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# SCREENING AND ISOLATION OF MOLDS FROM *MURGHAS* (SILAGE) BAGS

<sup>1</sup>Akshay Darekar\*, <sup>2</sup>Sanket karpe, <sup>3</sup>Ganesh Gangarde, <sup>4</sup>Harshal Wandhekar, <sup>5</sup>Akhilesh Rudrabhate, <sup>6</sup>Rushikesh Jadhav Department of Microbiology, Karamshibhai Jethabhai Somaiya College of Art, Commerce and Science Kopargaon, Maharashtra, India

Abstract: Cattle farming is a supplementary business for farmers to improve or boost their income along with agriculture. Good health and an appropriate diet for cattle have a direct impact on the economic product obtained from cattle. Availability of forage for the whole year is the biggest challenge in front of farmers and to overcome it making of Murghas (silage) a great option. Ensilage is the conserved and fermented forage. Some microbial contaminants are responsible for the spoilage of ensilage. In this research article, a study has been carried out on the isolation of the fungal contaminants responsible for spoiling the surface of silage. Along with the isolation of fungal contaminants, this research also provides the solution or remedy to overcome or reduce the problem of ensilage spoilage by molds. High salt concentration shows a reduction in the growth of molds.

IndexTerms - Silage, contaminants, Molds, Fungi, NaCl, Salinity

#### I. INTRODUCTION

Domestic animals such as cattle, sheep, horses, and buffalos have great commercial importance [1]. Cattle farming and milk production from cattle is the supplementary business for farmers to improve or boost their income [2]. The animal feed and appropriate diet of cattle have a direct impact on the health of cattle [3]. The production of milk, flesh, or other products from cattle is also affected by feedings [4]. Forage or fodder can be given to the cattle in fresh form or it can be stored for the future [5]. The traditional method used to store forage was the making of Hay [6]. For lats 50 years making of Murghas (silage) hold popularity and great demand in forage preservation in the regions of intensive cattle farming for milk and flesh [6]. Forage with high moisture content can be preserved by the fermentation process is termed silage [7]. Silage making process comprises more than 50% of the total amount of preserved forage [8]. The main purpose behind silage making is to conserve forage to be used during the seasons of fresh forage unavailability [9]. The crops like maize, sorghum, and wheat can be used for making silage due to their high nutritional value [10]. Instead of the grains, the whole crop is used in silage [11]. The process of making silage involves harvesting the crop, chopping the crop, and storage of chopped crop or forage in a silo or airtight plastic bags by following the addition of required preservatives [12]. In the silage bag, forage has to be filled compactly to reduce the trapped air amount in ensiled forage mass [13]. The rapid achievement of acidic or low pH is the main principle behind the ensilage [14]. The low pH is obtained by lactic acid fermentation and anaerobic condition [15]. The acidification or fermentation process continues for several days from the day of packaging [16]. It also takes several months to complete the acidification of silage. During the fermentation process number of different microorganisms able to grow in an anaerobic condition such as lactic acid bacteria, yeast, clostridia, and enterobacteria compete with each other for available nutrients [6]. The silage microflora is of two types, desirable and undesirable microflora. Lactic acid bacteria fall under the category of desirable microflora and clostridia, enterobacteria are falls under the category of an undesirable one. yeasts, listeria, and molds also fall under undesirable aerobic spoiler of silage [17]. Usually, lactic acid bacteria dominate the process of fermentation and it results in decreased pH [18]. Due to the acidic pH, normal microorganisms which can act as a contaminant or forage spoilers cannot be able to grow in silage [19]. Hence silage can be stored for a longer period until the nutritional value of ensilage reduces due to the breakdown of proteins and amino acids [20]. The presence of enterobacteria, clostridia, yeasts, and acetic acid bacteria in silage can cause the synthesis of some toxic components of silage like biogenic amines production by enterobacteria [21]. The abundant presence of clostridia in silage causes an increase in pH which is susceptible to the growth of other contaminating microorganisms. The presence of yeast causes loss of dry matter and it also reduces the sugar contained for acid production due to excess production of ethanol [22]. Acetic acid bacteria cause spoilage of entire maize crop silage. Molds are usually aerobic microorganisms. Molds show growth on the upper surface of silage [23]. It is also an indication of proper compaction or sealing of silage bags. The successful conservation of silage is primarily depending on the appropriate silagemaking technique [24]. The silage quality is also dependent on the composition of the crop including soluble sugar, nitrate, dry matter, lactic acid bacteria, and preservatives added to silage [25].

#### II. MATERIAL AND METHODS

#### 1. Sampling

Samples from the upper surface of Murghas (silage) bags contaminated with fungus were collected from the village Jawalke of Kopargaon, Maharashtra, India. The sample was collected in clean polythene bags and used for further studies.



Figure 1. Silage bag infected with fungus

#### 2. Physico-chemical test of Murghas (silage) sample

The Murghas (silage) bags were tested for Physico-chemical parameters including pH and Temperature during the sampling [26].

#### 3. Screening of sample

Initially, the Murghas (silage) sample contaminated with the fungus was inoculated on different nutrient media-containing plates [27]. The media used for the screening were Potato Dextrose agar, Congo Red Yeast Extract Mannitol Agar (CRYEMA), and Malt Extract agar [28]. The plates inoculated with the sample were incubated at room temperature for 2 to 3 days and the growth of fungus on the plates was observed.

#### 4. Isolation and Morphological characterization

The pure culture of screened fungus was made using the same growth media. The morphological characterization was done by observing the fungal colonies on respective culture media plates. After morphological analysis, the fungal staining of the isolated fungus was performed using the dye Lactophenol cotton blue, and the results of the microscopic examination were recorded [29].

# 5. NaCl treatment to reduce the growth of fungal isolate

High salinity can inhibit the growth of microorganisms. To overcome the problem of fungal contamination in silage bags the isolated fungus was grown on high salt-containing media. The growth media was prepared by varying the salt concentration from 5% to 10% and the ability of high salt concentration to inhibit the growth of the isolated fungus was tested. The plates were incubated for 2 to 3 days at room temperature [30] [31].

#### III. RESULT AND DISCUSSION

# 1. Physico-chemical characters of Murghas (silage)

After analyzing the silage sample for their Physico-chemical properties, the pH of silage was found to be acidic due to the presence of essential microflora and the anaerobic fermentation process. The presence of lactic acid bacteria in the silage is the main reason for the acidic pH of Murghas (silage). The temperature of the silage bags was found to be increased due to the microbial metabolism and fermentation process in silage bags. The temperature of ensilage was found more than  $55^{\circ}$ C.

#### 2. Screening of sample

After 2 to 3 days of incubation, the plates of Malt Extract Agar inoculated with silage sample were observed for the morphological characterization. Among the different types of fungi, Molds were observed on culture media plates with white, greenish, and some black colored filamentous colonies.



Figure 2. Screening of Silage samples for the presence of fungal spoilers.

## 3. Isolation and Morphological characterization

after screening and making pure culture of each fungal colony on different growth media, the growth was observed after 2 to 3 days of incubation. Which were further characterized by staining with lactophenol cotton blue for the microscopic examination. Under the microscope, the mycelial structure of all fungal isolates was observed. Some Fungal isolates were showing Septate hyphal structure while some were sowing aseptate hyphal structure under the microscope. The one is showing a long conidiophore structure with globule vesicles under the microscope. The isolated molds which spoil the silage are found to be *Fusarium*, *Aspergillus*, *Mucor*, *and penicillium* based on microscopic and morphological examination.



Figure 3. The isolated fungal colonies on different media-containing plates.

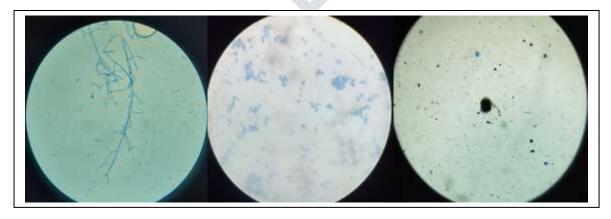


Figure 4. Microscopic examination of fungal isolate using lactophenol cotton blue dye

# TABLE 1. Cultural and Morphological Characterization of Fungal Isolates.

Fungal Isolates	Cultural Characters	Morphological Characters
Mucor	Large white-coloured colonies which then turns into black.	Erect Sporangiospores swells at the tip for sporangia formation.
Penicillium spp.	Fast-growing Green coloured colonies.	Branched conidiophores, septate hyaline hyphae
Aspergillus spp.	Olive to lime green colonies.	Septate and hyaline hyphae. Uncoloured and roughened conidiophores.
Fusarium spp.	White creamy to white greyish coloured colonies.	Septate hyphae with erect conidiophore and conidia.

#### 4. Effect of NaCl treatment on the growth of fungal isolates

Results were observed after inoculating and incubating the fungal isolates on the culture media containing high salt concentration. By observing culture media plates after 2 to 3 days of incubation, it was found that the growth of the fungi was inhibited. The media containing 5% NaCl shows less inhibition of fungi as compared to the media containing 10% NaCl concentration. It means that the increase in salt concentration or salinity of growth media can be able to inhibit, more growth of fungi. This result can be the remedy to overcome the problem of ensilage spoilage due to fungi. The addition of salt on the upper surface of the silage bag during the forage filling and packaging process can prove effective against spoilage and wastage of ensilage.

#### **IV. Conclusion**

Cattle farming is the growing supplementary business for the farmers along with agriculture. To improve the yield of economical products, obtain from cattle, it is important to maintain the good health and diet of the cattle. Availability of forage for the whole year is the biggest challenge in front of cattle farmers. To make forage available for whole years, in many countries silage making practices use to be followed. Ensilage is found to be the best fermented and storage fodder for the cattle. Some microbial contaminants are responsible for the spoilage of the ensilage, which includes yeasts, molds, acetic acid bacteria, enterobacteria, some species of bacilli, etc. In this research, article studies have been carried out on the molds which usually spoil the surface of silage. The isolated molds which spoil the silage are found to be *Fusarium*, *Aspergillus*, *Mucor*, *and penicillium*. To remove or reduce the growth of those fungal contaminants the Sodium Chloride treatment was given to the isolates and it was found that the growth of those isolates can be inhibited by the increased concentration of salt in silage. On the surface of silage, the addition of salt can be a good remedy against those fungal contaminants.

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#### REFERENCES

- 1. Yimer, N., & Rosnina, Y. (2014). Chromosomal Anomalies and Infertility in Farm Animals: A Review. *Pertanika Journal of Tropical Agricultural Science*, 37(1).
- 2. Millogo, V., Ouédraogo, G. A., Agenauml, S., & Svennersten-Sjaunja, K. (2008). Survey on dairy cattle milk production and milk quality problems in peri-urban areas in Burkina Faso. *African Journal of Agricultural Research*, 3(3), 215-224.
- 3. Albright, J. L. (1993). Feeding behavior of dairy cattle. *Journal of dairy science*, 76(2), 485-498.
- 4. Salami, S. A., Luciano, G., O'Grady, M. N., Biondi, L., Newbold, C. J., Kerry, J. P., & Priolo, A. (2019). Sustainability of feeding plant by-products: A review of the implications for ruminant meat production. *Animal Feed Science and Technology*, 251, 37-55.
- 5. Wilkinson, J. M., & Rinne, M. (2018). Highlights of progress in silage conservation and future perspectives. *Grass and Forage Science*, 73(1), 40-52.
- 6. Driehuis, F., & Elferink, S. O. (2000). The impact of the quality of silage on animal health and food safety: a review. *Veterinary Quarterly*, 22(4), 212-216.

- 7. Bolsen, K. K., Ashbell, G., & Weinberg, Z. G. (1996). Silage fermentation and silage additives-Review. *Asian-Australasian journal of animal sciences*, 9(5), 483-494.
- 8. Roer, A. G., Johansen, A., Bakken, A. K., Daugstad, K., Fystro, G., & Strømman, A. H. (2013). Environmental impacts of combined milk and meat production in Norway according to a life cycle assessment with expanded system boundaries. *Livestock Science*, 155(2-3), 384-396.
- 9. Owen, E., Smith, T., & Makkar, H. (2012). Successes and failures with animal nutrition practices and technologies in developing countries: A synthesis of an FAO e-conference. *Animal Feed Science and Technology*, 174(3-4), 211-226.
- 10. Barretto, R., Buenavista, R. M., Rivera, J. L., Wang, S., Prasad, P. V., & Siliveru, K. (2021). Teff (Eragrostis tef) processing, utilization and future opportunities: a review. *International Journal of Food Science & Technology*, 56(7), 3125-3137.
- 11. Ferraretto, L. F., Shaver, R. D., & Luck, B. D. (2018). Silage review: Recent advances and future technologies for whole-plant and fractionated corn silage harvesting. *Journal of dairy science*, 101(5), 3937-3951.
- 12. Wilkinson, J. M., Bolsen, K. K., & Lin, C. J. (2003). History of silage. Silage science and technology, 42, 1-30.
- 13. Sylvester, T. R. (2010). An evaluation of the effects of two different inoculants on the quality of potato hash silage for grower pigs (Doctoral dissertation, [Bloemfontein?]: Central University of Technology, Free State).
- 14. Čabarkapa, I., Palić, D., Plavšić, D., Vukmirović, Đ., & Čolović, R. (2010). The influence of a bacterial inoculant on reduction of aerobic microflora during ensiling of alfalfa. *Food and Feed research*, 37(1), 23-26.
- 15. Fu, W., & Mathews, A. P. (1999). Lactic acid production from lactose by Lactobacillus plantarum: kinetic model and effects of pH, substrate, and oxygen. *Biochemical engineering journal*, *3*(3), 163-170.
- 16. Li, Y., & Nishino, N. (2013). Changes in the bacterial community and composition of fermentation products during ensiling of wilted I talian ryegrass and wilted guinea grass silages. *Animal Science Journal*, 84(8), 607-612.
- 17. JP Aakash Pawar\*, 2Vijay Kadam, 3Saish Joshi, 4Abhishek Rashinkar. Scientist The official Journal of Scientist R Academy 1 (1), 155-162
- 18. Silva, V. P., Pereira, O. G., Leandro, E. S., Da Silva, T. C., Ribeiro, K. G., Mantovani, H. C., & Santos, S. A. (2016). Effects of lactic acid bacteria with bacteriocinogenic potential on the fermentation profile and chemical composition of alfalfa silage in tropical conditions. *Journal of dairy science*, 99(3), 1895-1902.
- 19. AD Aakash Pawar\*, Yogesh Chaudhari, Pradnya Kadu, Rushikesh Jadhav. Scientist The official Journal of Scientist R Academy 1 (1), 163-173
- 20. Kondo, M., Shimizu, K., Jayanegara, A., Mishima, T., Matsui, H., Karita, S., ... & Fujihara, T. (2016). Changes in nutrient composition and in vitro ruminal fermentation of total mixed ration silage stored at different temperatures and periods. *Journal of the Science of Food and Agriculture*, 96(4), 1175-1180.
- 21. Ávila, C. L. S., & Carvalho, B. F. (2020). Silage fermentation—updates focusing on the performance of micro-organisms. *Journal of Applied Microbiology*, 128(4), 966-984.
- 22. Seglar, B. (2003). Fermentation analysis and silage quality testing.
- 23. Wilkinson, J. M., & Davies, D. R. (2013). The aerobic stability of silage: key findings and recent developments. *Grass and Forage Science*, 68(1), 1-19.
- 24. Chedly, K., & Lee, S. (2000). Silage from by-products for smallholders. FAO Plant Production And Protection Papers, 85-96.
- 25. Yitbarek, M. B., & Tamir, B. (2014). Silage additives. Open Journal of Applied Sciences, 2014.
- 26. Cao, C., Bao, W., Li, W., Zhao, F., Kwok, L. Y., Zhang, W., & Zhang, H. (2021). Changes in physico-chemical characteristics and viable bacterial communities during fermentation of alfalfa silages inoculated with Lactobacillus plantarum. *World Journal of Microbiology and Biotechnology*, 37(7), 1-14.
- 27. Kurniati, E., Arfarita, N., Imai, T., Higuchi, T., Kanno, A., Yamamoto, K., & Sekine, M. (2014). Potential bioremediation of mercury-contaminated substrate using filamentous fungi isolated from forest soil. *Journal of Environmental Sciences*, 26(6), 1223-1231.
- 28. PG Yogesh Chaudhari\*, Monali Khairnar, Shital Kashid, Aakash Pawar, Pradnya Kadu. Scientist The official Journal of Scientist R Academy 1 (1), 223-238
- 29. Shamly, V., Kali, A., Srirangaraj, S., & Umadevi, S. (2014). Comparison of microscopic morphology of fungi using lactophenol cotton blue (LPCB), iodine glycerol and congo red formaldehyde staining. *Journal of clinical and diagnostic research: JCDR*, 8(7), DL01.
- 30. McMillen, B. G., Juniper, S., & Abbott, L. K. (1998). Inhibition of hyphal growth of a vesicular-arbuscular mycorrhizal fungus in soil containing sodium chloride limits the spread of infection from spores. *Soil Biology and Biochemistry*, 30(13), 1639-1646.
- 31. Al Tamie, M. S. (2014). Effect of salinity on the fungal occurrence in Al-Shega Area at Al-Qassim, Saudi Arabia. *Research Journal of Microbiology*, 9(6), 287-295.
- 32. Al Tamie, M. S. (2014). Effect of salinity on the fungal occurrence in Al-Shega Area at Al-Qassim, Saudi Arabia. *Research Journal of Microbiology*, 9(6), 287-295.