

FT-IR analysis of methanol extract of leaves of *Mollugo lotoids*

Dr.M.Chandran,
Associate Professor
Department of Zoology
Thiruvalluvar University,
Serkadu,Vellore-632 115.

Abstract

Mollugo lotoids are otherwise called as *Glinus lotoids*, This plant is grow well in all types of soil. These plants are growing at ground level and creep on the ground. Seeds of this plant are traditionally used as anthelmintic for the prevalent tapeworm infestation. In the present investigation, the FTIR analysis of methanol leaf extract of these plants has the following bioactive phytochemicals such as - in amino acids, =CH₂ in vinyl compounds, 1,2,4-trisubst benzenes, 1,2,4-trisubst benzenes, CH-O-H cyclic alcohols, *t*-butyl in hydrocarbons, NO₂ in sulfonamides, C-N in primary amides, COO⁻ in carboxylic acid salts, NH₂ in aminoacids, C≡C in alkynes (monosubst), N≡N in diazonium salts, CH₃ and -CH₂ in Aliphatic compounds and OH in alcohol and phenols in fraction-I. The phytochemicals Cl-C=O in acid chlorides, C-I in indo compounds, Pyridines, C≡C-H in alkynes, - (CH₂)_n- in hydrocarbons, C-Cl in chloro compounds, CH₂ =C in vinylidenes, SO₃H in sulfonic acids, C-O-C in esters and lactones, SO₂ sulfonyl chlorides, C=O in β- diketones, C=O in ketones, C=C=C in allenes, -PH in phosphines, CH₃ and CH₂- in aliphatic compounds and OH in alcohols and phenols in fraction -II.

Key Words: *Mollugo lotoides*, *Glinus lotoides*, FT-IR, bioactive phytochemicals, anthelmintic.

Introduction

About three-quarters of the world population have used traditional medicine for their health care (Gilani and Atta-ur-Rahman, 2005). Plants have been an important part of sophisticated traditional medicine systems for thousands of years (Gurib-Fakim, 2006). Medicinal plants are gaining popularity because of several advantages such as fewer side effects, better patient compliance, relatively low cost and high accessibility as well as high acceptability due to a long history of use (Veramani and Garg, 2002). The Indian subcontinent is a vast repository of medicinal plants that are used in traditional

medical treatments (Ballabh and Chaurasia, 2007).

Recent records reported that 80% of the people living in rural areas use medicinal herbs as primary healthcare system (Sakarkar and Deshmukh, 2011). In India, around 20,000 medicinal plants have been recorded however traditional communities are using only 7,000 - 7,500 plants for curing different diseases (Kamboj, 2000). *Glinus lotoides* (*Mollugo lotoides*) is one of the plant selected for the present study is indigenous to Ethiopia, It is used as anthelmintic (Pankhurst, 1965 and Kloos *et al.*, 1978), antimicrobial (Abdel-Hameed, *et al.*, 2008), antifungal and antitumor medicines (Chopra *et al.*, 1956) used to cure anal fistula, boils and wounds (Hamed *et al.*, 1996).

Materials and method

The fresh leaves of *Mollugo lotoides* collected from moisture agricultural land near vallimalai Murugan Temple, Vellore Tamilnadu were dried under shaded place in the laboratory for complete evaporation of moisture content. Thereafter, these dried leaves were grounded well in a mixer and make into fine powder. This fine powder of leaf was packed inside the thimble of the Soxhlet apparatus. The required amount of methanol was filled inside the bottom flask. The temperature of the solvent was set in the heating mantle. Few minutes after, the green colour extract come from the siphon was collected in a conical flask and dried under sunlight to evaporate the solvent. Then dried phytochemicals containing extract were stored for FT-IR study.

Biology of study plants

The plant *Glinus lotoides* L is locally known as “Mettere” is an annual or short- living perennial prostrate herb. This plant belongs to the family *Molluginaceae* which contains 16 genera and about 100 species.

Description

The plant *Glinus lotoides* is a small annual herb grow with the branches of 20- 50cm long. All parts contain densely stellate-pubescent and short gray hairs. Leaves are barely petiolate or with stellate-hairy, arranged an opposite or whorl type on branches with to 1-3 cm long. The lamina is mostly very widely obovate to obovate, 8–23mm long, 7–14mm wide, margins entire, both dorsal

and ventral surfaces greyish-green color. Flowers are greenish in color with 5mm long and arranged as 3-8 clusters in the axillary region, shortly pedicelled, fascicled, sessile. Sepals are densely hairy in the outside. Stamens are 10-15 in number with some staminodes. Ovary contains 1-1.5mm long 5 styles. Seeds are small, brown or black color and present inside the ovoid capsule. Flowering and fruiting are present throughout years.

Distribution

This plant is native to Eurasia and Africa and mostly abundant in tropical, subtropical and warm temperature area worldwide such as Ethiopia, Sudan, Uganda, Egypt, India, Pakistan, and South America and in all mainland states of Australia.

Classification

Kingdom: Plantae

Order: Caryophyllales

Family: Molluginaceae

Genus: *Glinus*

Species: *G. lotoides*

Phytocompounds

Saponin: The plant *Glinus lotoides* L contains the following phytocompounds such as (1) 3-O- $[\beta$ -D-glucopyranosyl-(1 \rightarrow 4)- α -L-arabinopyranosyl]-oleanolic acid- (28 \rightarrow 1) - β -D-glucopyranosyl ester, (2) 3-O- β -L-arabinopyranosyl -22-O- β -D-glucopyranosyl (4 \leftarrow 1)- α -L-rhamnopyranosyl 1-15 β -hydroxyhopan-6-one (3) 3-O- β -L-arabinopyranosyl-15-O- β -D-glucopyranosyl-22 β -hydroxyhopan-6-one (4) 3-O- β -D-glucopyranosyl-(4 \leftarrow 1)- β -L-arabinopyranosyl-22 β -hydroxyhopan-6-one (5) 3-O- β -L-arabinopyranosyl-22- α -L-rhamnopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-5 β -H-16- β -hydroxyhopane (6) 3-O- β -L-arabinopyranosyl-22-O- β -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-5 β -H-16- β -hydroxyhopane (7) 3-O- β -D-xylopyranosyl -(1 \rightarrow 2)- α -L-rhamnopyranosyl -6 α -O- β -D-xylopyranosyl -22- β -O- β -D-glucopyranosyl-16 β -hydroxyhopane (8) 3-O- β -D-xylopyranosyl -(1 \rightarrow 2)- α -L-rhamnopyranosyl -22- β -O- β -D-glucopyranosyl-6 α ,16 β -dihydroxyhopane (9) 3-O-D-xylopyranosyl-6 α -O- β -D-xylopyranosyl -16 β -O- β -D-xylopyranosyl-22 β -

hydroxyhopane (10) 3-O- β -D-xylopyranosyl-16 β -O- α -L-arabinopyranosyl-6 α ,22- β -dihydroxy hopane (11) 3-O- β -D-xylopyranosyl-6 α -O- β -D-xylopyranosyl-16 β ,22- β -dihydroxyhopane (12)3-O- β -D-xylopyranosyl-22- β -O- β -D-glucopyranosyl-16 β -hydroxyhopan-6-one.

Flavonoids:Apigenin-7-O-glucoside, isovitexin, luteolin-7-O-glucoside, vicenin-2 and vitexin-2''-O-glucoside.

Isoflavonoids:5,7,2',4' - tetrahydroxy-6- (3,3-dimethylallyl) isoflavone; 5,7,4'-trihydroxy- 6,3'-di-(3,3-dimethylallyl) isoflavone and 5,7,2',4'-tetrahydroxy-6; 3'-di-(3,3- dimethylallyl) isoflavone.

Others: β -amyrin, campesterol, α -spinasterol, β -sitosterol and lupeol

Medicinal uses

Glinus lotoides are traditionally used as anthelmintic to prevent tapeworm infection. In Punjab region this plant is used to cure the diarrhoea, applied to itches and skin diseases by traditional native practitioners.

Fig -1 Plant *Mollugo lotoides* with leaves and flowers.



Results and discussion

Glinus lotoidesis are mostly distributed in tropical and subtropical region such as Ethiopia, Sudan, Uganda, Egypt, India, Pakistan and South Africa. In Ethiopia, the seeds of *Glinus lotoides* are traditionally used as anthelmintic for the prevalent tapeworm infestation (Djote, 1978; Endale et al., 1997, 1998), taenicidal activity ((Djote, 1978). Hence, in the present investigation, the FT-IR analysis made on the methanol extract of leaf of *Glinus lotoidesis* showed the following peak values such as 559.36, 640.37, 823.6, 1064.71, 1240.23, 1357.89, 1415.75, 1598.99, 1610.56, 2133.27, 2351.23, 2935.66, 3402.43, 3782.41, 3942.5 and 3986.86 in fraction-I indicates the existence of bands such as C-C-O- rocking, =CH₂ twistings, CH out-of-plane deformation, C-O stretch, Skeletal vibration, NO₂ antisym stretch, C-N Stretch (Amide III band), COO⁻ in antisym stretch, NH₂ Deformation-broad band, C≡C Stretch, N≡N in stretch- aq soln, CH antisym and sym stretching, -OH stretch (solids and liquids) -OH stretch (solids and liquids and -OH stretch (solids and liquids) and the functional groups such as - in amino acids, =CH₂ in vinyl compounds, 1,2,4-trisubst benzenes CH-O-H cyclic alcohols, *t*-butyl in hydrocarbons, NO₂ in sulfonamides, C-N in primary amides, COO⁻ in carboxylic acid salts, NH₂ in aminoacids, C≡C in alkynes (monosubst), N≡N in diazaonium salts, CH₃ and -CH₂ in Aliphatic compounds, OH in alcohol and phenols, OH in alcohol and phenols, OH in alcohol and phenols and OH in alcohol and phenols respectively. The peak values such as 563.21, 644.22, 723.31, 850.61, 1068.56, 1255.66, 1381.03, 4440.83, 1631.78, 1705.07, 2357.01, 2798.7, 2864.29, 2926.01, 3367.71, 3687.9, 378.41, 3842.2, 3861.49 and 3944.43 in fraction -II indicates the existence of Chain deformation (two bands), O-C-O bend, CH₂ rocking in methylene chains; intensity depends on chain length, Si-C in organosilicon compounds, SO₃H Sym stretch, C-O-C antisym stretch, SO₂ antisym stretch, CH₃ antisym deformation, C=O Stretch and enol form, C=O Stretch and fairly broad, -PH Stretch- sharp peak, -CH₃ Stretching modes, CH₃ and -CH₂ antisym and sym stretching, CH₃ and -CH₂ antisym and sym stretching, -OH Stretch (solids and liquids), OH Stretch (solids and liquids), OH Stretch (solids and liquids), OH Stretch (solids and liquids), OH Stretch (solids and liquids) and OH Stretch (solids and liquids) and the presence of bands related functional groups such as C_nH_{2n+1} in alkyl

groups O-C-O in esters, $-(CH_2)_n-$ in hydrocarbons, Si-C stretch, SO_3H Sulfonic acids, C-O-C esters, lactones, SO_2 Sulfonyl chlorides, CH_3 Aliphatic compounds, C=O in β diketones, C=O Carboxylic acids, -PH in phosphines, $-CH_3$ attached to O or N, CH_3 and $-CH_2$ in aliphatic compounds, CH_3 and $-CH_2$ in aliphatic compounds, -OH in alcohol and phenols, -OH in alcohol and phenols, -OH in alcohol and phenols, -OH in alcohol and phenols, -OH in alcohol and phenols, and -OH in alcohol and phenols respectively. These results showed an agreement with the seeds of *Glinus lotoides* contain 10% crude saponin (Endale et al., 1998) and 14% oil (Biftu et al., 1979). The lipophilic compounds in the plant comprise fatty acids, glycosides of sitosterols, stigmasterol, flavonoids and waxes (Biftu et al., 1979) and five triterpenoidal saponines (Glinusides A, B, C, D and E) have been isolated from the n-butanol fraction of *Glinus lotoides* var. *dicatamnoides* (Hamed and El-Elmary, 1999).

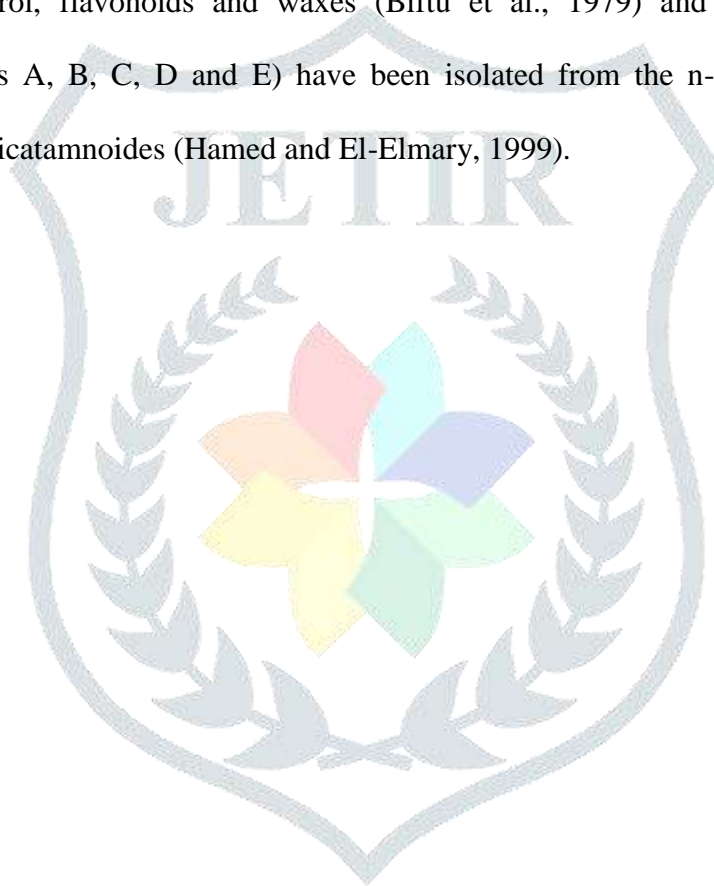
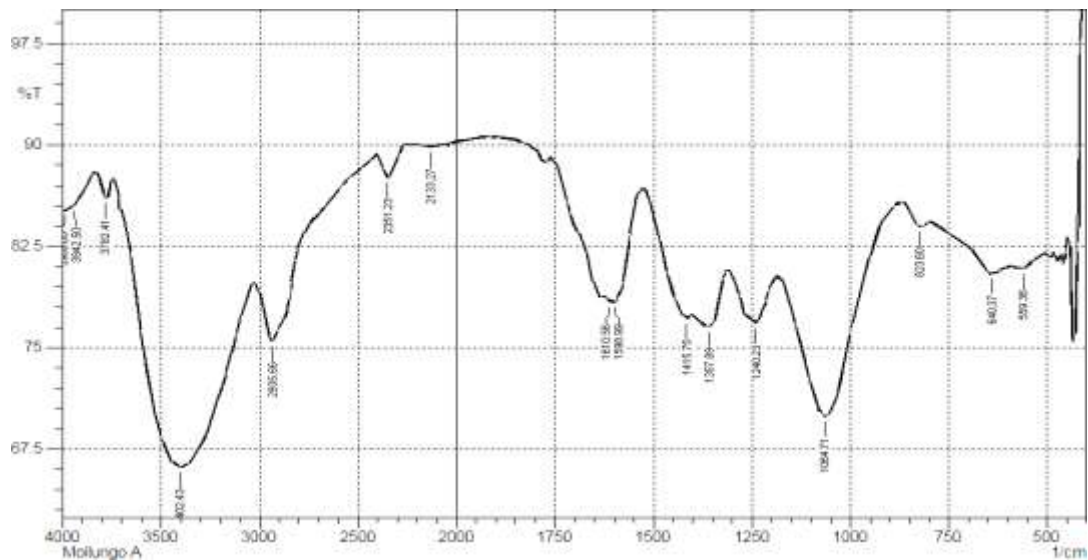
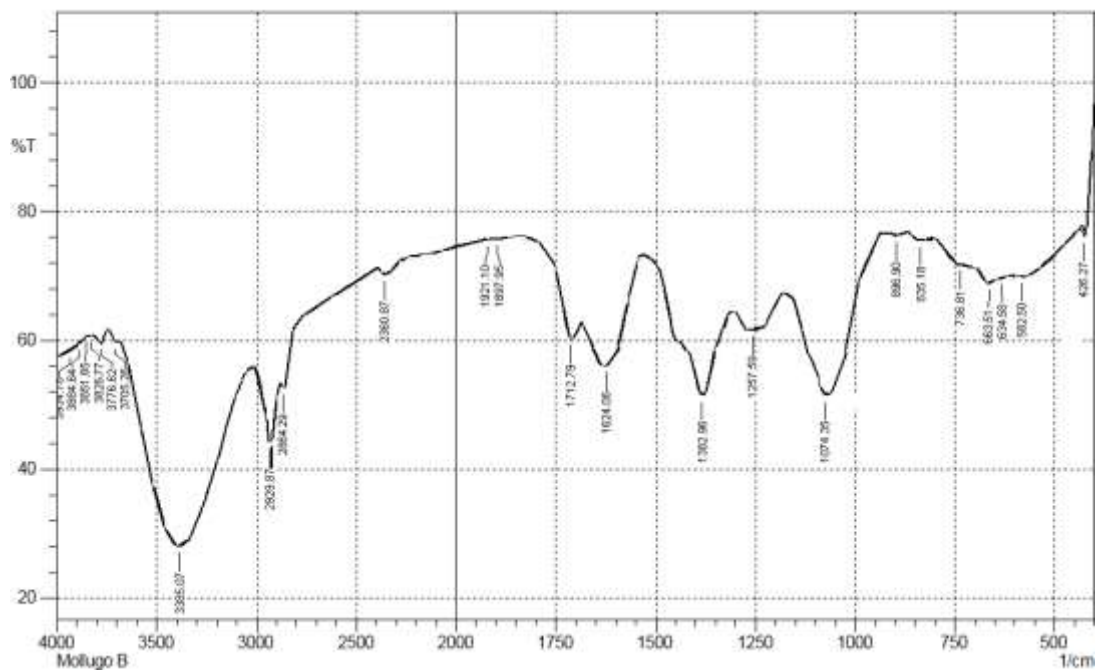


Table-1. Peak values, band type and functional group for FTIR (Fourier Transform Infrared Spectroscopy) spectra of fraction I methanol extract of *Mollugo lotoid*

No	Peak value	Bonds	Functional group
1.	559.36	- rocking	- in amino acids
2.	640.37	=CH ₂ twistings	=CH ₂ in vinyl compounds
3.	823.6	CH out- of- plane deformation	1,2,4-trisubst benzenes
4.	1064.71	C-O stretch	CH-O-H cyclic alcohols
5.	1240.23	Skeletal vibration	<i>t</i> -butyl in hydrocarbons
6.	1357.89	NO ₂ antisym stretch	NO ₂ in sulfonamides
7.	1415.75	C-N Stretch (Amide III band)	C-N in primary amides
8.	1598.99	COO ⁻ in antisym stretch	COO ⁻ in carboxylic acid salts
9.	1610.56	NH ₂ Deformation;broad band	NH ₂ in aminoacids
10.	2133.27	C≡C Stretch	C≡C in alkynes (monosubst)
11.	2351.23	N≡N in stretch; aq soln	N≡N in diazaonium salts
12.	2935.66	CH antisym and sym stretching	CH ₃ and -CH ₂ in Aliphatic compounds
13.	3402.43	-OH stretch (solids and liquids)	OH in alcohol and phenols
14.	3782.41	-OH stretch (solids and liquids)	OH in alcohol and phenols
15.	3942.5	-OH stretch (solids and liquids)	OH in alcohol and phenols
16.	3986.86		OH in alcohol and phenols

Table-2. Peak values, band type and functional group for FTIR (Fourier Transform Infrared Spectroscopy) spectra of fraction II methanol extract of *Mollugo lotoid*

No	Peak value	Bonds	Functional group
1.	426.27	Cl-C=O plane deformation	Cl-C=O in acid chlorides
2.	582.5	C-I Stretch	C-I in indo compounds
3.	634.58	plane ring deformation	Pyridines
4.	663.51	C≡C-H bending	C≡C-H in alkynes
5.	736.81	CH ₂ rocking in methylene chains; intensity depends on chain length.	-(CH ₂) _n - in hydrocarbons
6.	835.18	C-Cl stretch	C-Cl in chloro compounds
7.	896.9	CH ₂ out-of-plane	CH ₂ =C in vinylidenes
8.	1074.35	SO ₃ H sym stretch	SO ₃ H in sulfonic acids
9.	1257.59	C-O-C Antisym stretch	C-O-C in esters and lactones
10.	1382.96	SO ₂ Antisym stretch	SO ₂ sulfonyl chlorides
11.	1624.06	C=O stretch and enol form	C=O in β- diketones
12.	1712.79	C=O stretch ; 20cm ⁻¹ lower if unsaturated	C=O in ketones
13.	1897.95	C=C=C antisym stretch	C=C=C in allenes
14.	1921.1	C=C=C antisym stretch	C=C=C in allenes
15.	2360.87	-PH stretch and sharp peak	-PH in phosphines
16.	2864.29	CH antisym and sym stretching	CH ₃ and CH ₂ - in aliphatic compounds
17.	2929.87	CH antisym and sym stretching	CH ₃ and CH ₂ - in aliphatic compounds
18.	3385.07	-OH Strtech (Solids and Liquids)	OH in alcohols and phenols
19.	3705.26	-OH Strtech (Solids and Liquids)	OH in alcohols and phenols
20.	3776.62	-OH Strtech (Solids and Liquids)	OH in alcohols and phenols
21.	3826.77	-OH Strtech (Solids and Liquids)	OH in alcohols and phenols
22.	3851.85	OH Strtech (Solids and Liquids)	OH in alcohols and phenols

Fig-2.FTIR Pattern of fraction –I methanol extract of leaf of *Mollugo lotoides*Fig -3.FTIR Pattern of fraction –II methanol extract of leaf of *Mollugo lotoides*

References

- Ballabh ,B and Chaurasia, O.P.2007. Traditional medicinal plants of cold desert Ladakh-used in treatment of cold, cough and fever. *Journal of Ethnopharmacolog* ,**112**:341-349.
- Biftu, T., Abegaz, B., Teffera, S., 1979. Fatty acid composition of “mettere” (*Glinus lotoides* Linne) seeds by gas liquid chromatography (GLC). *SINET: Ethiopian Journal of Science* 2, 19–22.
- Chopra.1956.A review of Medicinal plants from India .*Indian .J.Pharmacol*, **8**(5):416-420.
- Djote, M., 1978. Taenicidal activity of *Glinus lotoides* (Aizoaceae). *Journal of Ethiopian Pharmaceutical*

- Association 3, 9–11.
- Endale, A., Getachew, M., Gebre-Mariam, T., 1997. In vitro taenicidal activity of the seeds of *Glinus lotoides* on *Hymenolepis nana* worms. *Ethiopian Pharmaceutical Journal* 15, 46–51.
- Endale, A., Kassa, M., Gebre-Mariam, T., 1998. In vivo anthelmintic activity of the extracts of *Glinus lotoides* in albino mice infested with *Hymenolepis nana* worms. *Ethiopian Pharmaceutical Journal* 16, 34–41.
- Gilani, A and Atta-ur-Rahman.2005. Trends in ethnopharmacology. *Journal of Ethnopharmacology*,**100** :43–49.
- Gurib-Fakim, A.2006. Medicinal plants: traditions of yesterday and drugs of tomorrow.*Molecular Aspects of Medicine* ,**27**: 1–93.
- Hamed, A.I., El-Elmary, N.A., 1999. Triterpenoidal saponins from *Glinus lotoides* Var *dictamnoides*. *Phytochemistry* 50, 477–480.
- Hamed,A.I., Springuel,I., El-Emary,N.A., Mitome,H., Miyaoka, Hand Yamada,Y. 1996.Triterpenoidal saponin glycosides from *Glinus lotoides*. *Phytochemistry* ,**43** : 183 –188.
- Kamboj ,V.P. 2000. Herbal medicine. *Current Science* ,78:35-38.
- Kloos,H.,Tekle,A.,Yohannes, L and Yosef,A.1978.Preliminary studies of Traditional Medicinal plants in nineteen markets in Ethiopia: use patterns and public health aspects – *Ethiopian medical Journal* ,**16**:33-43.
- Pankhurst,R. 1965.An historical examination of traditional Ethiopian Medicine and surgery . *Ethiopian medical Journal* , **3**:157-172.
- Sakarkar, D and Deshmukh,V. 2011. Ethnopharmacological review of traditional medicinal plants for anticancer activity. *International Journal of Pharma Tech Research* ,**3**:298- 308.
- Veramani, K and Garg,S.2002. Herbal medicines for sexually transmitted diseases and AIDS. *Journal of Ethnopharmacology* , **80**:46–66.