gigaspora decipiens</i>	+	+	+	-
28.	<i>Gigaspora gigantia</i>	-	+	-	+
29.	<i>Gigaspora margarita</i>	+	+	+	-
30.	<i>Gigaspora ramisporophora</i>	+	+	-	+
31.	<i>Gigaspora rosea</i>	+	-	+	+
32.	<i>Glomus aggregatum</i>	+	+	+	-
33.	<i>Glomus albidum</i>	-	+	-	+
34.	<i>Glomus arborensense</i>	+	-	+	-
35.	<i>Glomus austral</i>	+	-	+	+
36.	<i>Glomus celedonium</i>	-	+	+	-
37.	<i>Glomus canadense</i>	+	-	+	+
38.	<i>Glomus citricola</i>	-	+	+	-
39.	<i>Glomes clarum</i>	+	+	+	+
40.	<i>Glomus clavisporum</i>	-	+	-	+
41.	<i>Glomus constrictum</i>	+	+	+	-
42.	<i>Glomus diaphanum</i>	+	-	+	-
43.	<i>Glomus dimorphicum</i>	-	+	+	-
44.	<i>Glomus fasciculatum</i>	+	-	+	+
45.	<i>Glomus fistulosum</i>	+	+	+	-
46.	<i>Glomus fragilistratum</i>	-	+	+	+

47.	<i>Glomus fuegianum</i>	+	-	+	-
48.	<i>Glomus geosporum</i>	+	-	+	+
49.	<i>Glomus globiferum</i>	+	+	-	+
50.	<i>Glomus glomeratum</i>	+	-	+	+
51.	<i>Glomus heterosporum</i>	+	-	+	-
52.	<i>Glomus hoi</i> Berch	+	-	+	+
53.	<i>Glomus intraradices</i>	+	+	+	-
54.	<i>Glomus intraradix</i>	+	-	+	+
55.	<i>Glomus invermaium</i>	+	+	-	+
56.	<i>Glomus liquidambaris</i>	-	+	+	+
57.	<i>Glomus macrocarpum</i>	+	-	+	-
58.	<i>Glomus maculosum</i>	-	+	-	+
59.	<i>Glomus microaggregatum</i>	+	-	+	-
60.	<i>Glomus microcarpum</i>	-	+	-	+
61.	<i>Glomus minuta</i>	+	-	+	-
62.	<i>Glomus monosporum</i>	-	+	-	+
63.	<i>Glomus mosseae</i>	+	-	+	-
64.	<i>Glomus multicaule</i>	-	+	-	+
65.	<i>Glomus multisubtensum</i>	+	-	+	-
66.	<i>Glomus pakistanica</i>	-	+	-	+
67.	<i>Glomus palladium</i>	+	-	+	-
68.	<i>Glomus pulvinatum</i>	-	+	-	+
69.	<i>Glomus rubiformis</i>	+	-	+	-
70.	<i>Glomus segmentatum</i>	-	+	-	+
71.	<i>Glomus sinuosa</i>	+	-	+	-
72.	<i>Glomus tenebrosum</i>	-	+	-	+

73.	<i>Glomus warcupli</i>	+	-	+	-
74.	<i>Intraspora schenckii</i>				
75.	<i>Scutellospora arenicola</i>	-	+	-	+
76.	<i>Scutellospora auriglobosa</i>	+	-	+	-
77.	<i>Scutellospora calospora</i>	-	+	-	+
78.	<i>Scutellospora pellucida</i>	-	+	+	-
79.	<i>Scutellospora sculata</i>	+	-	+	-
80.	<i>Scutellospora tricalyptra</i>	+	+	-	+
81.	<i>Scutellospora verrucosa</i>	-	+	-	+

References:

1. Copetta A., G. Lingua, and G. Berta, 2006. Effects of three AM fungi on growth, distribution of glandular hairs, and essential oil production in *Ocimum basilicum* L. var. Genovese. *Mycorrhiza*. 16:485-494.
2. Gerdemann JW and Nicolson TH. 1963. Spores of mycorrhizal *Endogone* species extracted from soil by wet sieving and decanting. *Trans Br Mycol Soc* 46: 235-244.
3. Giovannetti M., and C. Sbrana, 1998. Meeting a non-host: the behavior of AM fungi. *Mycorrhiza*. 8:123–130.
4. Khaliel A.S., 1988. Influence of vesicular-arbuscular mycorrhizae in some desert plants and correlation with edaphic factors. In: Mahadevan A, Raman N, Natarajan K (eds) *Mycorrhiza for Green Asia*, first asian conference on mycorrhizae. University of Madras, India, pp 56–59.
5. Khaosaad T., H. Vierheilig, M. Nell, K. Zitterl-Eglseer, and J. Novak, 2006. Arbuscular mycorrhiza alters the concentration of essential oils in *oregano* (*Origanum* sp., Lamiaceae). *Mycorrhiza* 16:443–446.

6. Raman N., N. Nagarajan, and S. Gopinathan, 1993. Occurrence of VAM fungi in Kolli hills of Tamil Nadu, India. In: Boger (eds) Proceedings of the second asian conference on mycorrhiza, Indonesia, pp 51–55.
7. Sanders I.R., and A.H. Fitter, 1992. Evidence for differential responses between host-fungus combination of vesicular arbuscular mycorrhizae from a grassland. *Mycol Res.* 96:415– 419
8. Schenck MC and Perez Y. 1990. *Manual for the identification of VA mycorrhizal fungi.* Synergistic Publ, Gainesville,Florida,USA. 283 p.
9. Sengupta A., and S. Chaudhari, 1990. Vesicular arbuscular mycorrhiza (VAM) in pioneer Salt marsh plants of the Gangesriver delta in West Bengal (India). *Plant soil.*122:111-113.
10. Sundar S.K., A. Palavesam, and B. Parthipan, 2010. Effect of native dominant AM fungus and PGPRs on growth and biochemical characteristics of medicinally important *Indigofera aspalathoides* Vahl.ex. DC. *Int J Biol Biotechnol.*7(1–2):59–67.
11. Vierheilig H., H. Gagnon, D. Strack, and W. Maier, 2000. Accumulation of cyclohexenone derivatives in barley, wheat and maize roots in response to inoculation with different arbuscular mycorrhizal fungi. *Mycorrhiza* 9:291–293.