

METHOD DEVELOPMENT AND VALIDATION OF RP-HPLC METHOD FOR ESTIMATION OF BOSUTINIB IN BULK AND PHARMACEUTICAL DOSAGE FORM

Ms. Sonal S. Fande, Mrs. Deepali S. Ghorpade

Department of quality assurance, Government College of pharmacy, Amravati.

ABSTRACT

An accurate, simple, precise, rapid and robust RP-HPLC method has been developed for estimation of bosutinib in bulk and pharmaceutical dosage form. The separation was achieved on C18 column (250 mm×4.6mm, 5µm), using Acetonitrile: Methanol: water (80:5:15) as a mobile phase, at flow rate 1.0 ml/min. Detection was carried out at 246 nm and drug eluted with a retention time were 5.63 min. Beer's law was obeyed with the concentration range of 20-100µg/ml with correlation coefficient 0.999. The method has been validated according to ICH guidelines for linearity, accuracy, precision, robustness, range, LOD and LOQ. The method was found to be accurate, simple, precise, robust and rapid. The proposed method was convenient for quantitative routine analysis and quality control of bosutinib in bulk and pharmaceutical dosage form.

Keywords- bosutinib, RP-HPLC, Validation.

INTRODUCTION

Bosutinib is a cancer medication prescribed to treat leukemia¹. It operates by inhibit protein associated with cancer cell growth in order to relieve symptoms, prevents the spread of cancer cells and aid other treatments. The U.S. FDA approved bosutinib in 2012 to treat chronic myelogenous leukemia (CML), blood and bone marrow disease that usually affects older adults. The drug is designed to inhibit tyrosine kinases such as bcr-abl and src used to treat chronic myelogenous leukemia (CML)^{2,3}.

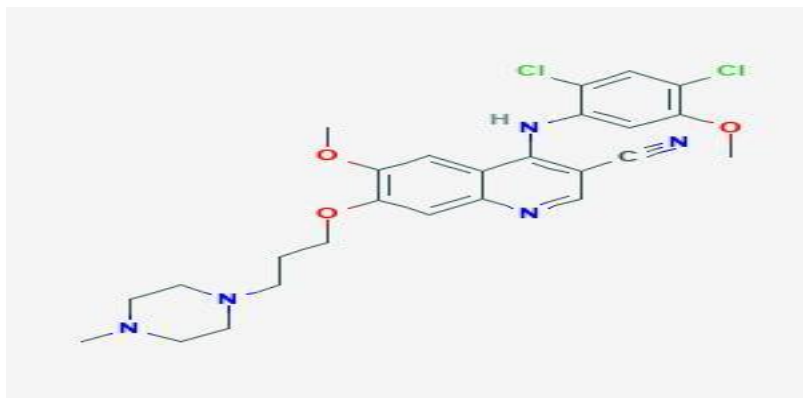


Fig. No. 1 Chemical structure of Bosutinib

The chemical name of bosutinib is 4-(2, 4-dichloro-5-methoxyanilino)-6-methoxy-7-[3-(4-methylpiperazin-1yl) propoxy] quinoline-3-carbonitrile. It has a molecular formula of $C_{26}H_{29}Cl_2N_5O_3$ and molecular weight of 530.45 g/mol. It has the structure formula (fig.1). Bosutinib is a yellow crystalline powder which is soluble in dimethyl sulfoxide, ethanol, and acetonitrile^{4,5}.

The literature survey revealed that the drug has been estimated by liquid chromatography method in biological fluids like human plasma and ultraviolet detection, RP-HPLC method for pharmaceutical formulation, RP-HPLC method in bulk form has been reported so far.

The aim of present work is to develop and validate simple, accurate, robust, rapid and precise RP-HPLC method for estimation of bosutinib in bulk and pharmaceutical dosage form.

MATERIALS AND METHODS

Instruments

Chromatographic separation was performed on water 2695 HPLC system equipped with C18 column (250mm × 4.6mm, 5.0 μm), Single pump, degasser, Dual λ absorbance detector and injector with 10 μl loop volume.

Chemicals and reagents

Bosutinib pure form was obtained from cubic lab Ankleshwar. The Acetonitrile (HPLC grade), Water (HPLC grade), Methanol (HPLC grade), Orthophosphoric acid (AR grade) were used.

Preparation of mobile phase

The mobile phase consisted of Acetonitrile: Methanol: Water (80:05:15 v/v/v) and pH adjusted to 7.0 with ortho phosphoric acid was used. The elution mode used was Isocratic. Mobile phase was filtered through a 0.22 μm nylon membrane filter and degassed prior to use.

Preparation of standard stock solution

A standard stock solution of Bosutinib (1000 μg/ml) was prepared by dissolving 100 mg of drug in 100 ml of volumetric flask and dissolve with 100 ml of Acetonitrile. The obtained concentration of bosutinib was 1000 μg/ml. The resulting solution was sonicated for 15 min.

Optimization of analytical wavelength

Optimization of analytical wavelength was done by scanning bosutinib standard solution in the range of 200-400 nm. By observing the spectra of standard solution λ_{max} at 246 nm were taken for trials to develop UV method.

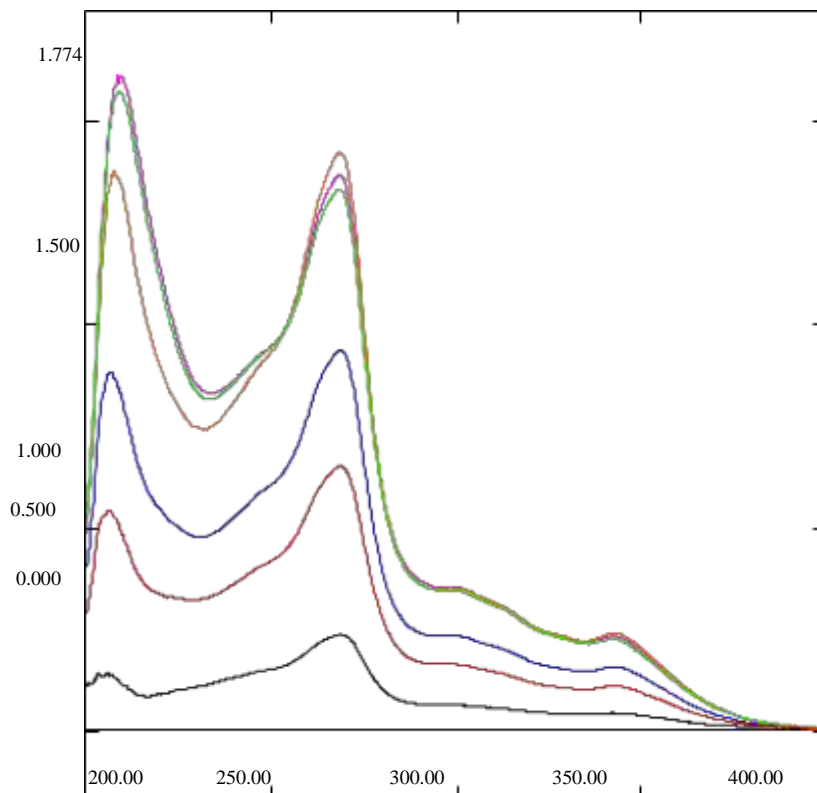


Fig 2. UV spectra of bosutinib

Optimized chromatographic conditions

Optimization of chromatographic conditions were done by performing different trials by taking different mobile phase and varying their compositions, flow rates. Finally an optimized chromatogram was obtained. The system suitability parameters were.

Retention Time (min)	: 5.63
Peak Area	: 689554
Theoretical plates	: 8653
Asymmetry (10%)	: 0.26
Resolution	: 9.56

Procedure for calibration curve

The calibration curve was prepared in a concentration range of 20-100 $\mu\text{g/ml}$ by taking 0.2-1.0 ml of standard stock in 10 ml volumetric flask and volume was made up to the mark with mobile phase. The resulting solution were filtered through 0.45 μ membrane filter paper and the filtrate was used for analysis.

Estimation of bosutinib in tablet dosage form

The synthetic mixture of bosutinib was prepared. Below all ingredients were shift and blend to make uniformity of mixing. Take Powder equivalent about 10 mg BST in 100 ml volumetric flask. Dissolve Content in 25 ml of Acetonitrile and Sonicated for 15min. Dilute up to 100 ml with Solvent shake vigorously, Filter solution through Whatman filter paper No. 42 and further dilute. The sample formulations were injected and chromatograms were recorded at 246 nm. The amount of drug estimated from the calibration curve.

Sr. No	Drug-Excipient Name	Quantity (mg)	Quantity (mg)
1	Bosutinib	10	200
3	Starch	30	600
4	Magnesium Stearate	10	200
5	Lactose	20qs	400qs
Total		100	2000

RESULT AND DISCUSSION

A reverse phase HPLC method was developed by keeping in mind system suitability parameters i.e. tailing factor (T), number of theoretical plate (N), run time and cost effectiveness. The optimized method developed resulted in the elution of bosutinib at 5.63 min. fig 3 represents standard chromatogram of bosutinib. The total run time is 10 min. The system suitability tests are the integral part of method development and are used to ensure adequate performance of the chromatographic system. Retention time (R_t), number of theoretical plates (N), and peak asymmetric factor were evaluated for six replicate injections of the standard at working concentration. The results given in table 1.

In order to test the applicability of the developed method to a synthetic mixture was chromatographed at working concentration 60 $\mu\text{g/ml}$. The sample peak was identified by comparing the retention time with the standard drug. System suitability parameters were within the acceptance limits, ideal for the chromatographed sample. Integration of standard peak area was done and drug concentration was determined using the peak area concentration relationship obtained in the standardization step. The protocol affords reproducible assay of the drug in the sample ranging between 98-102%, which is the standard level in the any pharmaceutical quality control.

The obtained chromatogram showed no interference from excipients fig 3. The system suitability parameters were within the limits. The developed method was simple, economical and the retention time that showed that the method was rapid. The developed method was validated as per ICH guidelines.

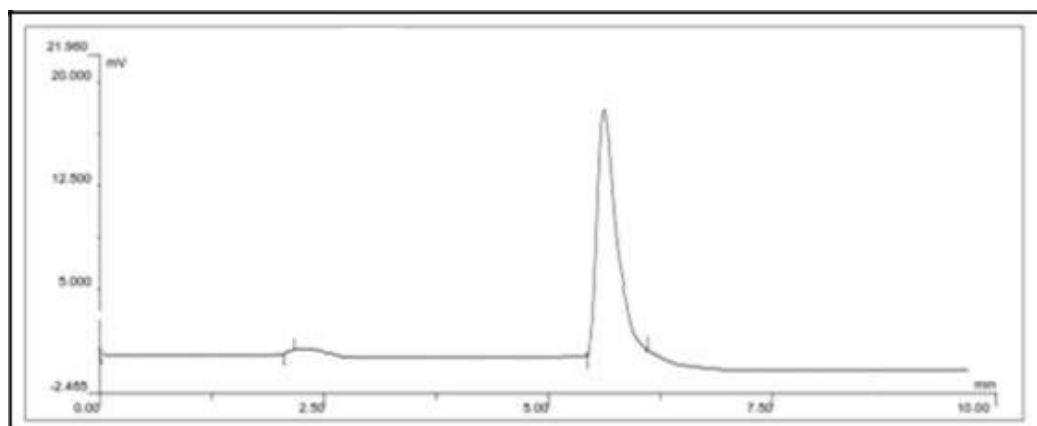


Fig 3. Chromatogram of bosutinib

PARAMETERS	Obser	IP'2010
	BST*	Specification
Retention Time (min)	5.63	RSD<1%
Peak Area	689554	-
Theoretical plates	8653	Not less than 2000
Asymmetry (10%)	0.26	Not greater than 2
Resolution	9.56	>2

Table 1 Result from system suitability studies

METHOD VALIDATION

Validation is the analytical method is the process that establishes by the laboratory studies in which performance characteristics of the method meet the requirements for the intended analytical application. RP-HPLC developed the method was validated according to International Conference on Harmonization (ICH) guidelines for validation of analytical procedures. The method was validated for the parameters like system suitability, specificity, linearity, precision, accuracy, robustness, and ruggedness, limit of detection (LOD) and limit of quantification (LOQ)^{6,7}.

Specificity

Specificity was checked for the interference of excipients in the analysis of sample solution and was determined by injecting sample solution with added excipients under optimized chromatographic conditions to demonstrate separation of bosutinib in from excipients. There is no interference of excipient peak on the peak of bosutinib indicating the high specificity of method^{8,9}.

Linearity

Calibration curve was plotted for different concentration of sample solution prepared from stock solution. Figure 4 shows linearity over the concentration range of 20-100 µg/ml shown in table 02. Each sample was injected six times and calibration curve was constructed by plotting the peak area verses drug concentration.

Concentration (µg/ml)	Peak area
20 µg/ml	30782.8
40 µg/ml	62367.5
60 µg/ml	95638
80 µg/ml	126485.7
100 µg/ml	158640.2

Table 2 linearity data for bosutinib

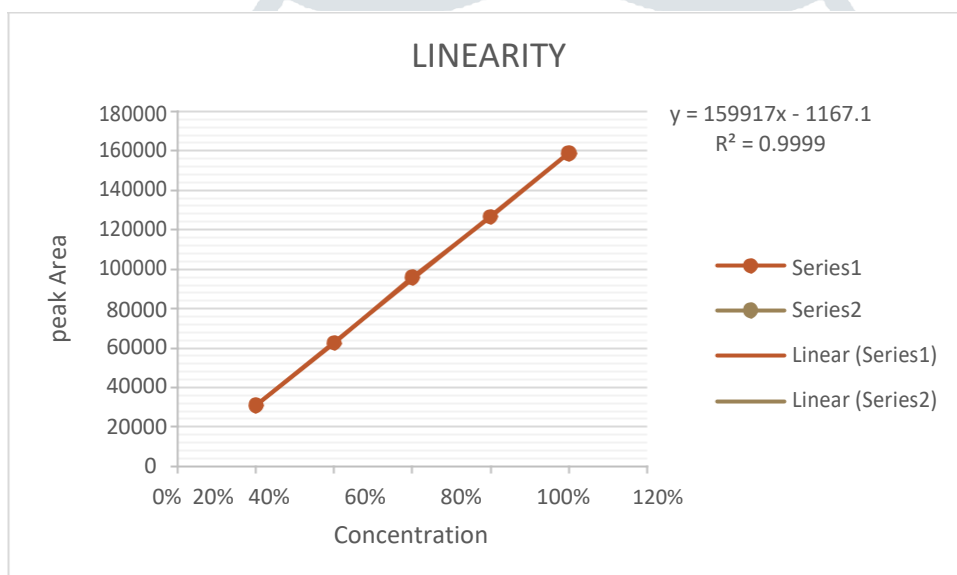


Figure 4. Calibration curve for bosutinib

Regression parameter	Bosutinib
Regression equation	$y = 159917x - 1167.1$
Slope	159917
Intercept	1167.1
Correlationcoefficient (r ²)	0.9999

Table 3. Regression parameters table for bosutinib

Precision

The precision of an analytical procedure expresses the closeness of agreement between a series of measurements obtained from multiple sampling of the same homogeneous sample under the prescribed conditions. The precision was Intraday and inter day precision¹⁰.

Intraday precision- it is determined by analyzing three different concentration solution of bosutinib within the same day with specific interval. Intraday precision is estimated by analyzing 40 μ g/ml, 60 μ g/ml, 80 μ g/ml, 100 μ g/ml within the same day.

Interday precision- it is determined by analyzing three different concentration of solution of bosutinib in three different days over the period of week. Inter day precision is estimated by above mention concentration for three different days over the period of week.

Sr no	40 μ g/ml	60 μ g/ml	80 μ g/ml
1	62245	95645	126494
2	62344	95630	126483
3	62350	95633	126583
AVG	62313	95636	126520
SD	58.96609	7.937254	54.83612
% RSD	0.094629	0.008299	0.043342

Table 4. Intraday precision

S r. N o	Day 1			Day 2			Day 3		
	40 μ g/ml	60 μ g/ml	80 μ g/ml	40 μ g/ml	60 μ g/ml	80 μ g/ml	40 μ g/ml	60 μ g/ml	80 μ g/ml
1	622 24	956 30	126 500	623 24	956 21	126 510	622 01	956 25	126 460
2	623 60	956 45	126 440	623 32	956 60	126 502	622 31	956 31	126 450
3	623 50	957 00	126 450	623 60	956 01	126 555	623 31	956 45	126 440
A V G	623 11.3 3	956 58.3 3	126 463 .3	623 38.6 7	956 27.3 3	126 522. 3	622 54.3 3	956 33.6 7	126 450
S D	75.7 979 8	36.8 555 7	32. 145 5	18.9 032 6	30.0 055 6	28.5 715 5	68.0 685 9	10.2 632	10
% R S D	0.12 164 4	0.03 852 8	0.0 254 19	0.03 032 3	0.03 137 8	0.02 258 2	0.10 934	0.01 073 2	0.00 790 8

Table 5. Interday precision

Accuracy

The accuracy of the method was obtained by spiking 80, 100 and 120% of bosutinib standard concentration, in which the amount of marketed formulation kept constant and the amount of pure drug was varied. Solution were prepared in triplicates and accuracy was indicated by % recovery which was between 99.83 to 100.18%. The result were shown in table 6.

Level	Amount taken	Amount added	Total amount	Found area	Concentration	% recovery
80 µg/ml	20	16	36	56390	35.99	99.97
	20	16	36	56450	36.03	100.08
	20	16	36	56299	35.94	99.83
100 µg/ml	20	20	40	62698	39.94	99.85
	20	20	40	62760	39.97	99.85
	20	20	40	62771	39.98	99.92
120 µg/ml	20	24	44	69322	44.08	100.18
	20	24	44	69110	43.94	99.86
	20	24	44	69185	43.99	99.98

Table 6. Accuracy of bosutinib

Robustness

Robustness was carried by varying three parameters deliberately from the optimized chromatographic conditions like mobile phase, flow rate and pH. The RSD was found to be < 2, shown in table 7.

Parameters	values	Peak area	AVG	SD	% RSD
PH	7.0	689554	689553.3	295.4426	0.042847
	7.01	689545			
	6.95	689561			
Flow rate	1.0	689554	689535.3	133.665	0.019384
	1.1	689231			
	0.9	689821			
Mobile phase composition (ACN:methanol:water)	80:05:15	689554	689546.3	8.020806	0.001163
	82:03:15	689676			
	70:10:20	689409			

Table 7. Robustness results of bosutinib

Limit of detection and limit of quantification

The LOD and LOQ of the present method were calculated based on standard deviation of the response and slope of linearity curve^{11,12}. The LOD & LOQ values of bosutinib were found to be 0.00062 ug/ml and 0.001879 ug/ml.

CONCLUSION

Thus, the developed method was found to easy, simple, rapid, robust, selective and precise for the routine estimation of bosutinib in bulk and pharmaceutical dosage form.

REFERENCES

- 1) Bowles, P., Busch, F. R., Leeman, K. R., Palm, A. S., & Sutherland, K. (2015). Confirmation of Bosutinib Structure; Demonstration of Controls to Ensure Product Quality. *Organic Process Research and Development*, 19(12), 1997–2005. <https://doi.org/10.1021/acs.oprd.5b00244>.
- 2) Assi, R., Kantarjian, H., Short, N. J., Daver, N., Takahashi, K., Garcia-Manero, G., ... Jabbour, E. (2017). Safety and Efficacy of Blinatumomab in Combination With a Tyrosine Kinase Inhibitor for the Treatment of Relapsed Philadelphia Chromosome-positive Leukemia. *Clinical Lymphoma, Myeloma and Leukemia*, 17(12), 897–901. <https://doi.org/10.1016/j.clml.2017.08.101>
- 3) Pasic, I., & Lipton, J. H. (2017). Current approach to the treatment of chronic myeloid leukaemia. *Leukemia Research*, 55, 65–78. <https://doi.org/10.1016/j.leukres.2017.01.005>.
- 4) Moy, B., Neven, P., Lebrun, F., Bellet, M., Xu, B., Sarosiek, T., ... Lang, I. (2014). Bosutinib in Combination With the Aromatase Inhibitor Exemestane: A Phase II Trial in Postmenopausal Women With Previously Treated Locally Advanced or Metastatic Hormone Receptor-Positive/HER2-Negative Breast Cancer. *The Oncologist*, 19(4), 346–347. <https://doi.org/10.1634/theoncologist.2014-0022>
- 5) Ono, C., Hsyu, P. H., Abbas, R., Loi, C. M., & Yamazaki, S. (2017). Application of physiologically based pharmacokinetic modeling to the understanding of bosutinib pharmacokinetics: Prediction of drug-drug and drug-disease interactions. *Drug Metabolism and Disposition*, 45(4), 390–398. <https://doi.org/10.1124/dmd.116.074450>.
- 6) Xu, Y., Huang, X. ce, Dai, S., Xiao, Y., & Zhou, M. tao. (2015). A simple method for the determination of Bosutinib in rat plasma by UPLC-MS/MS. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 1004, 93–97. <https://doi.org/10.1016/j.jchromb.2015.09.030>
- 7) Wang, L., Tang, L., Zheng, Y., Pan, G., Zhu, W., Pan, C., & Zhu, L. (2015). Determination of bosutinib in mice plasma and tissue by UPLC-MS/MS and its application to the pharmacokinetic and tissue distribution study. *Analytical Methods*, 7(21), 9184–9189. <https://doi.org/10.1039/c5ay01529d>.
- 8) Sumimoto, T., Nakahara, R., Sato, Y., & Itoh, H. (2018). A quantitative method for the determination of bosutinib in human plasma using high-performance liquid chromatography and ultraviolet detection. *Journal of Clinical Laboratory Analysis*, 32(1). <https://doi.org/10.1002/jcla.22201>.
- 9) Naga Sindhu, S., Srinivasa Rao, Y., Hemant Kumar, T., & Vara Prasada Rao, K. (2015). Method development and validation of RP-HPLC method for estimation of imatinib mesylate in pure and pharmaceutical dosage form. *Der Pharmacia Lettre*, 7(3), 33–38.
- 10) Chaudhari, V. L., & Kulkarni, A. A. (2016). *RP-HPLC METHOD FOR ESTIMATION OF BOSUTINIB IN BULK*. 5(12), 417–424. <https://doi.org/10.20959/wjpr201612-7485>.
- 11) Article, R., Mohammad, A. S., Opera, B. M., Opera, B. M., & Nadu, T. (2018). *Estimation of anticancer and antiviral drugs in bulk and varied dosage forms by advanced analytical techniques : a detailed review*. 7(10), 1651–1667. <https://doi.org/10.20959/wjpps201810-12570>.
- 12) Jadhav, P. B., Gajare, G. K., & Ambedkar, B. (2016). *Development and validation of an rp-hplc method for bosutinib in bulk form*. 6(3), 599–603.

13)

