



Antimicrobial Activities of Marine Actinobacteria *Streptomyces sp.* isolated from Machilipatnam coast area

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Abstract : Actinobacteria of marine origin are considered to be a good and efficient source of secondary metabolites having been developed for clinical use. *Streptomyces* is the largest genus of actinobacteria belonging to the family Streptomycetaceae. It is gram positive with enhanced GC content. *Streptomyces* is synonymous with the production of numerous naturally originated antibiotics having a multitude of uses. It has been established that the coastal wetland inhabiting actinobacteria possess broad spectrum antimicrobial activities. This forms the basis that they are able to generate newer bioactive substances. In the present investigation the antimicrobial activities of marine actinobacteria- *Streptomyces sp.*, namely, *Streptomyces alboniger*, *Streptomyces coelicolor* and *Streptomyces griseus* that were isolated from the Machilipatnam coast area was studied against the clinical bacteria consisting of Gram positive *Staphylococcus aureus*, *Streptococcus faecalis* and Gram negative *Proteus vulgaris* and *Salmonella typhi*.

IndexTerms - Actinobacteria, Streptomyces, antimicrobial, bioactive, metabolites.

I. INTRODUCTION

The most interesting and appreciable feature of *Streptomyces sp.* is the production of secondary metabolites, specifically antibiotics. Streptomycetaceae family is part of the phylum Actinobacteria, the order Actinomycetales and the genus *Streptomyces* is the sole member of the family by Anderson and Wellington, (2001). The studies of Chen *et al.*, (2021) established the coastal wetland inhabiting actinobacteria exhibiting broad spectrum antimicrobial activities in addition to PKS and NRPS genes. Wang *et al.*, (2020) presented the antimicrobial activities of almost 313 novel natural products of marine actinomycetes from 1976 to 2019. Hamed *et al.*, (2019) isolated actinomycetes inhabiting the marine regions of the Red sea and the Suez gulf with antagonistic properties. Phongsopitapan *et al.*, (2019) established the antagonistic properties of nearly 41 marine actinomycetes of Thailand region. The studies of Subramani and Aalbersberg, (2012) imply that Streptomycetes group amongst actinomycetes establish it to be economically important. This genus is accredited with producing more than fifty percent of the known antibiotics.

The studies of Ramesh and Mathivanan, (2009), Peela *et al.*, (2005) established the prevalence of diversified antimicrobial activity and enzyme production in marine originated actinomycetes. This has increased the pronouncement of industrially important marine actinomycetes, the major one being *Streptomyces* from the Bay of Bengal which produced the best enzymes and molecules. The work of Kathiresan *et al.*, (2005) caused the isolation of marine actinomycetes from mangroves and estuaries the strains of which proved to act as agrobased fungicides. The relevant antagonistic properties of 107 marine actinomycetes that was screened of the Konkan coast of Maharashtra was established by Gulve and Deshmukh, (2012). Sivakumar *et al.*, (2011) were able to isolate 78 types of marine actinomycetes from the Bay of Bengal along the Pudimadaka coast of Andhra Pradesh wherein *Streptomyces* was the most prevailing strain exhibiting anti bacterial and anti fungal activity. Jiang *et al.*, (2017) studied chalcones from marine derived *Streptomyces* species and their antimicrobial activities. Cho *et al.*, (2020) discussed the antibacterial activity of chromomycins from the marine derived *Streptomyces microflavus*.

Dharmaraj, (2010) highlighted that within the marine actinobacterial group *Streptomyces* is represented by the greatest number of species and varieties. Patel *et al.*, (2020) observed and studied the antibacterial activity of marine bacterial pigments from the Arabian sea. Elnaby *et al.*, (2016) worked on the properties pertaining to antibacterial and anticancer action of silver nanoparticles that are synthesized extracellularly and obtained from marine *Streptomyces rochei* MHM13. Similar studies by Rana *et al.*, (2018) were based on the antimicrobial potency of silver nanoparticles generated by *Streptomyces sp.* RHS16 against certain fish pathogens. Dholakiya *et al.*, (2017) predicted the antioxidant and antibacterial activities of newer actinobacterial strain *Streptomyces variabilis* RD-5 that was isolated from the gulf of Khambhat Gujarat. Luo *et al.*, (2020) worked on certain natural marine antibacterial products as lobophorin L M, spirotetronate antibiotics and ansamycins that were derivatives of the marine *Streptomyces sp.* 4506. The studies of Janardhan *et al.*, (2014), Jiao *et al.*, (2013) established and emphasized marine mangrove based actinomycetes possessing potent ability to produce bioactive compounds. Many researchers have focused on gifted antimicrobial compounds isolated from marine actinomycetes from 1991-2013. The observations of Behie *et al.* (2017), Kamjam

et al., (2017) have correlated the production of newer bioactive metabolites with actinobacterial adaptations to extreme climatic conditions.

Streptomyces has the ability to produce cellulose with cellulolytic waste materials used in degradation studies characterized from the marine environments. Actinobacteria isolated from marine sediments and also molluscs form the major source for many extracellular enzymes as studied in Vellar estuary by Patil *et al.*, (2001). This inferred the characterization of 104 isolates of which 77 of them were observed to show inhibitory activity against one of the pathogens. *Actinopolyspora sp.* have shown antagonistic activity against the Gram positive bacteria that were isolated from the west coast of India. On the contrary it exhibited no antagonistic activity against the Gram negative bacteria as reported by Kokare *et al.*, (2004). Marine actinobacteria are featured in the bioprospecting of enzymes secreted with plenty of industrial applications by Sahu *et al.*, (2005). *Streptomyces albus* having good scope for industrial production of extracellular L-glutaminase reported by Umamaheswary *et al.*, (2005). Actinobacterial studies imply and spread information about the antibiotic bioactive producing properties that could be utilized for human welfare as discussed and explained by Sahu *et al.*, (2006). *Streptomyces albidoflavus* has shown anti-tumour properties isolated from the Pichavaram mangroves by Siva Kumar *et al.*, (2005). Marine actinobacterial strains to date remain the most superior ones in research by Lakshmipathy *et al.*, (2010). *Streptomyces* isolated from urban soils have shown to be resistant to multiple antibiotics by Zotchev, (2012). Learn-Han *et al.*, (2012) exemplified the marine derived genus of *Streptomyces* having produced 289 secondary metabolites basing on the information provided by marinlit database.

Selvakumar *et al.*, (2012) characterized the marine microorganisms producing vitamin B12 and the remarkable antagonistic activity of glucose utilizing potential of *Streptomyces sp.* *Streptomyces avermitilis* is synonymous with avicta a nematicidal product explored by Cabrera *et al.*, (2012). The studies of Antunes *et al.*, (2014) confirmed 7600 compounds being produced under *Streptomyces species*. Rana and Salam, (2014) isolated and characterized 23 marine actinomycetes with at least 7 bringing in inhibition of both Gram negative and positive strains with a 20 mm inhibitory zone. *Streptomyces* species also involved in the control of certain plant pathogens as reported by Al-Askar *et al.*, (2015). The prospective magnitude of marine microorganisms involved in the making of constructive metabolic products being cytotoxic, antibacterial and antifungal was revealed by Sivalingam *et al.*, (2019). Study and observations by various researchers has proved that marine originated *Streptomyces* being abundantly available is a potential source for novel anti microbial activities although microorganisms of mangrove origin gained not enough recognition as pointed by Liua *et al.*, (2008). Nature has risen as most important source for newer antibiotics by Kpehn and Carter, (2005) wherein alternative selected isolation procedures have been suggested by Sathiyaseelan and Saranraj, (2016). Therefore this present research work laying more emphasis on the isolation of the marine actinobacteria, *Streptomyces sp.* with specific antagonistic properties from the sea and mangrove regions of coastal Machilipatnam.

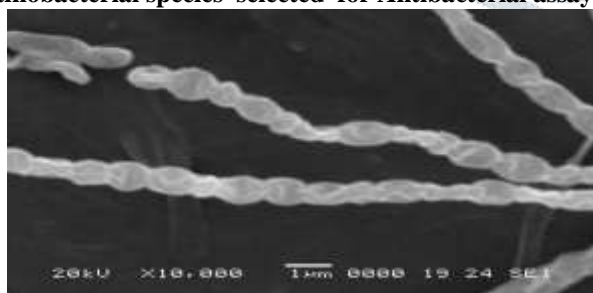
Materials and methods: Antibacterial activity

The antagonistic activities of the thus isolated actinobacteria during the investigative period were evaluated against the clinical bacteria being Gram positive *Staphylococcus aureus* and *Streptococcus faecalis* and Gram negative *Proteus vulgaris* and *Salmonella typhi*.

Assay for antagonistic activity

The often occurring marine actinobacteria, *Streptomyces alboniger*, *Streptomyces coelicolor* and *Streptomyces griseus* that were isolated during the period of investigation were selected to assess their antagonistic activity against the clinical bacteria considered. The test conducted by the cross streak method on modified nutrient agar medium as given by Dholakiya *et al.* (2017). The surface of the modified agar was streaked singly and later left to incubate at $28 \pm 2^\circ\text{C}$ for a period of one week according to Balagurunathan and Subramaniam (2001). The actinobacterial growth was observed and identified as being ribbon like wherein the clinical bacterial streaking at right angles to the actinobacterial streak. They were further subjected to incubation at 28°C . The zone of inhibition measured based on the time frame ranging from 24 to 48 hrs wherein the control plate was devoid of the actinobacterial inoculum in comparison to the usual bacterial growth. The inhibitory zone of > 15 mm is measured as an important activity apprehensive in the combat against a wide range of pathogens.

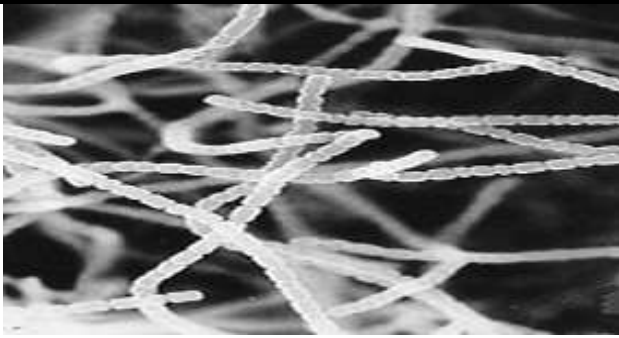
Actinobacterial species selected for Antibacterial assay



Streptomyces alboniger



Streptomyces coelicolor



Streptomyces griseus

Extraction of Antibacterial Principle

The particular opposed actinobacterial isolates infused into a broth of casein agar in a shake flask method with speed being 200-250 rpm. The incubatory duration was 1 week at 28°C. Broth cultures subjected to filtration twice done initially by a supreme quality whatman filter paper and later by the millipore filter procured from millipore millex HV hydrophilic PVDF comprising of size of 0.45µm. The filtrates transferred under sterile conditions into the flask stored at 4°C to proceed with assessment and investigation. This was followed by the slow and individual addition of the solvents alcohol, chloroform, ethyl acetate and methanol to a similar volume of the filtrate. These were mixed well and subjected to a 5000 rpm centrifugation wherein the time period was 10 min by Remya and Vijayakumar, (2008).

Antibiotic disc preparation

The method adopted to study the evaluation of the antibacterial extracts with diverse cultures of bacteria was of Kirby-Bauer disc diffusion. This is also considered and categorized as the antibiotic susceptibility test. Sterilized filter paper discs were used on to which 200 µl of the extract was impregnated and then completely dried. This was done under aseptic conditions.

Antibacterial assay

The assay involved the nutrient agar plates being readied under aseptic conditions. This was followed by impregnating the selected bacteria comprising of 18-24 hours previous culture. The swab method used for spreading the bacterial culture with cotton swabs was adopted. The antibiotic discs thus extracted and impregnated were slowly placed with the pathogenic bacterial swab culture. The incubation time for the plates considered was 1 to 2 days thereby checked the zone of inhibition making a note of the zone of clearance in mm. Among the tested ethyl acetate extracts shown good growth inhibition against the test organisms. Therefore for the assessment and analytical studies TLC and HPLC techniques were done on the ethyl acetate extracts.

Purification of Antibacterial Compounds: Thin Layer Chromatography

Thin layer chromatography with silica gel plates collected from Merck Art. 5735 Kiesselgel 60 F 254 used for partial purification of antibacterial principles to detect the compounds. The dried crude extracts of the selected three actinobacteria dissolved in methanol individually spotted on the silica plate developed in the chromatographic solvent system ratio being 3:1:1 v/v consisting n-butanol-acetic acid-water (BAW). The developed thin layer chromatographic plates air dried overnight to remove any traces of the solvents. Thus separated compounds were visualized under UV at 254 nm absorbance also at 365 nm fluorescence. The active spots were detected by bioautography by Betina (1973), Choma and Grzelak (2011). The retention factors (RF) of the active spots were further calculated.

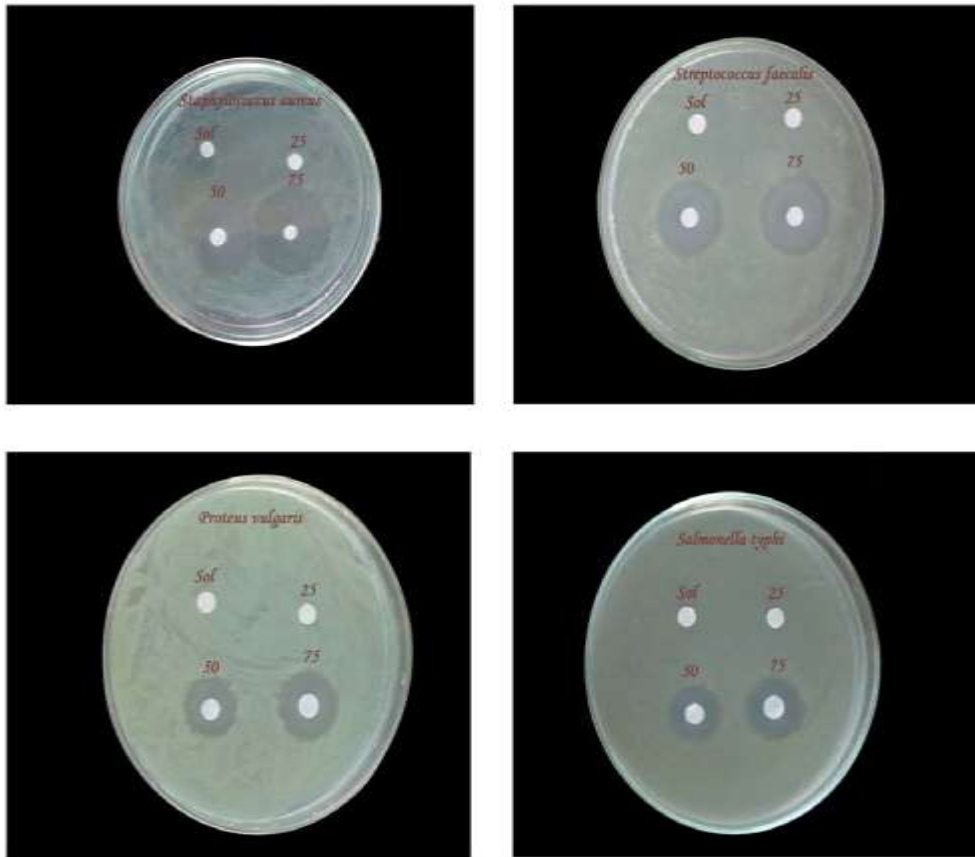
HPLC analysis

The high performance liquid chromatographic (HPLC) technique performed for separation identification of the antimicrobial compound utilizing a Shimadzu HPLC UV-VIS detector and LC-18 column. A 20 µl supernatant injected separately into the HPLC system maintaining flow rate 2.5 mL/min with a back pressure 250 psi. The compounds thus read at 235 nm. The samples run for 20 min after which the retention time RT was noted. RT was compared with standard thereby identifying the antimicrobial compounds.

Results: Antibacterial studies

This present study involved ethyl acetate extracts of three species of marine actinobacteria being screened for four clinical bacterial strains causing diseases in humans. The actinobacterial extract screened for its activity based on zone of inhibition comparing with the standard drug ciprofloxacin. The average values of inhibition zone of actinobacterial extracts screened against bacteria as shown. Maximum activity found with *Streptomyces alboniger* against *Staphylococcus aureus* as also inferred by Remya and Vijayakumar, (2008). Similarly *Streptomyces coelicolor* exerted maximum antibacterial activity against *Staphylococcus aureus* and *Streptococcus faecalis*; *Streptomyces griseus* against *Staphylococcus aureus* and *Proteus vulgaris*. The zone of inhibition localized 14-18 mm near to the standard drug ciprofloxacin. The inhibition zone 17 to 20 mm is exhibited. The actinobacterial extracts 25, 50 and 75 µg/mL screened for antibacterial activity seen only at concentrations of 50 and 75 µg/mL but not with 25 µg/mL wherein the activity found maximum at the concentration of 75 µg/mL for all extracts.

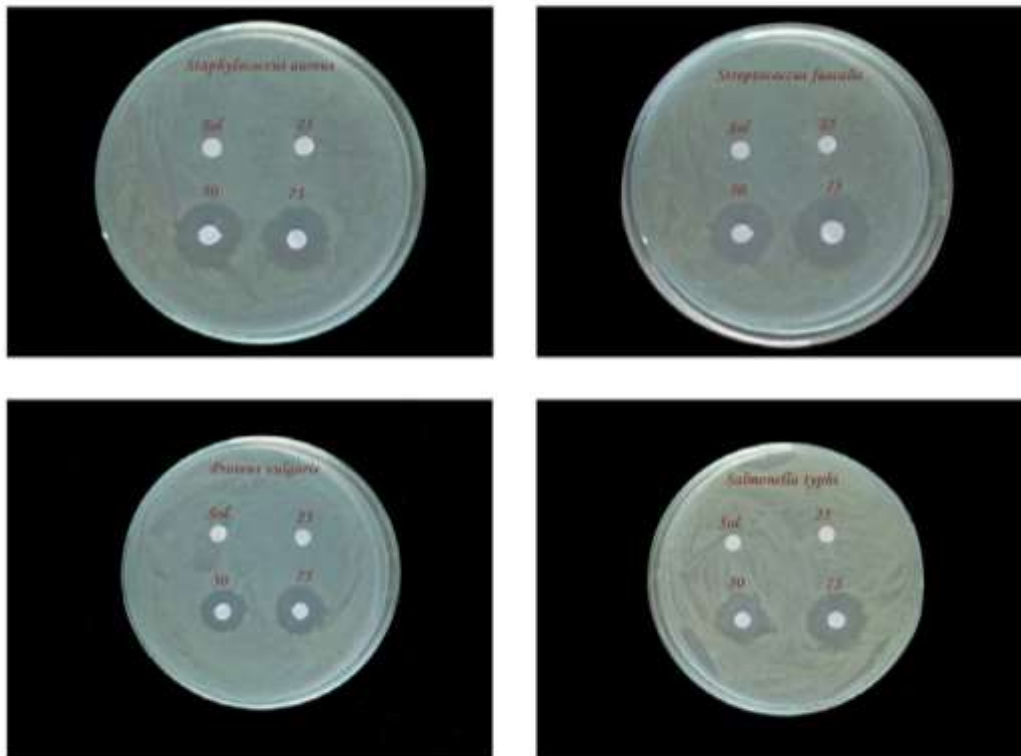
Effect of *Streptomyces alboniger* : extracts at different concentrations
(25, 50 and 75 g/ml) against bacteria



Sol - Ethyl acetate (showed nil effect against the microorganisms under test)
25,50 & 75 - Different concentrations of *S.alboniger* extract



Effect of *Streptomyces coelicolor* : extracts at different concentrations
(25, 50 and 75 g/ml) against bacteria

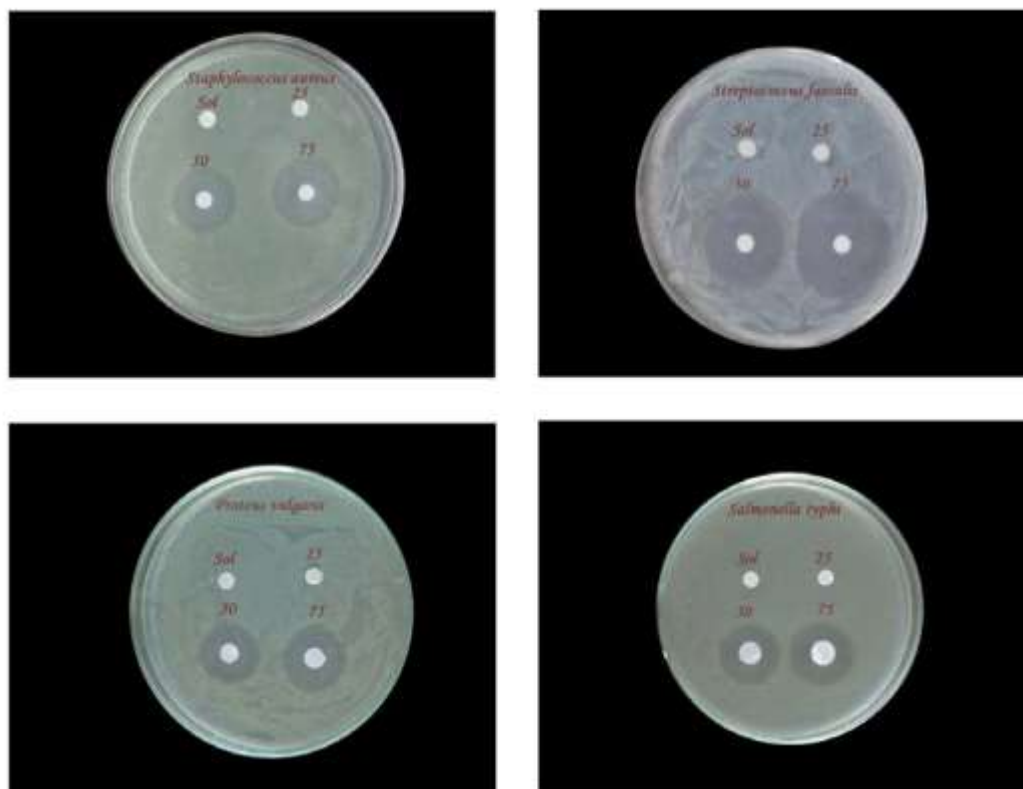


Sol - Ethyl acetate (showed nil effect against the microorganisms under test)

25,50 & 75 - Different concentrations of *S.coelicolor* extract



**Effect of *Streptomyces griseus*: extracts at different concentrations
(25, 50 and 75 g/ml) against bacteria**



**Sol - Ethyl acetate (showed nil effect against the microorganisms under test)
25,50 & 75 - Different concentration of *S.griseus* extract**

Analysis of the antibacterial compounds identified:

HPLC being used in analytical estimation of different antibiotics in research. In this present investigation HPLC profile of antibacterial compounds identified from *Streptomyces alboniger*, *Streptomyces coelicolor* and *Streptomyces griseus* is shown

Bioactive compounds identified in the ethyl acetate extract of species of *Streptomyces*

S. No	Name of marine actinobacteria	RT (min)	Name of the bioactive compounds	Molecular formula	MW	Peak area%	Compound nature	Activity based on scientific literature
1	<i>Streptomyces alboniger</i>	16.41	Pamamycin	C ₃₆ H ₆₃ N ₇ O ₇	621	6.86	Antibiotic	Antibacterial
2	<i>Streptomyces coelicolor</i>	3.313	Cephalexin	C ₁₆ H ₁₇ N ₃ O ₄ S.H ₂ O	365.4	100	Antibiotic	Antibacterial
3	<i>Streptomyces griseus</i>	10.36	Streptomycin	(C ₂₁ H ₃₉ N ₇ O ₁₂) ₂ - 3H ₂ SO ₄	1457.41	100	Antibiotic	Antibacterial

Discussion

Adaptation of marine actinobacteria to extreme climatic conditions as high salinity, pressure and temperature has modified their physiological conditions to survive and elaborate novel bioactive metabolites. Actinobacteria of marine origin offer a promising basis to discover newer biologically active components by Betancur *et al.*, (2017). The marine sediments are often considered to be dominant in origin with respect to actinobacteria involved in the production of beneficial bioactive metabolic products by Girao *et al.*, (2019). Studies pertaining to the marine sediments by Elliah *et al.*, (2002, 2004) of the river Krishna acted as a vivid actinomycetes source in the screening of bioactive metabolites. The studies of Valli *et al.*, (2012) validated that most of the marine antagonistic strains were of actinobacterial origin. Thus the exploitation of marine actinobacteria in antibiotic production stands good in research in the fight against increased pathogen toxicity.

Amongst the marine microorganisms actinomycetes is the most investigated and researched group particularly the genus *Streptomyces* which is credited as having produced the maximum number of antibiotics by Dharmaraj, (2010), Tenebro *et al.*, (2021). This study reveals the major marine species thus isolated of coastal Machilipatnam sediments being *Streptomyces*. The findings based on the studies by Barka *et al.*, (2016) Cumsille *et al.*, (2017) imply the availability of *Streptomyces* as intraspecies

exhibiting distinctive adaptability thereby outcompeting other bacteria in the production of antibiotics. The present study recorded the isolation of 27 species of which three species screened and shown potent antibiotic activity. The three species thus isolated and screened as part of the present investigation characterized by the presence of spores with surface morphology being smooth in accordance to the streptomycetes specification by Tresner *et al.*, (1967). The observations pertaining to this work has implicated that there were three species with good specific activity in defense of bacteria both Gram positive and negative bacteria. Studies of the ethyl acetate extracts, of *Streptomyces alboniger*, *Streptomyces coelicolor*, and *Streptomyces griseus* shown apt effects to 50, 75 µg/ml concentrations. This lead to the identification of pamamycin, cephalixin and streptomycin based on HPLC study involving the above three sample extracts. Amongst which pamamycin being characterized by certain exclusive characteristics stimulates the aerial mycelium in *Streptomyces alboniger*

The studies of Hashimoto *et al.*, (2011) explained the effect of the aerial mycelium in the antibiotic production in *Streptomyces sp.* wherein the inducing compound was pamamycin-607. It induced the production of puromycin in *Streptomyces alboniger* and streptomycin in *Streptomyces griseus* and virginiamycin M1 in *Streptomyces sp.*91. Hoz *et al.*, (2017) described four paulomycin derivatives from *Streptomyces albus* J1074 whereas its biosynthesis was studied by Aranzazu *et al.*, (2016). Victor *et al.*, (2021) studied with reference to the production of 2 antibiotics from *Streptomyces coelicolor* A3(2). Odumosu *et al.*, (2017) emphasized and studied the antimicrobial activities of *Streptomyces coelicolor* AOBKF977550 in the African estuaries. Hence *Streptomyces sp.* being the most abundant is often explored more as there is an everlasting requirement for research in antibiotics by Procopio *et al.*, (2012). *Streptomyces coelicolor* studies on the antibacterial compounds were found to show retention time 3.313 with respect to the absorption peaks.

Research studies by Yague *et al.*, (2013) reflected in the pre sporulation stages of *Streptomyces* in optimizing the differentiation of hyphae in the production of antibiotics. The observations of Manteca and Sanchez, (2009) included differential studies of mycelium and also the production of antibiotics in *Streptomyces coelicolor*. The first isolation of *Streptomyces griseus*, given by Krainsky, (1914) during outbreak of first world war by Waksman and Henrica, (1948) from the soils of Russia as studied by Kuznetsov *et al.*, (1987). Streptomycin an antibiotic far ranging thereby inhibiting bacteria both Gram positive and negative studied by Al-Ansari *et al.*, (2019). Among the different actinobacterial strains isolated from coastal regions of Guntur district Andhra Pradesh India one potent strain with broad-spectrum antagonistic activity identified as *Streptomyces rediverticillatus* VJMS-8 by Swapna and Vijayalakshmi, (2020). This work made the use of *Streptomyces griseus* extracts of ethyl acetate able to cause the growth and inhibition of bacteria both Gram positive and negative bacteria which inferred the presence of streptomycin in the extract in confirmation with the HPLC analysis. The initial and early studies of Carvajal, (1946) are still in support that few strains of *Streptomyces griseus* researched from other sources produced streptomycin with variable function.

References:

- [1]. Al Ansari M, Alkubaisi N, Vijayaragavan P, Murugan K (2019): Antimicrobial potential of Streptomyces sp. to the Gram positive and Gram negative pathogens. *Journal of Infection and Public Health*.
- [2]. Al-Askar AA, Baka ZA, Rashad YM, Ghoneem KM, Abdulkhair WM, Hafez EE, Shabana YM (2015): Evaluation of Streptomyces griseorubens E44G for the biocontrol of Fusarium oxysporum f.sp. lycopersici: ultrastructural and cytochemical investigations. *Annals of Microbiology*.
- [3]. Anderson AS and Wellington EM (2001): The taxonomy of streptomycetes and related genera. *International Journal of Systematic and Evolutionary Microbiology*.
- [4]. Antunes TC, Borba MP, Spadari CC (2014): Screening of actinomycetes with activity against clinical isolates of gram positive cocci with multiresistant profile. *Journal of Advanced and Scientific Research*.
- [5]. Aranzazu G, Rodriguez M, Brana AF, Mendez C, Salas JA, Olano (2016): New insights into paulomycin biosynthesis pathway in Streptomyces albus J1074 and generation of novel derivatives by combinatorial biosynthesis. *Microbial Cell Factories*.
- [6]. Balagurunathan R and Subramaniam A (2001): Antagonistic streptomycetes from marine sediments. *Journal of Advanced Biosciences*.
- [7]. Barka EA, Vatsa P, Sanchez L, Gaveau-Villiant N, Jacquard C, Hans-Peter K, Clement C, Ouhdouch Y, Van Wezel GP (2016): Taxonomy, physiology, and natural products of actinobacteria. *Microbiology and Molecular Biology Reviews*.
- [8]. Behie SW, Bonet B, Zacharia VM, McClung DJ, Traxler MF (2017): Molecules to ecosystems: actinomycetes natural products in-situ. *Frontiers in Microbiology*.
- [9]. Betancur LA, Gaybor SJN, Villarraga DMV, Sarmiento NCM, Maldonado LA, Moreno ZRS, Gonzalez AA, Gonzalez GFP, Puyana M, Castellanos FAR (2017): Marine actinobacteria as a source of compounds for phytopathogen control: an integrative metabolic-profiling/bioactivity and taxonomical approach. *Public Library of Science ONE Journal*.
- [10]. Betina V (1973): Bioautography in paper and thin-layer chromatography and its scope in the antibiotic field. *Journal of Chromatography*.
- [11]. Cabrera JA, Menjivar RD, Dababat AA, Sikora RA (2012): Properties and nematocidal performance of avermectins. *Phytopathology*.
- [12]. Carvajal F (1946): Studies on the structure of Streptomyces griseus. *Mycologia*.
- [13]. Chen L, Wang Z, Du S, Wang G (2021): Antimicrobial activity and functional genes of actinobacteria from coastal wetland. *Current Microbiology*.
- [14]. Cho E, Kwon OS, Chung B, Lee J, Sun J, Shin J, Oh KB (2020): Antibacterial activity of chromomycins from a marine derived Streptomyces microflavus. *Marine Drugs*.
- [15]. Choma IM and Grzelak CE (2011): Bioautography detection in thin-layer chromatography. *Journal of Chromatography A*.
- [16]. Cumsille A, Undabarrena A, Gonzalez V, Claverias F, Rojas C, Camara B (2017): Biodiversity of actinobacteria from the south Pacific and the assessment of Streptomyces chemical diversity with metabolic profiling. *Marine Drugs*.
- [17]. Dharmaraj S (2010): Marine Streptomyces as a novel source of bioactive substances. *World Journal of Microbiology and Biotechnology*.
- [18]. Dholakiya RN, Kumar R, Mishra H, Mody KH, Jha B (2017): Antibacterial and antioxidant activities of novel actinobacteria strain isolated from gulf of Khambhat, Gujarat. *Frontiers in Microbiology*.

- [19]. Elliah P, Bapi Raju KVVSN, Adinarayana K, Adinarayana G (2002): Bioactive actinomycetes from Krishna River sediments of Andhra Pradesh. *Hindustan Antibiotics Bulletin*.
- [20]. Elliah P, Ramana T, Bapi Raju KVVSN, Sujatha P, Uma Shanker AM (2004): Investigation on marine actinomycetes from Bay of Bengal near Karnataka coast of Andhra Pradesh. *Asian Journal of Microbial Biotechnology and Environmental Sciences*.
- [21]. Elnaby HMA, Elala GMA, Raouf MA, Hamed MM (2016): Antibacterial and anticancer activity of extracellular synthesized silver nanoparticles from marine *Streptomyces rochei* MHM13. *The Egyptian Journal of Aquatic Research*.
- [22]. Girao M, Ribeiro I, Ribeiro T, Azevedo IC, Pereira F, Urbatzka R, Leao PN, Carvalho WF (2019): Actinobacteria isolated from *Laminaria ochroleuca*: A source of new bioactive compounds. *Frontiers in Microbiology*.
- [23]. Gulve RM and Deshmukh AM (2012): Antimicrobial activity of the marine actinomycetes. *Research Gate*.
- [24]. Hamed MM, Abdelfattah LSH, Fahmy NM (2019): Antimicrobial activity of marine actinomycetes and the optimization of culture conditions for the production of antimicrobial agents. *Journal of Pure and Applied Microbiology*.
- [25]. Hashimoto M, Katsura H, Kato R, Kawaide H, Natsume M (2011): Effect of pamamycin-607 on secondary metabolite production by *Streptomyces* sp. *Bioscience, Biotechnology and Biochemistry*.
- [26]. Hoz JF, Mendez C, Salas JA, Olano C (2017): Novel bioactive paulomycin derivatives produced by *Streptomyces albus* J1074. *Molecules*.
- [27]. Janardhan A, Kumar AP, Viswanath B, Saigopal DVR, Narasimha G (2014): Production of Bioactive Compounds by Actinomycetes and Their Antioxidant Properties. *Biotechnology Research International*.
- [28]. Jiang S, Zhang L, Pei X, Deng F, HU D, Chen G, Wang C, Hong K, Yao X, Gao H (2017): Chalcomycins from marine derived *Streptomyces* sp. and their antimicrobial activities. *Marine Drugs*.
- [29]. Jiao W, Zhang F, Zhao X, Hu J, Suh JW (2013): A novel alkaloid from marine-derived actinomycete *Streptomyces xinghaiensis* with broad-spectrum antibacterial and cytotoxic activities. *Public Library of Sciences ONE Journal*.
- [30]. Kamjan M, Sivalingam P, Deng Z, Hong K (2017): Deep Sea Actinomycetes and Their Secondary Metabolites. *Frontiers in Microbiology*.
- [31]. Kathiresan K, Balagrunathan R, Masilamani Selvam M (2005): Fungicidal activity of marine actinomycetes against phytopathogenic fungi. *Indian Journal of Biotechnology*.
- [32]. Kokare CR, Mahadik KR, Kadam SS, Chopade BA (2004): Isolation, characterization and antimicrobial activity of marine halophilic AH1 from west coast of India. *Current Science*.
- [33]. Kpehn FE and Carter GT (2005): The evolving role of natural products in drug discovery. *Nature Reviews Drug Discovery*.
- [34]. Kuznetsov VD, Filippova SN, Poltorak VA (1987): Proposed neotype *Streptomyces ruber* (Krainsky, 1914) Waksman et Henrici, 1948. *Mikrobiologiya*.
- [35]. Lakshmi pathy TD, Prasad ASA, Kannabiran K (2010): Production of biosurfactant and heavy metal resistance activity of *Streptomyces* sp. VITDDK3 – a novel halo tolerant actinomycetes isolated from saltpan soil. *Advanced Biological Research*.
- [36]. Learn-Han L, Yoke-Kqueen C, Shiran MS, Vui-Ling CMW, Nurul Syakima AM, Son R, Andrade HM (2012): Identification of actinomycete communities in Antarctic soil from Barrientos island using PCR-denaturing gradient gel electrophoresis. *Genetics and Molecular Research*.
- [37]. Liua Z, Shi Y, Zhang Y, Zhou Z, Li W, Huang Y, Rodrigues C, Goodfellow M (2008): Classification of related species and the genus *Streptomyces* as *Streptomyces yanii* sp.nov. *International Journal of Systematic Evolutionary Microbiology*.
- [38]. Luo M, Tang I, Dong Y, Huang H, Deng Z, Sun Y (2020): Antibacterial natural products lobophorin L and M from the marine-derived *Streptomyces* sp-4506. *Natural Products Research*.
- [39]. Manteca A and Sanchez J (2009): *Streptomyces* development in colonies and soils. *Applied Environmental Microbiology*.
- [40]. Manishaben Jaiswal, "CYBERCRIME CATEGORIES AND PREVENTION", International Journal of Creative Research Thoughts (IJCRT), ISSN:2320-2882, Volume.7, Issue 1, pp.526-536, February 2019, Available at: <http://www.ijcrt.org/papers/IJCRT1134229.pdf>
- [41]. Odumosu BT, Buraimoh, Okeke CJ, Ogah JO, Michel Jr FC (2017): Antimicrobial activities of the *Streptomyces* coelicolor strain AOBKF977550 isolated from a tropical estuary. *Journal of Taibah University for Science*.
- [42]. Patel N, Dwivedi M, Jadeja S, Begum R (2020): Antibacterial activity of marine bacterial pigments obtained from Arabian sea water samples. *Journal of Pure and Applied Microbiology*.
- [43]. Patil R, Jeyaskaran G, Shanmugan SA, Shakila RJ (2001): Control of bacterial pathogens, associated with fish diseases, by antagonistic marine actinomycetes isolated from marine sediments. *Indian Journal of Marine Science*.
- [44]. Peela S, Kurada VVSNB, Terli R (2005): Studies on antagonistic marine actinomycetes from Bay of Bengal. *World Journal of Microbiology and Biotechnology*.
- [45]. Phongsopitanum W, Suwanborirux K, Tanasupawat S (2019): Distribution and antimicrobial activity of Thai marine actinomycetes. *Journal of Applied Pharmaceutical Sciences*.
- [46]. Procopio RE de Lima, da Silva IR, Martins MK, de Azevedo JL, de Araiyo JM (2012): Antibiotics produced by *Streptomyces*. *The Brazilian Journal of Infectious Diseases*.
- [47]. Ramesh S and Mathivanan N (2009): Screening of marine actinomycetes isolated from the Bay of Bengal, India for antimicrobial activity and industrial enzymes. *World Journal of Microbiology and Biotechnology*.
- [48]. Rana FelB, Samia SA, Hanan AG, Soraya AS (2018): Characterization and antimicrobial activity of AgNPS synthesized by *Streptomyces* sp.RHS16 against fish pathogens. *IOSR Journal of Pharmacy and Biological Sciences*.
- [49]. Rana S and Salam MD (2014): Antimicrobial potential of actinomycetes isolated from soil samples of Punjab. *Indian Journal of Microbiology and Experimentation*.

- [50]. Remya M and Vijayakumar R (2008): Isolation and characterization of marine antagonistic actinomycetes from west coast of India. *Facta Universitatis Medical Biology*.
- [51]. Sahu MK, Sivakumar K, Kannan (2006): Marine realm: A treasure house for bioprospecting. *Asian Journal of Microbiology and Biotechnology, Environmental Science*.
- [52]. Sahu MK, Sivakumar K, Kannan L (2005): Isolation of actinomycetes from different samples of the Vellar estuary, southeast coast of India. *Pollution Research*.
- [53]. Sathiyaseelan K and Saranraj P (2016): Antagonistic activity of marine actinobacteria- A Review. *Indo-Asian Journal of Multidisciplinary Research*.
- [54]. Selvakumar P, Balamurugan G, Viveka S (2012): Microbial production of vitamin B12 and antimicrobial activity of glucose utilizing marine derived *Streptomyces* sp. *International Journal of Chemical Technology Research*.
- [55]. SivaKumar K, Haritha R, Jagan Mohan YSYV, Ramana T (2011): Screening of marine actinobacteria for antimicrobial compounds. *Research Journal of Microbiology*.
- [56]. Sivakumar K, Sahu MK, Kathiresan K (2005): An antibiotic producing marine *Streptomyces* from the Pichavaram mangrove environment. *Journal of Annamalai University*.
- [57]. Sivalingam P, Hong K, Pote J, Prabakar K(2019): Extreme Environment *Streptomyces*: Potential sources for new antibacterial and anticancer drug leads? *International Journal of Microbiology*.
- [58]. Subramani R and Aalbersberg (2012): Marine actinomycetes: an ongoing source of novel bioactive metabolites. *Microbiological Research*.
- [59]. Swapna MM and Vijayalakshmi M (2020): Influence of cultural parameters on the production of bioactive metabolites by *Streptomyces rectiverticillatus* VJMS-8 isolated from south coastal regions of Andhra Pradesh, India. *International Journal of Pharmaceutical Sciences and Research*.
- [60]. Tenebro CP, Trono DJVL, Vicera CVB, Sabido EM, Ysulat Jr JA, Macaspac AJ, Tampus KA, Fabrigan TA, Saludes JP, Dalisay DS (2021): Multiple strain analysis of *Streptomyces* species from Phillipine marine sediments reveals intraspecies heterogeneity in antibiotic activities. *Science Reports*.
- [61]. Tresner HD, Backus EJ, Hayes JA (1967): Morphological Spore types in the *Streptomyces hygrosopicus* - like complex. *Applied Microbiology*.
- [62]. Umamaheswary K, Sahu MK, Sivakumar K, Thangaradjou T, Sumitha D, Kannan L (2005): Investigations on L-glutaminase producing actinomycetes strain LG-33 from the estuarine fish, *Mugil cephalus* (Linnaeus, 1758). *Environmental Ecology*.
- [63]. Valli S, Sugasini SS, Aysha OS, Nirmala P, Vinoth Kumar P, Reena A (2012): Antimicrobial potential of actinomycetes species isolated from marine environment. *Asian Pacific Journal of Tropical Biomedicine*.
- [64]. Victor TMM, Ndlovu TM, Filho M, Pessela BC, Bull S, Ward AC (2021): Production and evaluation of two antibiotics of *Streptomyces coelicolor* A3(2), prodigiosin and actinomorphodin using micro-porous culture. *Chemical Engineering and Processing-Process Intensification*.
- [65]. Wang C, Lu Y, Cao S (2020): Antimicrobial compounds from marine actinomycetes. *Archives of Pharmaceutical Research*.
- [66]. Yague P, Lopez M, Rioseras B, Sanchez J (2013): Pre-sporulation Stages of *Streptomyces* Differentiation: State-of-the-Art and Future Perspectives. *FEMS Microbiology Letters*.
- Zotchev SB (2012): Marine actinomycetes as an emerging resource for the drug development pipelines. *Journal of Biotechnology*.