Toxicity of Endosulfan on Total Lipid and Protein of Liver, Kidney and Gonads of Fish Channa punctatus (Bloch.)

Dr. Mohan Kumar Department of Zoology, LNM University, Darbhanga.

Abstract

The current study includes the toxicity effect Endosulfan on fish. The alterations induced by chronic (30 days) exposure of the fish Channa punctatus to a sublethal concentrations (0.01 ppm conc.) of Endosulfan on the profile of total protein and lipid in the liver, kidney and gonads. The liver and kidney showed significant depletion of total protein content amounting. Estimation of total lipid was recorded significantly. The present study therefore points towards a severe metabolic dysfunction in response to Endosulfan toxicity in the fish Channa punctatus (Bloch.)

Keywords: Endosulfan, Channa punctatus, Toxicity, Protein contents, Lipid contents.

INTRODUCTION

Organochlorine insecticides have strong insecticidal properties and broad applications due to their low cost of large scale production. However, some organochlorine pesticides have been banned due to their persistent residual characteristics and unexpected toxicities to non-target organisms in the environment, (Jonsson, et.al., 1993). While much less persistent than other organochlorines, endosulfan is known to be neurotoxic and highly toxic to fish (Sharma, et. al.2011). In addition, this compound at the concentration of 1 µg L⁻¹ has been shown to be genotoxic to fish, affecting reproduction (Dutta et.al., 2006). Endosulfan is one of 123 pesticides that have been listed as potential endocrine disruptors (Mckinlay et.al., 1993). Even though endosulfan is highly toxic to aquatic life, it is still being used in many countries. Therefore, methods for the fast and low cost assessment of the adverse effects of endosulfan in aquatic ecosystems need to be developed. Accurate and precise techniques have elucidated the unknown toxic effects of compounds, including endosulfan (Park DS, et.al., 2015). Its toxicity to fish has been reported at LC₅₀ values lower than those of terrestrial animals (Chow *et.al.*, 2013).

The fish, Channa punctatus (Bloch), locally known as "Garai", having the presence of suprabranchial accessory respiratory organs, an air-breathing teleost and endosulfan were selected for present study.

MATERIALS & METHODS

The air-breathing teleost *Channa punctatus* procured live from the local fish market were washed with 0.1% KMnO₄ solution to remove dermal infection if any. Healthy fish of average length (9–12cm) and weight (21–25 g) were acclimated for 15 days to laboratory conditions. The fish were fed with chopped goat liver every day adlibitum. Running tap water was used in all the experiments and the fish were adjusted to natural photoperiod and ambient temperature. No aeration was done.

Static acute bioassays were performed to determine LC₅₀ values of endosulfan for 24, 48, 72 and 96 hours following the methods of APHA, AWWA & WPCF (1985). The LC₅₀ values for these periods were 8.25 ppm, 6.25 ppm, 4.25 ppm and 3.25 ppm respectively. The sub-lethal concentration was determined following the formula of Hart et al. (1945). Twenty acclimated fish were exposed to a sub-lethal concentration (0.01 ppm) of endosulfan for 30 days. Side by side same number of fish as that of experimental one was maintained as the control group. At the end of exposure period the fish were anaesthetized with 1:4000 MS 222 (tricane, methane, sulfonate, sandoz) for two minutes. The liver, kidney, testis and ovary were quickly dissected out, weighed to nearest mg and processed for the quantitative estimation of total protein by the methods of Varley et al. (1980). The total lipid extraction was done by done by the method Folch et. al. (1957).

RESULTS

The protein profiles of liver, muscle, testis and ovary in response to endosulfan exposure showed a significant decline. The liver and kidney showed statistically more significant decline. The liver and kidney showed statistically more significant (P < 0.001) decline i.e. 31% in , while 30% in liver. The testis showed significant (P < 0.05) while ovary showed significant at (P < 0.01). The testis showed decline 19% while ovary 17%. Total protein in the control liver, kidney, testis and ovary was estimated to be 102.19±1.81, 75.006±1.04, 80.48±1.41, and 121.01±1.89 respectively. As against there, the total protein profiles in the experimental lots were 50.08±1.96 45.08 ± 0.01 , 60.16 ± 0.98 and 70.08 ± 0.01 respectively (Table-I).

TABLE - I

Profiles of total protein (mg/g wet tissue) in tissue of Channa punctatus chronically exposed to endosulfan for 30 days. Values are mean \pm SE 0f 5 observations.

| Tissue | Control | Endosulfan treated |
|--------|-------------|--------------------|
| Liver | 102.19±1.81 | 50.08±1.96 |
| Kedney | 75.006±1.04 | 45.08±0.01 |
| Testis | 80.48±1.41 | 60.08±0.01 |
| Ovary | 121.01±1.89 | 70.08±0.01 |

Value are mean \pm SE of 5 observations, Significant level =P<0.05

The estimation of total lipid was recorded significantly decreases under the experimental concentration and duration of endosulfan exposure, and was to be 70.08±0.01 mg/g in liver, 18.49±0.83 mg/g in kidney, 12.54±0.77 mg/g in testis and 16.05 ± 0.66 overy.

TABLE - II Profiles of total lipid (mg/g wet tissue) in tissue of Channa punctatus chronically exposed to endosulfan for 30 days. Values are mean \pm SE 0f 5 observations.

| Tissue | Control | Endosulfan treated |
|--------|------------|--------------------|
| Liver | 28.10±1.56 | 15.40±.56 |
| Kedney | 20.68±1.17 | 18.49±0.83 |
| Testis | 16.74±0.72 | 12.54±0.77 |
| Ovary | 21.19±0.97 | 16.05±0.66 |

Value are mean \pm SE of 5 observations, Significant level =P<0.05

DISCUSSION

The level of tissue protein and lipid in control fish recorded in the present study indicates that total proteins and lipids are the largest contributors to the wet weight of the tissues after water. Previous workers have also reported decline in tissue protein profiles in a number of fish species exposed to various pesticides and endosulfan. Ramalingam and Ramalingam (1982) noted a steady decline in the total protein of liver and muscle after 7 and 15 days exposure of the fish, Sarotherodon mossambicus to malathion and mercury and correlated it with an intensive proteolysis. Similarly, a significant decrease in the protein content was recorded by Kumar and Ansari (1984) in the Zebra fish, Brachydanio rerio, exposed to malathion and suggested inhibition of protein synthesis by the toxicant. Proteins being involved in the architecture and physiology of the cell seem to occupy a key role in the cell metabolism. The observed significant depletion of tissue protein in the present case denotes high catabolic potency of those organs and may be attributed to the intensive proteolysis and utilization of their degradation products for metabolism under the toxic influence of Herbiclon. They might have been fed into TCA cycle through aminotransferase system to cope with the excess demand of energy during stressful situations as suggested by Jha (1991). The loss of gonadal proteins may also be associated to the direct action of pyrethroids leading to arrest of vitellogenesis in ovary and loss of germ cells in testis (Jha and Jha, 1995). Moreover, the decreased protein contents might also be attributed to the tissue destruction, necrosis, or disturbance of cellular function and consequent impairment in protein synthetic machinery (Srivastava, et al., 1995). The liver of Clarias gariepinus exposed to the cypermethrin showed hyperplastic hepatic and necrosis of hepatic cells (Andem A. B. et al. 2016). The toxicity was found to increase with endosulfan concentration, various structural changes were already induced

on the morphology of the vital organs, i.e. gill, liver and kidney even with exposure to low, sublethal endosulfan concentration (Nordin, et. al. 2018).

The estimation of total lipid was recorded significantly decreases under the experimental concentration and duration of endosulfan exposure. Previous workers have similar reported decline in total lipid of different tissue profiles in a number of fish species exposed to various pesticides and endosulfan. The decrease might have occurred mainly due to altered lipid metabolism and energy demand in fishes under stress of toxicants.

CONCLUSION

The test fish Channa punctatus when exposed to sub lethal concentration of endosulfan (0.01 ppm) for 30 days, significant decrease in total lipid and total protein content in the tissue of all four organs, liver, kidney, testis and ovary. The decrease might have occurred mainly due to altered lipid and protein metabolism and energy demand in fishes under stress of toxicants.

ACKNOWLEDGEMENT

The authors are thankful to the Department of Zoology, LNM University, Darbhanga, for the provision of laboratory facilities used in this study.

REFERENCES

- Andem A. B. Ibor O. R., Joseph A. P., Eyo V. O., Edet A. A., 2016: Toxicological Evaluation and Histopathological Changes of Synthetic Pyrethroid Pesticide (Cypermethrin) Exposed to African Clariid Mud Catfish (Clarias gariepinus) Fingerlings. International Journal of Toxicological and Pharmacological Research; 8(5); 360-367.
- APHA, 1985: Standard methods for the examination of water and waste water (16th Ed). American Public Health Assoc., Washington D.C.
- Chow WS, Chan WK, Chan KM, 2013: Toxicity assessment and vitellogenin expression in zebrafish (Danio rerio) embryos and larvae acutely exposed to bisphenol A, endosulfan, heptachlor, methoxychlor and tetrabromobisphenol A. J. Appl. Toxicol. 33:670–678.10.1002/jat.v33.7
- Deshmukh, D. R. 2015: Toxicity of endosulfan on protein level in a freshwater fish Wallago attu. J. Trends. Life Sci.Res., 3(2):15-18.
- Dutta HM, Misquitta D, Khan S., 2006: The effects of endosulfan on the testes of bluegill fish, Lepomismacrochirus: a histopathological study. Archives of Environmental Contamination and Toxicology. 51:149-156
- Folch, J., Lees, M and SloanS, G.H., 1957: Asimple method for the isolation and purification of lipids from animal tissues. J. Biol. Chem., 226:497-507.

- Hart, W.B., Dondoroff, P. and Greenbank, J., 1945: The evaluation of toxicity of industrial wastes, chemicals and other substances to freshwater fishes. Atlantic Refining Company. Phil. Part (1): 317-326.
- Jha, B.S., 1991: Alterations in the protein and lipid contents of intestine, liver and gonads in the lead exposed freshwater murrel, *Channa punctatus* (Bloch). J. Ecobiol. 3(1): 29-34.
- Jha, B.S. and Jha, M.M., 1995b: Biochemical effects of nickel chloride on the liver and gonads of the freshwater climbing perch, *Anabas testudineus*, (Bloch). Proc. Nat. Acad. Sci. (India) 65B(1): 39-46.
- Jonsson CM, Toledo MCF, 1993: Bioaccumulation and elimination of endosulfan in the Fish Yellow Tetra (Hyphessobrycon-Bifasciatus). *Bulletin of Environmental Contamination and Toxicology*. 1993;50:572-577
- Kumar, K. and Ansari, B.A., 1984: Malathion toxicity effect on the liver of the fish, *Brachydanic reno* (Cyprinidae). Ecotoxical Environ. Sal. 23: 199-205.
- Mckinlay R, Plant JA, Bell JNB, Voulvoulis N., 2008: Endocrine disrupting pesticides: implications for risk assessment. *Environment International*. 34:168-183
- Nordin1, N. Ibrahim1, S.A. Ahmad, N.l Hamidin1, F.A. Dahalan1, and M.Y. Abd. Shukor, 2018. Endosulfan Toxicity to Anabas testudineus and Histopathological Changes on Vital Organs. E3S Web of Conferences **34**, 02055.
- Park DS, Jeon HJ, Park ES, 2015: Highly selective biomarkers for pesticides developed in *Eisenia fetida* using SELDI-TOF MS. Environ. Toxicol. Pharmacol. 39:635–642.10.1016
- Ramalingam, K. and Ramalingam, K., 1982: Effects of sublethal levels of DDT, malathion and mercury on tissue proteins of *Sarotherodon mossambicus* (Peters). Proc. Ind. Acad. Sci. (Anim. Sci.) 91(6): 501–505.
- Sharma A, Mishra M, Ram KR, Kumar R, Abdin MZ, Chowdhuri DK. 2011: Transcriptome analysis provides insights for understanding the adverse effects of endosulfan in Drosophila melanogaster. *Chemosphere*. 82:370-376
- Sarma K, Pal AK, Grinson-George, 2015: Effect of sub-lethal concentration of endosulfan on lipid and fatty acid metabolism of spotted murrel, *Ghana punctatus*. J. Environ. Biol.36:451–454.
- Srivastava, A.K., Singh, N.N. and Srivastava, A.K., 1995: Bio-chemical change in freshwater. Indian Cat. Fish. Following exposure to sublethal concentration of propoxur J. Freshwater Biol. 7(4) 257-260.
- Varley, H. Gowenlock, A.H. and Bell, M., 1980: Practical clinical Bio-chemistry, Vol. I, General topics and commoner tests. William Heinemann Medical Books Ltd., London.