

# Potential leads for development of new antimalarial drugs

Savita A.Patil, Amol Thete, Sudarshan Shelke, Jyoti Kondhalkar, Dwarkanath bhagat

Bioreinventors LLP, Pune; Department of Biotechnology.

## Abstract:

*Malaria is a dreadful disease caused by the Abstract:*

*parasite Plasmodium and is transmitted to humans by mosquitoes. Malaria remains an important public health problem, especially in endemic regions of India. Globally, malaria remains a leading infectious disease, and especially so in Sub-Saharan Africa and South East Asia. Global efforts are underway to eliminate malaria and to use a multi-pronged strategy where drugs play a crucial part. The most effective present day line of treatment option is based on the artemisinin-based drugs. The malarial parasites are developing resistance to the artemisinin class of drugs; it is likely that one day these drugs will be ineffective. Therefore, there is an urgent need to develop new classes of anti-malaria drugs with novel modes of action. Cladosporin (also known as asperentin), 3,4-dihydro-6,8-dihydroxy-3-(6-methyl tetrahydropyran-2-ylmethyl) isocoumarin, is an important secondary metabolite isolated from Cladosporium cladosporioides in 1971. It is the major compound of C. cladosporioides, but a minor metabolite of other fungal sources including Aspergillus flavus. In Present study we have assessed the fungal metabolite-inspired molecules (Cladosporin stereoisomers) as potential lead antimalarials. Novel synthetic routes were developed in the laboratory to access this natural product. In addition, the team has synthesized all the possible stereoisomers of Cladosporin using novel synthetic organic chemistry protocols. After the successful synthesis of all eight compounds (called Cladologs), the teams tested it against malaria parasites to address their potency. Enzyme and structure-based studies were done to address mechanistic details of the drug interactions. The important cladologs were co-crystallized with the target enzyme lysyl-tRNA synthetase of malaria parasite in order to provide atomic details. The bases for wide differences in antimalarial potency between various*

*stereoisomeric forms of cladosporin using an elegant chemistry, strong biochemistry and modern structure-based methods. Three categories of molecules as potent, moderately potent and non-potent were identified based on target binding and parasite killing. The demonstrations validated two most potent stereoisomers of cladosporin so this information will allow their development for drug-like properties. The significance of chirality in modern drug discovery has also been highlighted through these efforts.*

## Introduction:

Malaria is a common and life-threatening disease in many tropical and subtropical areas. There are currently over 100 countries and territories where there is a risk of malaria transmission, and these are visited by more than 125 million international travellers every year. Each year many international travellers fall ill with malaria while visiting countries/territories where malaria is endemic, and well over 10 000 are reported to become ill with malaria after returning home; however, underreporting means that the real figure may be considerably higher. International travellers to countries/territories with ongoing local malaria transmission arriving from countries with no transmission are at high risk of malaria infection and its consequences because they lack immunity. Migrants from countries/territories with malaria transmission living in malaria-free countries and returning to their home countries to visit friends and relatives are similarly at risk because of waning or absent immunity. Travellers who fall ill during travel may find it difficult to access reliable medical care. Travellers who develop malaria upon returning to a country that is malaria-free face particular problems: medical personnel may be unfamiliar with malaria, the diagnosis may be delayed, and effective antimalarial medicines may not be registered and/or available, resulting in progression to fatality rates.—severe and complicated malaria and, consequently, high case Fever occurring in a traveller

within 3 months of leaving a country in which there is risk of malaria is a potential medical emergency and should be investigated urgently to exclude malaria. In the absence of rapid access to reliable diagnostic facilities, stand-by emergency treatment (SBET) is indicated.

In Present study we have assessed the fungal metabolite-inspired molecules (Cladosporin stereoisomers) as potential lead antimalarials. Novel synthetic routes were developed in the laboratory to access this natural product. In addition, the team has synthesized all the possible stereoisomers of Cladosporin using novel synthetic organic chemistry protocols. After the successful synthesis of all eight compounds (called Cladologs), the teams tested it against malaria parasites to address their potency. Enzyme and structure-based studies were done to address mechanistic details of the drug interactions. The important cladologs were co-crystallized with the target enzyme lysyl-tRNA synthetase of malaria parasite in order to provide atomic details. The bases for wide differences in antimalarial potency between various stereoisomeric forms of cladosporin using an elegant chemistry, strong biochemistry and modern structure-based methods. Three categories of molecules as potent, moderately potent and non-potent were identified based on target binding and parasite killing. The demonstrations validated two most potent stereoisomers of cladosporin so this information will allow their development for drug-like properties. The significance of chirality in modern drug discovery has also been highlighted through these efforts.

### Materials and Methodology:

#### Protein Sequence retrieval and Primary analysis:

Protein sequence of protein minor nucleoprotein Cladosporin (ribosomal protein S5 [*Aspergillus flavus*]) was retrieved from Gene bank database. The physicochemical analysis were calculated by ProtParam tool (<http://web.expasy.org/protparam/>), including *pI*, total number of negatively and positively charged residues, the instability index (II), aliphatic index, and grand average of hydrophilic (GRAVY).

#### Structural Charecterization:

Similarity search was carried out by using BLAST software (<https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE=Proteins>). SOPMA (Geourjon and Deléage, 1995) server (<https://npsaprabi.ibcp.fr/cgi->

[bin/npsa\\_automat.pl?page=npsa\\_sopma.html](http://bin/npsa_automat.pl?page=npsa_sopma.html)). SOPMA is using homologue method of Levin *et al.* According to this method; short homologous sequence of amino acids will tend to form similar secondary structure. As well it also done by using Phyre2 (<http://www.sbg.bio.ic.ac.uk/~phyre2/html/page.cgi?id=index>) software and visualized by using Chimera (<https://www.cgl.ucsf.edu/chimera/>) software.

#### Homology modeling and Model evaluation:

Homology modeling was used for determining 3D structure of protein. Then, BLASTP was performed against PDB (Protein Databank, Bernstein *et al.*, 1977) to retrieve the best suitable templates for homology modeling. Preferred hit contains maximum identity and lowest e-value that it was used as a template. The modeling of the 3D structure of the protein was performed by using Swiss-Modeler (<http://swissmodel.expasy.org/>) program (Arnold *et al.*, 2006; Bordoli *et al.*, 2009).

#### Molecular Docking:

Molecular docking is an attractive scaffold to understand drug-biomolecular interactions for the rational drug design and discovery, as well as in the mechanistic study by placing a molecule (ligand) into the preferred binding site of the target specific region of the DNA/protein (receptor) mainly in a non-covalent fashion to form a stable complex of potential efficacy and more specificity. The information obtained from the docking technique can be used to suggest the binding energy, free energy and stability of complexes. At present, docking technique is utilized to predict the tentative binding parameters of ligand-receptor complex beforehand. In this project docking was carried out between Cladosporin and ribosomal proteins5 to find proper drug structure for future applications.

#### Binding site Prediction:

The binding site of Cladosporin protein was predicted by RaptorX server (<http://raptorx.uchicago.edu/BindingSite/>). The binding site shows the small pockets of the tertiary structure where ligands bind to using the weak forces.

1] Organism : *Aspergillus flavus*

2] Protein : ribosomal protein S5

3] Accession id: Gen Bank: RAQ63165.1

4] Sequence:

>RAQ63165.1 ribosomal protein S5 [*Aspergillus flavus*]

MADAAPRGRGGFGRGDRGGDRGRGRRRRGGK  
 QEEKEWQPVTKLGRVLKAGKITSMEQIYLHSLPIKEY  
 QIVDFFLPKLKDEVMKIKPVQKQTRAGQRTRFKAVVI  
 IGDSEGHIGLGIKTSKEVATAIRAAITIAKLAVLPVRRG  
 YWGSNLGEPHSLPVKQSAKCGSVSVRLIPAPRGTLV  
 ASPAVKRLQLAGVQDAYTSSSGSTKTLENTLKATFL  
 AVVNTYGFLLTPNLWKETKLIRSPLEEFQDVLVLRQGKKY

## 5) Drug used:

- I) **Source name :** *Aspergillus flavus*  
 II) **Chemical compound :** Cladosporin  
 III) **Pubchem id:** 13990016  
 IV) **Molecular Formula:** [C<sub>16</sub>H<sub>20</sub>O<sub>5</sub>](#)  
 V) **Chemical Names:** Cladosporin  
 Asperentin  
 UNII-81PR0D5FI4  
 81PR0D5FI4  
 35818-31-6

VI) **Related compounds with annotation:**

1. Taleranol
2. Mellein

3. (4S)-8,16,18-Trihydroxy-4-methyl-3-oxabicyclo[12.4.0]octadeca-1(14),15,17-trien-2-one
4. Altenuene
5. Isocoumarin, 3,4-dihydro-6,8-dihydroxy-3-(6-methyl-tetrahydro-2H-pyran-2-yl)

**Results and discussion:****Protein Sequence retrieval and Primary analysis:**

The physicochemical analysis of Ribosomal Protein S5 protein was performed using ProtParam and results were shown in Table 1. Protein contains 259 amino acids with molecular weight 28274.85 Dalton and Theoretical pI 10.54

Sr.No.	Parameters	Values
1	Molecular weight	28274.85 D
2	Theoretical pi	10.54
3	Instability index	37.17
4	Extinction coefficients	25440
5	Total number of negatively charged residues (Asp + Glu):	21
6	Total number of positively charged residues (Arg + Lys):	47
7	Aliphatic index:	86.22
8	GRAVY -	-0.396

**Table 3. Physico-chemical properties of Ribosomal Protein**

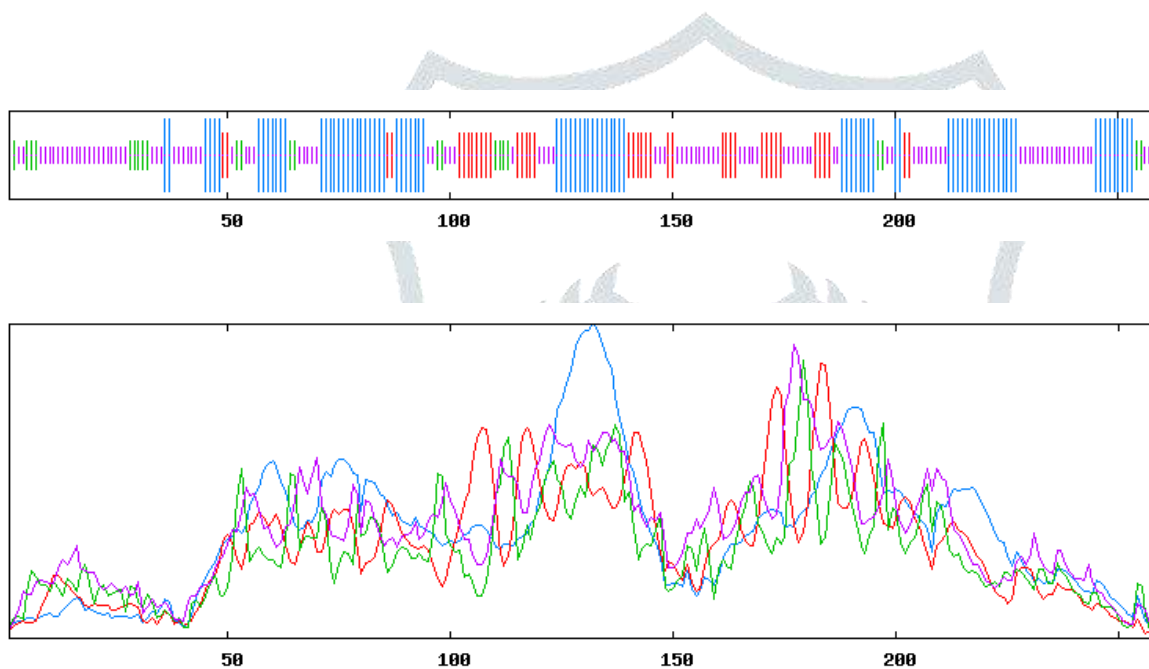
ProtParam tool computed that the protein is basic in nature and stable on the basis of parameters Theoretical pi and instability index. According to the GRAVY index protein is hydrophilic. The aliphatic index of a protein is 86.22 which defined as the relative volume occupied by aliphatic side chains (alanine, valine, isoleucine, and leucine). The total number of positively charged residues (Arg+Lys 47) was found higher than the total number of negatively charged residues (Asp+Glu 21).

**Structural Characterization:**

The secondary structure of the protein was predicted using SOPMA server. It was observed that predominant with alpha helix (33.98%) followed by random coil (41.31%), and extended strand (15.83%). Random coils have important functions in proteins for flexibility and conformational changes such as enzymatic turnover (Buxbaum, 2007).

**Table 4. Secondary structure of protein using SOPMA**

Sr.No.	Parameters	Values
1	Alpha Helix	80
2	Bita Sheets	23
3	Random coils	107
4	Extended strand	41

**Figure 5. Secondary structure of protein using SOPMA****Phyre2 Secondary structure prediction:**

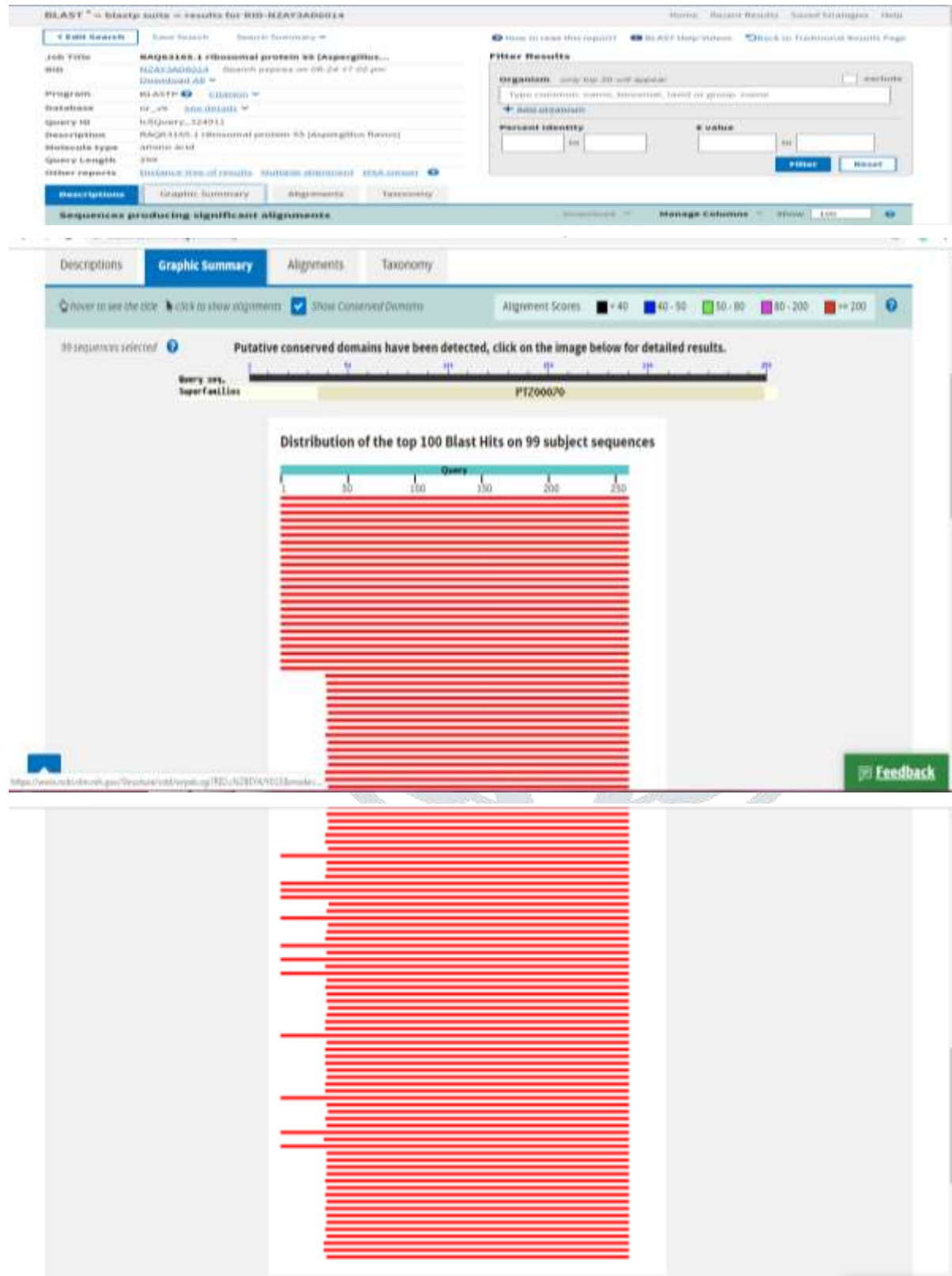
According to structure prediction by Phyre2 ribosomal s5 protein is known to antagonize interferon signaling by binding host karyopherin a proteins, The crystal structures and accompanying biochemical analysis map differences between pathogenic and nonpathogenic viruses, offer templates for drug design, and provide the three dimensional framework necessary for biological dissection of the many functions of ribosomal S5 protein.



analysis 118 structures of resolution of at least 2.0 Angstroms and R-factor no greater than 20.0 a good quality model would be expected to have over 90% in the most favored regions

(A,B,L) it also showed that only 4.2% residues in outlier region, 16.9% allowed region indicating that the models were of reliable and good quality.

**BLASTP Results:**



Descriptions | Graphic Summary | Alignments | Taxonomy

Sequences producing significant alignments

Download Manage Columns Show 100

select all 0 sequences selected

Description	Max Score	Total Score	Query Cover	E value	Pos. Ident	Accession
<input type="checkbox"/> 40S ribosomal protein (Aspergillus flavus AF70)	521	521	100%	0.0	100.00%	K0C0Z577.1
<input type="checkbox"/> 40S ribosomal protein S2 (Aspergillus oryzae R0640)	521	521	100%	0.0	100.00%	WP_005572513.1
<input type="checkbox"/> 40S ribosomal protein S2 (Aspergillus oryzae NRRL 11137)	521	521	100%	0.0	100.00%	WP_015493186.1
<input type="checkbox"/> ribosomal protein S2 (Aspergillus fumigatus)	521	521	100%	0.0	100.00%	XP_022384132.1
<input type="checkbox"/> Ribosomal protein S2.96 terminal domain protein (Aspergillus parasiticus S010)	521	521	100%	0.0	99.61%	KM57495.1
<input type="checkbox"/> 40S ribosomal protein S2 (Aspergillus nidulans CBS 702.7)	509	509	100%	0.0	96.53%	F7901231.1
<input type="checkbox"/> hypothetical protein A5FACQ5AFT_01671 (Aspergillus oryzae ATCC 16827)	509	509	100%	0.0	96.52%	XP_000926441.1
<input type="checkbox"/> Uncharacterized protein D3M27K5_03720 (Aspergillus nidulans)	508	508	100%	0.0	97.31%	XP_008089871.1
<input type="checkbox"/> 40S ribosomal protein S2 (Aspergillus terreus N1520)	507	507	100%	0.0	97.30%	XP_001217279.1
<input type="checkbox"/> 40S ribosomal protein (Aspergillus fumigatus)	506	506	100%	0.0	96.54%	WP025220.1
<input type="checkbox"/> ribosomal protein (Aspergillus terreus)	506	506	100%	0.0	96.54%	K0921643.1
<input type="checkbox"/> 40S ribosomal protein S2 (Penicillium subtilospora)	505	505	100%	2e-180	96.53%	GN052300.1
<input type="checkbox"/> hypothetical protein PDE_20301 (Penicillium notatum 134-2)	504	504	100%	3e-180	96.14%	EP531029.1
<input type="checkbox"/> hypothetical protein PF5NHL_01200201 (Penicillium ochrosporum)	504	504	100%	4e-180	96.54%	XP_012495413.1
<input type="checkbox"/> 40S ribosomal protein S2 (Aspergillus utiformis)	504	1009	100%	6e-180	96.53%	GN050936.1
<input type="checkbox"/> ribosomal protein (Aspergillus ochrosporum)	505	505	100%	6e-180	96.54%	K05

Download GenPept Graphics

40S ribosomal protein, partial [Aspergillus flavus AF70]

Sequence ID: K0C0Z577.1 Length: 295 Number of Matches: 1

Range 1: 17 to 295 GenPept Graphics

Score	Expect	Method	Identifiers	Positives	Gaps
521 bits (1343)	0.0	Compositional matrix adjust.	259/259 (100%)	259/259 (100%)	0/259 (0%)

```

Query 1  MDAQPRGRGFGSRGDRGGDRGGRGRGRGRGGKQEEKEWQPVTKLGRLVKAGKITSME 60
          MDAQPRGRGFGSRGDRGGDRGGRGRGRGRGGKQEEKEWQPVTKLGRLVKAGKITSME
Sbjct 37  MDAQPRGRGFGSRGDRGGDRGGRGRGRGRGGKQEEKEWQPVTKLGRLVKAGKITSME 96

Query 61  QIYLHSLPIKEYQIVDFFLPKLDEVWIKPVKQTRAGQTRFKAVVIIGDSEGHIGLG 120
          QIYLHSLPIKEYQIVDFFLPKLDEVWIKPVKQTRAGQTRFKAVVIIGDSEGHIGLG
Sbjct 97  QIYLHSLPIKEYQIVDFFLPKLDEVWIKPVKQTRAGQTRFKAVVIIGDSEGHIGLG 156

Query 121  IKTSKEVTAIRAAITIAKLAVLPVRGYNWGSILGEPHSLPVQOSARCGSVSVRLIPAP 180
          IKTSKEVTAIRAAITIAKLAVLPVRGYNWGSILGEPHSLPVQOSARCGSVSVRLIPAP
Sbjct 157  IKTSKEVTAIRAAITIAKLAVLPVRGYNWGSILGEPHSLPVQOSARCGSVSVRLIPAP 216

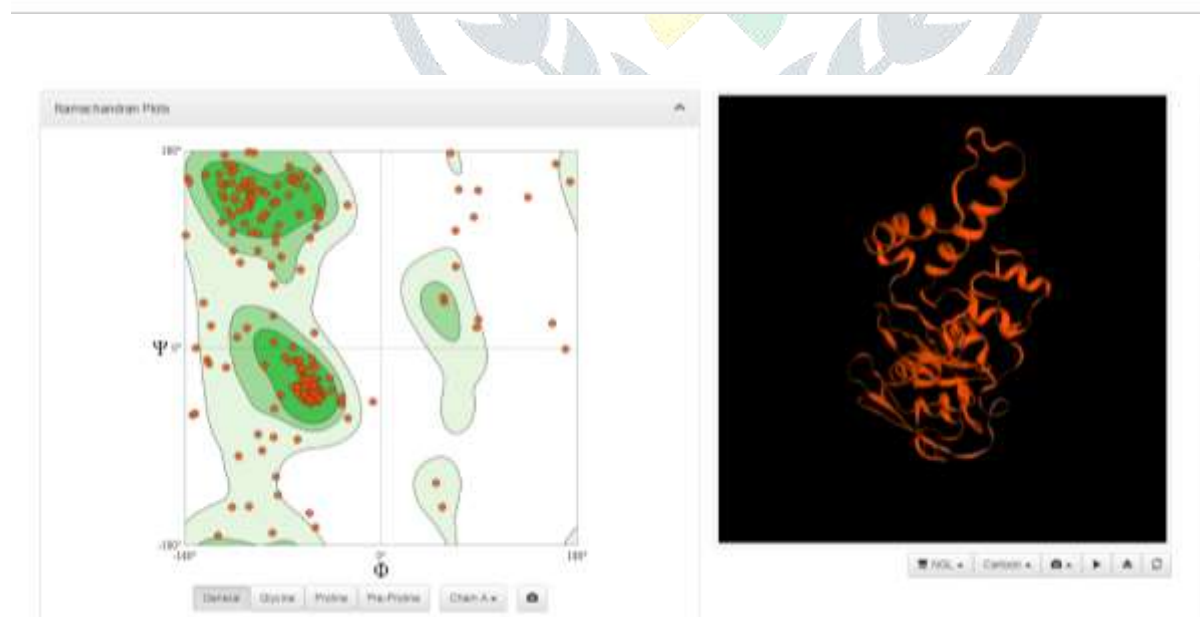
Query 181  GTGLVSPAWRLLQLAGVQDAYTSSGOSTKLENTLKATFLAVVITYGFLTPWVKETK 240
          GTGLVSPAWRLLQLAGVQDAYTSSGOSTKLENTLKATFLAVVITYGFLTPWVKETK
Sbjct 217  GTGLVSPAWRLLQLAGVQDAYTSSGOSTKLENTLKATFLAVVITYGFLTPWVKETK 276

Query 241  LIRSPLEEFQVLRGKKY 259
          LIRSPLEEFQVLRGKKY
Sbjct 277  LIRSPLEEFQVLRGKKY 295
    
```

Download GenPept Graphics

Next Previous Descriptions

SWISS modeling results




The SWISS-MODEL template library (SMTL version 2019-08-14, PDB release 2019-08-09) was searched with BLAST ([Camacho et al.](#)) and HHBlits ([Remmert et al.](#)) for evolutionary related structures matching the target sequence in Table T1. For details on the template search, see Materials and Methods. Overall 664 templates were found (Table T2).



Models:

The following models were built (see Materials and Methods "Model Building"):

Model #01	File	Built with	Oligo-State	Ligands	GMQE	QMEAN
	<a href="#">PDB</a>	ProMod3 Version 2.0.0.	monomer	None	0.75	-9.78

QMEAN

-9.78

C $\beta$

-5.13

All Atom

-6.21

solvation

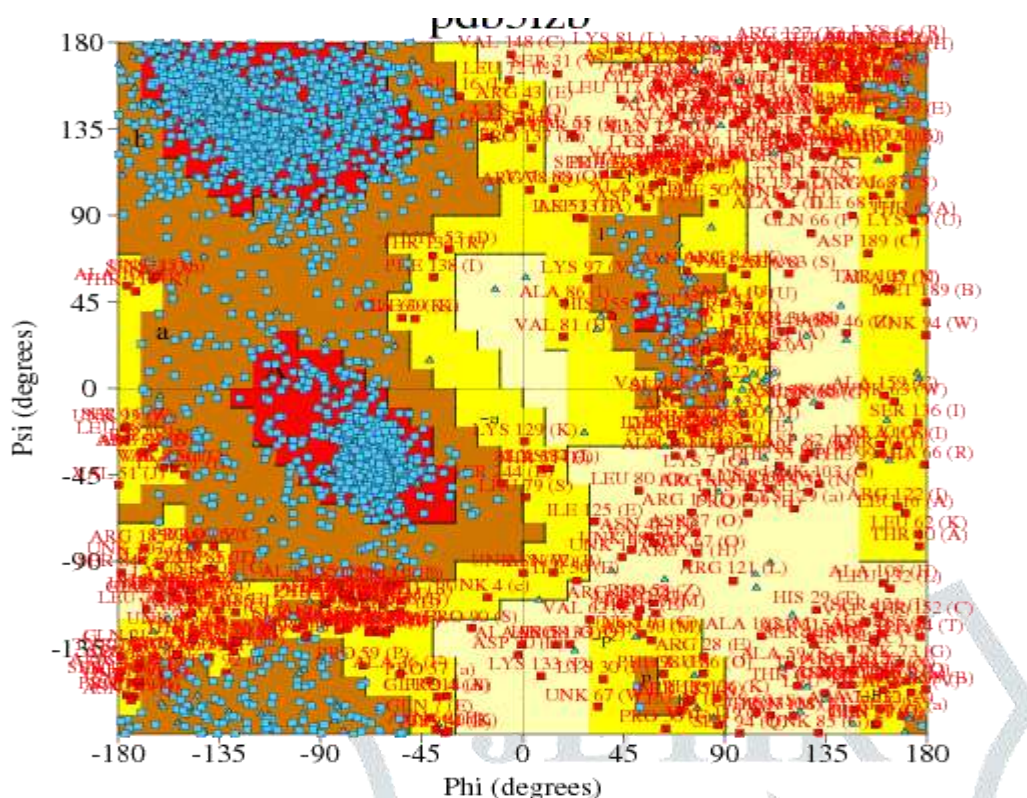
-5.04

torsion

-7.45

Template	Seq Identity	Oligo-state	Found by	Method	Resolution	Seq Similarity	Range	Coverage	Description
<a href="#">4v3p.5.A</a>	59.53	monomer	HHblits	EM	34.00Å	0.48	1 - 258	0.99	40S ribosomal protein S2

Structure validation by Ramachandran plot:



quality model would be expected to have over 90% in the most favoured regions [A,B,L].

PROCHECK statistics

1. Ramachandran Plot statistics

	No. of residues	%-tage
Most favoured regions [A,B,L]	2394	72.7%**
Additional allowed regions [a,b,l,p]	530	16.1%
Generously allowed regions [~a,~b,~l,~p]	228	6.9%
Disallowed regions [XX]	142	4.3%*
Non-glycine and non-proline residues	3294	100.0%
End-residues (excl. Gly and Pro)	64	
Glycine residues	259	
Proline residues	140	
Total number of residues	3757	

2. G-Factors

Parameter	Average Score
Dihedral angles:-	
Phi-psi distribution	-0.81*
Chi1-chi2 distribution	-0.66*
Chi1 only	-0.37
Chi3 & chi4	0.39
Omega	<b>-1.30**</b>
Main-chain covalent forces:-	
Main-chain bond lengths	<b>-2.64**</b>
Main-chain bond angles	<b>-3.88**</b>
	<b>-3.36**</b>
OVERALL AVERAGE	<b>-1.78**</b>

Based on an analysis of 118 structures of resolution of at least 2.0 Angstroms and R-factor no greater than 20.0 a good

**G-factors** provide a measure of how **unusual**, or out-of-the-ordinary, a property is.

Values below -0.5\* - unusual

Values below -1.0\*\* - highly unusual

**Important note:** The main-chain bond-lengths and bond angles are compared with the Engh & Huber (1991) ideal values derived from small-molecule data. Therefore, structures refined using different restraints may show apparently large deviations from normality.

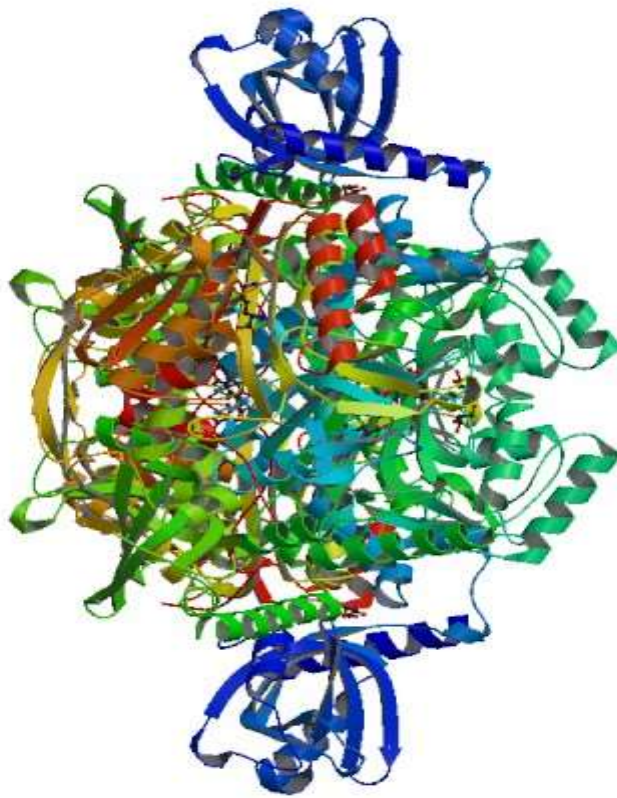
### Binding site prediction:

PDB code	Model	Length	% identity	Overlap	a.a.	Score	Ligands	Protein name
1. 1ubf(E)		254	71.5%	242	1418.2			Localization of the small subunit ribosomal proteins into a cryo-em map of <i>Saccharomyces cerevisiae</i> translating 80S rib
2. 1ubf(E)		283	81.5%	262	1358.9			Localization of the small subunit ribosomal proteins into a cryo-em map of <i>Trichum aestivum</i> translating 80S ribosome
3. 1ubf(R)		226	73.0%	222	1345.8	SF4 ANP		Structure of the 40S abo1 post-splicing complex in ribosome recycling and translation initiation
4. 1ubf(C)		217	75.4%	211	1339.7			Cryo-em structure of 40S-eIF1a-eIF1 complex from yeast
5. 1ubf(C)		217	75.4%	211	1339.7			Cryo-em structure of 40S-eIF1-eIF1a preinitiation complex
6. 1ubf(C)		217	75.4%	211	1339.7			Structure of the yeast <i>Kluyveromyces fragilis</i> small ribosomal subunit in complex with the cricket paralysis virus Ires
7. 1ubf(C)		217	75.4%	211	1339.7	MET		Cryo-em structure of a partial yeast 40S preinitiation complex
8. 1ubf(R)		220	73.7%	220	1331.1			Cryo-em structure of a late pre-40S ribosomal subunit from <i>Saccharomyces cerevisiae</i>
9. 1ubf(B)	X-ray 4.00Å	219	72.9%	218	1327.5	CHX		Yeast 80S ribosome. This entry consists of the 40S subunit o first 80s in the asymmetric unit.
10. 1ubf(B)	X-ray 4.00Å	219	72.9%	218	1327.5	CHX		Yeast 80S ribosome. This entry consists of the 40S subunit o second 80s in the asymmetric unit.
11. 1ubf(D)	X-ray 2.80Å	217	74.4%	211	1323.9	CHX		Crystal structure of lactimidomycin bound to the yeast 80S r
12. 1ubf(G)	X-ray 2.80Å	217	74.4%	211	1323.9	CHX		Crystal structure of lactimidomycin bound to the yeast 80S r
13. 1ubf(D)	X-ray 2.90Å	217	74.4%	211	1323.9	CHX		Crystal structure of cycloheximide bound to the yeast 80S r
14. 1ubf(D)	X-ray 2.90Å	217	74.4%	211	1323.9	CHX		Crystal structure of cycloheximide bound to the yeast 80S r
15. 1ubf(C)	X-ray 3.00Å	217	74.4%	211	1323.9			The structure of the eukaryotic ribosome at 3.0 Å resolution entry contains proteins of the 40S subunit, ribosome a
16. 1ubf(C)	X-ray 3.00Å	217	74.4%	211	1323.9			The structure of the eukaryotic ribosome at 3.0 Å resolution entry contains proteins of the 40S subunit, ribosome b
17. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of lycorine bound to the yeast 80S ribosome
18. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of lycorine bound to the yeast 80S ribosome
19. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of anisomycin bound to the yeast 80S ribosome
20. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of anisomycin bound to the yeast 80S ribosome
21. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of homoharringtonine bound to the yeast 80S
22. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of homoharringtonine bound to the yeast 80S
23. 1ubf(D)	X-ray 3.00Å	217	74.4%	211	1323.9	CHX		Crystal structure of naglactone E C bound to the yeast 80S r

### Molecular Docking:

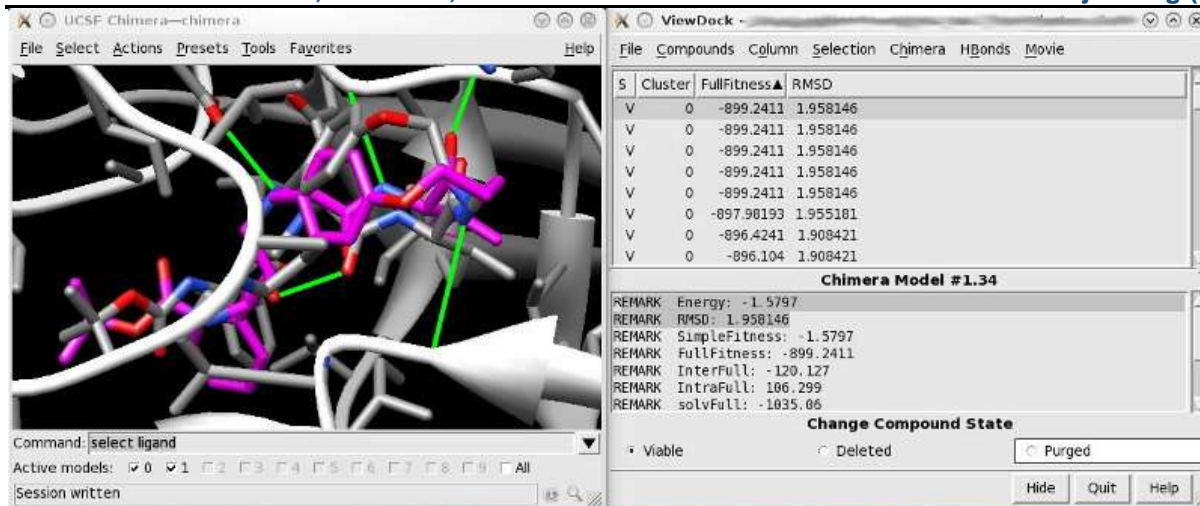
Molecular docking was done by using online free software for docking Swiss Dock. This website provides an access to **SwissDock**, a web service to predict the molecular interactions that may occur between a target protein and a small molecule. **S3DB**, a database of manually curated target and ligand structures, inspired by the [Ligand-Protein Database](#). SwissDock is based on the docking software [EADock DSS](#), whose algorithm consists of the following steps:

1. Many binding modes are generated either in a box (local docking) or in the vicinity of all target cavities (blind docking).
2. Simultaneously, their [CHARMM](#) energies are estimated on a grid.
3. The binding modes with the most favorable energies are evaluated with [FACTS](#), and clustered.
4. The most favorable clusters can be visualized online and downloaded on your computer.



Target for Docking Crystal structure of Cladosporin.-4YCU

Show	Cluster	Element	Fairness (kcal/mol)	ΔG (kcal/mol)
●	0	3	-2567.73	-4.82
●	0	1	-2567.73	-4.82
●	0	2	-2567.73	-4.82
●	0	3	-2567.73	-4.82
●	0	4	-2567.72	-4.82
●	0	5	-2567.73	-4.82
●	0	6	-2567.73	-4.82
●	0	7	-2567.73	-4.82
●	0	8	-2567.73	-4.82
●	0	9	-2567.73	-4.82
●	0	10	-2567.73	-4.82
●	0	11	-2567.73	-4.82
●	0	12	-2567.72	-4.82
●	0	13	-2567.72	-4.82
●	0	14	-2567.72	-4.82
●	0	15	-2567.72	-4.82
●	0	16	-2567.72	-4.82
●	0	17	-2567.72	-4.82
●	0	18	-2567.72	-4.81
●	0	19	-2567.71	-4.81
●	0	20	-2567.71	-4.81
●	0	31	-2567.68	-4.81



<http://www.swissdock.ch/img/material/viewdock.jpg> Image address.

## Conclusion

The Cladosporin (Rps5) protein is involved in the transcription of virus. The present study we analyzed the physicochemical properties of protein by using ProtParam tool. The 3D structure of protein was predicted using SWISS MODEL server. The final model was further evaluated by using Procheck and Ramachandran plot analysis. Binding site of the protein was studied using PDBsum database. From the present study it has been concluded that ribosomal protein s5 protein can be used as target for the inhibition of virus. The molecular structural insight encompasses to the development of new drug for inhibition of protein by using Cladosporin.

## References:

- [1] Cutler SJ, Cutler HG. (1999) Biologically active natural products: pharmaceuticals. CRC Press, New York, USA, p5.
- [2] Scott PM, van Walbeek W. (1971) Cladosporin a new antifungal metabolites from Cladosporin cladosporioids. The Journal of Antibiotics, XXIV, 747-755.
- [3] Grove JF. (1972) New metabolic products of *Aspergillus flavus*. Part I Asperentin its methyl ethers and 5-hydroxyasperentin. Journal of the Chemical Society, Perkin Transactions 1, 19, 2400-2406.
- [4] Springer JP, Cutler HG, Crumley FG, Cox RH, Davis EE, Thean JE. (1981) Plant growth regulatory effects and stereochemistry of cladosporin. Journal of Agricultural Food Chemistry, 29, 853-855.
- [5] Ellestad GA, Miranda P, Kunstmann MP. (1973) Structure of the metabolite LL-S490p from an unidentified *Aspergillus* species. Journal of Organic Chemistry, 58, 4204-4205.
- [6] John M, Krohn K, Florke U, Hans-Jurgen A, Draeger S, Schulz B. (1999) Biologically active secondary metabolites from fungi. 12.1 Oidiolactones A-F, labdane diterpene derivatives isolated from *Oidiodendron truncata*. Journal of Natural Products, 62, 1218-1221.
- [7] Wang S, Li X-M, Teuscher F, Li DL, Diesel A, Ebel R, Proksch P, Wang BG. (2006) Chaetopyranin, a benzaldehyde derivative, and other related metabolites from *Chaetomium globosum*, an endophytic fungus derived from the marine red alga *Polysiphonia urceolata*. Journal of Natural Products, 69, 1622-1625.
- [8] Flewelling AJ, Johnson JA, Gray CA. (2013) Antimicrobials from the marine algal endophyte *Penicillium sp.* Natural Product Communications, 8, 373-374.
- [9] Podojil M, Sedmera P, Vokoun J, Betina V, Baráthová H, Ďuračková Z, Horáková K, Nemeč P. (1978) Eurotium (*Aspergillus*) repens metabolites and their biological activity. Folia Microbiology, 23, 438-443.
- [10] Hoepfner D, McNamara CW, Lim CS, Studer C, Riedl R, Aust T, McCormack SL, Plouffe DM, Meister S, Schuierer S, Plikat U, Hartmann N, Staedtler F, Cotesta S, Schmitt EK, Petersen F, Supek F, Glynn RJ, Tallarico JA, Porter JA, Fishman MC, Bodenreider C, Diagana TT, Movva NR, Winzeler EA. (2012) Selective and specific inhibition of the *Plasmodium falciparum* lysyl-tRNA synthetase by the fungal secondary metabolite cladosporin. Cell Host & Microbe, 11, 654-63.
- [11] Bensch K, Groenewald JZ, Dijksterhuis J, Starink-Willemsse M, Andersen B, Summerell BA, Shin HD, Dugan FM, Schroers HJ, Braun U, Crous PW. (2010) Species and ecological diversity within the *Cladosporium cladosporioides* complex (Davidiellaceae, Apnodiales). Studies in Mycology, 67, 1-94.
- [12] Ellis MB. (1971) Dematiaceous Hyphomycetes. Commonwealth Mycological Institute.
- [13] Rahalison L, Hamburger M, Hostettmann K, Monod M, Frenk E. (1991) A bioautographic agar overlay method for the detection of antifungal compounds from higher plants. Phytochemical Analysis, 2, 199-203.
- [14] Danelutte AP, Lago JH, Young MC, Kato MJ. (2003) Antifungal flavanones and prenylated hydroquinones from *Piper crassinervium* Kunth. Phytochemistry, 64, 555-559.
- [15] Lago JH, Ramos CS, Casanova DC, Morandim Ade A, Bergamo D C, Cavalheiro AJ, Bolzani Vda S, Furlan M, Guimarães EF, Young MC, Kato MJ. (2004) Benzoic acid derivatives from *Piper* species and their fungitoxic activity against *Cladosporium cladosporioides* and *C. sphaerospermum*. Journal of Natural Products, 67, 1783-1788.

- [16] Benevides PJC, Young MCM, Giesbrecht AM, Roque NF, Bolzani VS. (2001) Antifungal polysulphides from *Petiveria alliacea* L. *Phytochemistry*, 57, 743-747.
- [17] Vieira MR, Milheiro A, Pacheco FA. (2001) Phaeohyphomycosis due to *Cladosporium cladosporioides*. *Medical Mycology*, 39, 135-137.
- [18] Kwon-Chung KJ, Schwartz IS, Rybak BJ. (1975) A pulmonary fungus ball produced by *Cladosporium cladosporioides*. *American Journal of Clinical Pathology*, 64, 564-568.
- [19] Grove JF, Pople M. (1981) The insecticidal activity of some fungal dihydroisocoumarins. *Mycopathologia*, 76, 65-67.
- [20] Anke H, Zaehner H, Koenig W. (1978) Metabolic products of microorganisms 170: On the antibiotic activity of cladosporin. *Archives of Microbiology*, 116, 253-257. 1600 Natural Product Communications Vol. 11 (10) 2016 Wang et al.
- [21] Sakagami Y, Sano A, Hara O, Mikawa T, Marumo S. (1995) Cladosporol,  $\beta$ -1,3-glucan biosynthesis inhibitor isolated from fungus, *Cladosporium cladosporioides*. *Tetrahedron Letters*, 36, 1469-1472.
- [22] Kobayashi E, Ando K, Nakano H, Iida T, Ohno, H, Morimoto M, Tamaoki T. (1989) Calphostins (UCN-1028), novel and specific inhibitors of protein kinase C. I. Fermentation, isolation, physico-chemical properties and biological activities. *The Journal of Antibiotics*, 42, 1470-1474.
- [23] Hirota A, Isogai A, Sakai H. (1981) Structure of cladospolide A, a novel macrolide from *Cladosporium fulvum*. *Agricultural and Biological Chemistry*, 45, 799-800.
- [24] Hirota A, Sakai H, Isogai A. (1985) New plant growth regulators, cladospolide A and B, macrolides produced by *Cladosporium cladosporioides*. *Agricultural and Biological Chemistry*, 49, 731-735.
- [25] Wang XN, Radwan MM, Tarawneh AH, Gao JT, Wedge DE, Rosa LH, Cutler HG, Cutler SJ. (2013) Antifungal activity against plant pathogens of metabolites from the endophytic fungus *Cladosporium cladosporioides*. *Journal of Agricultural and Food Chemistry*, 61, 4551-4555
- [26] Reese PB, Rawlings BJ, Ramer SE, Vederas JC. (1988) Comparison of stereochemistry of fatty acid and cladosporin biosynthesis in *Cladosporium cladosporioides* using 2H decoupled <sup>1</sup>H, <sup>13</sup>C NMR shift correlation. *Journal of the American Chemical Society*, 110, 316-318.
- [27] Rawlings BJ, Reese PB, Ramer SE, Vederas JC. (1989) Comparison of fatty acid and polyketide biosynthesis stereochemistry of cladosporin and oleic acid formation in *Cladosporium cladosporioides*. *Journal of the American Chemical Society*, 111, 3382-3390.
- [28] Hranueli D. (2001) Molecular biology of polyketide biosynthesis. *Food Technology and Biotechnology*, 39, 203-213.
- [29] Cattel L, Grove JF, Shaw D. (1973) New metabolites products of *Aspergillus flavus*. Part III. biosynthesis of asperentin. *Journal of the Chemical Society, Perkin Transactions 1*, 2626-2629.
- [30] Cochrane RVK, Sanichar R, Lambkin GR, Reiz B, Xu W, Tang Y, Vederas JC. (2016) Production of new cladosporin analogues by reconstitution of the polyketide synthases responsible for the biosynthesis of this antimalarial agent. *Angewandte Chemie International Edition*, 55, 664-668.

