



Bioactive Phytochemicals from Acanthaceae – Mini Review.

Rajeev Rattan

Associate Professor of Chemistry, Government College, Haripur, Kangra, HPU-Shimla, India.

Abstract : The leaves of the species of Acanthaceae have been traditionally used as herbal medicines by the native and nomadic communities mainly to alleviate the poisonous bites of insects and reptiles and for the treatment of external wounds and ulcers. The pharmacological activities reported from these species are antibacterial, antifungal, anti-pyretic, hepatoprotective, anti-inflammatory, cytotoxic, antioxidant, anti-platelet aggression and insecticidal. The traditional use, chemical constituents and pharmacological activities from family *Acanthaceae* have been selected for study. In literature, from this family phytochemicals mainly flavonoids, alkaloids, lignans, saponins, triterpenoids, steroids and fatty acids are recorded. The species of *Acanthaceae* has emerged as good source of traditional medicines. The chemical compounds isolated from these species have been reported for their pharmacological effects. Although, few experimental studies validated their traditional claim, but uncharacterized crude extract was employed in most of the activities. Such species need to be explored properly for their bioactive principle and exploited as potential drug. The review will help the researchers to select medicinally potential species of *Acanthaceae* for future research.

IndexTerms-Acanthaceae, Flavonoids, Alkaloids, Lignans, Triterpenoid Saponins.

INTRODUCTION

Plants are potential source of herbal medicines beside as resource of food and shelter. The medicinal plants have been the raw base for preparation of medicines in all the pharmacopeias including Chinese, Ayurveda, Unani, Siddha and native and ethnic communities. India is considered as gold mine of well recorded traditional practiced herbal medicines [1]. However, among the 250,000-400,000 plant species only 6% have been screened for biological potential and 15 % for phytochemical exploration [2]. The lack of systematic research, effectiveness, efficacy, disease specific formulations and place of natural product in primary healthcare system are the primary concerns. Still, it is estimated that about 25% drugs prescribed in modern medicines are derived from plants. In the essential list of WHO, out of 252 drugs 11% have plants origin. Despite of advanced treatment through modern medicines, 85 to 90% of world population consumes plants or plants derived medicines due to one reason or other [3]. The plants defense mechanism against the herbivorous animals and pathogens to biosynthesize defensive compounds particularly secondary metabolites have attracted the attention to isolate, characterize and lab scale formulation of these compounds for various biological effects. At the present time, over 12,000 bioactive compounds are identified in science. The recognized active compounds are screened from secondary metabolites. It is an important source with a variety of structural arrangements and properties. Pharmaceutical herbs have a wealthy source of secondary metabolites with fascinating biological activities [4].

Acanthaceae is a large family comprising of 4300 species and 346 genera. This family is cosmopolitan and distributed mostly in tropical and subtropical regions of the world. They are found in the equinoctial regions of both old and new world, with few species extending north into south of Europe, Pennsylvania and Japan and southwards to the Cape of Good Hope and southern coast of New Holland [5]. In India, many of the genera of *Acanthaceae* are peculiar to southern parts, Malayan Peninsula and Indian Archipelago. The species are spreaded from the forest of Silhet to the banks of Sutlej. Some genera have extended to the elevations of 7000 to 8000 feet of Himalayas [6]. All members of this family are shrubs, under shrubs, annual and perennial herbs [7]. Many species of *Acanthaceae* have been used traditionally as herbal drugs for the treatment of various ailments by the native and nomadic communities. The species have been investigated for extraction and isolation of chemical compounds responsible for various pharmacological effects. In the present review, phytochemicals mainly flavonoids, alkaloids, lignans, saponins, triterpenoids, steroids and fatty acids are recorded from the different genera of the family *Acanthaceae* have been selected for the study with the main focus on their structure and pharmacological effects reported in literature. Databases used for study were: Google scholar, PubMed, Scopus-Elsevier, AGRICOLA and Shodhganga.

Traditional uses of family Acanthaceae:

Mostly, leaves of the plants of *Acanthaceae* have been used for traditional purpose. Leaves of *Andrographis peniculata* are used to alleviate bites of poisonous insects and reptiles and for wounds and ulcers [8]. Paste of leaves of *Barberia grandicalyx* is used for snake bites [9]. The paste of onion bulbs and leaves of *Blepharis maderaspatens* are used for cuts and wounds [10]. Leaves part of the plant *Justicia gendrarussa* are used for fever, headache, vertigo and wounds [11]. *Justicia adhatoda* plant is used for rheumatism, fever, pneumonia, asthma, tuberculosis, expectorant, antiseptic and to relieve pain [12]. Decoction of whole plant of *Rungia linifolia* is used for ulcers [13]. Leaves of *Hygrophila auriculata* and *Justicia transquebariensis* are used for cough [11]. Infusion of two or three species is also used for various ailments. Ash of leaves of *Justicia betonica*, *Acathus pubescens* and *Justicia flava* has been used for cough, ulcers and flu [10]. The leaves of the species *Acanthus eminens*, *Aystasias chimperi*, *Dyschoristethum bergiiflora*, *Thunbergialata*, *Dyschoris teradicans* and *Lepidagathis scariosa* are infused to treat cough, skin

diseases, wounds, eye infections, anti-diarrhea, edema, pneumonia [10]. *Andrographis paniculata*, *Hygrophila spinosa*, *Barleria prionitis* and *Adhatoda vasica* are used traditionally as antiviral, antipyretic, antiasthmatic and in respiratory diseases [14].

Chemical Composition of Acanthaceae:

Flavonoids

Flavonoids belonging to different phytochemical classes, have been isolated and identified from different parts of *Acanthaceae*. From the genus *Blepharis* the isolated flavonoids belong to the classes flavones- apigenin, blephariside and dihydro flavones-naringenin, rutin aglycones glycosylated with D- glucose sugar and rarely esterified with coumaroyl moieties. The major flavonoids are apigenin 7-(3"-acetyl-6"-E-pcoumaroyl)-glucopyranoside, genistein-7-O-(6"- O-E-caffeoyl)-β-D glucopyranoside, naringenin-7-O-(3"-acetyl-6"-E-pcoumaroyl)-β-D-glucopyranoside were isolated from *Blepharis ciliaris*, *B. scindica* and *B. maderaspatensi* species.

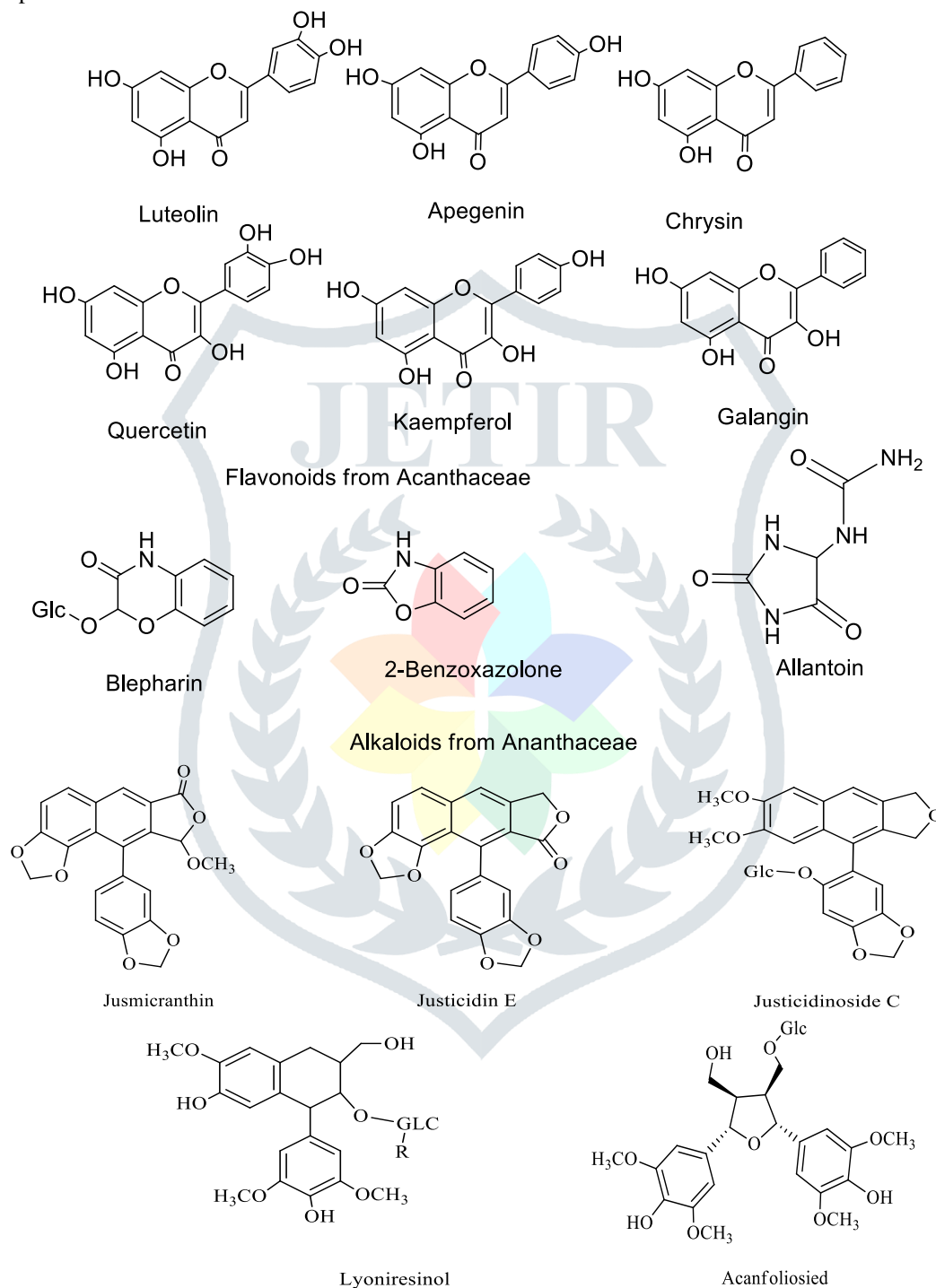


Fig-1 Common Flavonoids, Alkaloids, Lignanas from Acanthaceae.

From the genus *Justicia* -flavanol and dihydroxyflavonols – kaempferitrin, vitexin, 3',4'-Dihydroxyflavonol were identified from the species- *Justicia cataractae*, *J. gendarussa* and *J. spicigera* [14]. The species *Andrographis paniculata* [15] of *Acanthaceae* led to identify flavones and isoflavones and other class of polyphenyl compounds - andrographolide, 14-deoxy-11,12-dihydro Andrographolide, 14-Deoxyandrographolide, 7-O-methyl dihydro wogonin, (2S)-5,7,2',3'-tetramethoxyflavone, 12-Didehydro-andrographolide, 14-Deoxy-11-hydroxyandro grapholide, Neoandrographolide, andrographoside, 5-hydroxyl - 7,8,2',5'-tetramethoxy-flavone, 5-hydroxy-7,8,2',3'-tetra methoxyflavone, 5-hydroxy-7,2',6'-trimethoxyflavone, 5-hydroxy-7,2',3'-trimethoxyflavone. Different classes of flavonoids were identified from the genus *Ruellia*- Luteolin, Luteolin 7-O-glucoside, Apigenin 7-O-glucuronide, Cirsimaritin, Cirsimarin, Cirsiliol 4'-glucoside, Sorbifolin, Pedalitin, Apigenin, Apigenin 7-O-glucoside, Apigenin 7-O-glucuronide, Hispidulin 7-O-β-D-glucuronopyranoside, Comanthoside B, Hispidulin 7-O-α-L-Rha-

(1"→2")-O- β -D-glucuronopyranoside, Pectolinarigenin 7-O- α -L-Rha-(1"→2")-O- β -D-glucuronopyranoside, Nepetin 7-O-β-D-glucopyranoside, Demethoxycentaureidin 7-O-β-D-glucopyranoside, Pectolinarigenin 7-O- β-D-glucopyranoside, 7-O-Acetyl apigenin, Quercetin, Quercetin 3-O-glucoside, 5, 2', 3' -trihydroxy 7-O-glucoflavone, 5, 7, 4' -trimethoxy 3-ORhamnopyranoside, 2, 2', 4', 6'-tetrahydroxy-chalcone from the species- *Ruellia tuberosa*, *R. prostrata* and *R. brittoniana* [16].

Alkaloids

Few alkaloids including derivatives of hydroxamic acids have been also identified in some *Blepharis* species [17]. The bitter glucoside blepharin was first isolated from the *B. edulis* and its structure was later confirmed as blepharin i.e. *N*-deoxy-2,4-

Table1-Pharmacological activities from Acanthaceae.

Activity	Isolate/ extract	Species	Animals/ Species	Ref.
Antioxidant	Methanol, Aqueous	Juticia spicigera, Ruellia kerrii, Barleria cristata, Justicia procumbens, Strobilanthes aureculata, Rhinacanthus nasutus	DPPH/In vitro	[43,44]
Antibacterial	Methanol, Aqueous, chloroform,	Andrographis paniculata, Asteracantha logifolia, Andrographis paniculata, Rhinacanthus nasutus, Andrographis affinis	<i>B. subtilis</i> , <i>S. aureus</i> , <i>E. coli</i> , <i>B. pseudomallei</i> , <i>P. vulgaris</i> , <i>P. aeruginosa</i>	[45,46]
Antifungal	Acetone, Chloroform Dichloromethane	Hypoestes serpens, Andrographis paniculata, Rhinacanthus nasutus	<i>C. cucumerinum</i> <i>C. albicans</i> , <i>A. niger</i> , <i>P. oryzae</i>	[47,44]
Cytotoxic	Methanol, Elanoside, Justifortinol, Justicinol, Patentiflorin, 4-O-acetyl patentiflorin, Andrographolide, 14-deoxy andrographolide, 14-deoxy -11, 12-di hydroandrographolid e, Rhinacanthin-A, B, C, D	Justicia hyssopifolia, Justicia petentifolia, Andrographis paniculata, Rhinacanthus nasutus	Human cancer cells, Tumor cell lines, Human cervix adenocarcinoma	[48,45,44]
Anti-inflammatory	Ethanol, Aqueous, Methanol	Hygrophila spinosa, Dicliptera verticillata, Justicia grandrarussa, Barleria prionitis, Andrographis paniculata	Male Wister rats, key enzymes used for inflammation	[49,47,46]
Antiviral	Rhinacanthin-E, Rhinacanthin-F	Rhinacanthus nasutus	Variety of DNA and RNA virus.	[44]
Hepato protective	Andrographolide	Andrographis paniculata	Male Wister rats	[51]
Immuno modulatory	Aqueous	Rhinacanthus nasutus	Male rats	[44]
Anti-platelet aggregation	Rhinacanthin-A, B, C, G, H, I, K, M, Q	Rhinacanthus nasutus	Rabbits	[44]
Insecticidal	B-sitosterol-3-O-β-glucoside, Palmitic acid, Linaroside, Acetoside	Acanthus montanus	<i>Aedes aegypti</i>	[52]

dihydroxy-1,4- benzoxazin-3-one glucoside (N-deoxyDIBOA). Blepharigenin (N-deoxy-DIBOA) and blepharin were reported from *B. scindica* and 2-benzoxazolinone (BOA) has been isolated from the roots of *B. edulis*. Allantoin and betaine hydrochloride were isolated from *B. scindica*. Macrocylic polyamine alkaloids are found in several plant families. The principal basic unit of these alkaloids

is the polyamine spermidine or spermine, which is connected with one or two molecules of a 4-hydroxycinnamic acid. The roots of *Aphelandra species* contain as the major alkaloid the macrocylic spermine alkaloid, aphelandrine, Aphelandrine, N1,N5-Di-*p*-coumaroylspermidine, N5,N10]-di-*p*-coumaroylspermidine, N1,N5,N[10]-tri-*p*-coumaroyl spermidine, 18-*O*-Methylchaenorpine, iso-18-*O*-methylchaenorpine from *Aphelandra tetragona* and 6-hydroxy-aphelandrine, 6-acetoxy-aphelandrine from *Aphelandra fuscopunctata*. Polynuclear quinolinic based alkaloids are isolated from the methanol and ethyl acetate extract of the species of *Justicia*. 5*H*,6*H*-Quinindolin-11-one, 10*H*-Quindoline, Jusbetonin, Vasicine, Vasicinone, Allantoin, 6*H*-Quinindoline alkaloids from *J. betonica* and Vasicinol alkaloid from *J. adhatoda*. The alkaloids were tested for their pharmacological effects- Antitumor, bronchodilator, uterotonic, anti-inflammatory and anti-ulcer effects [18]. Indole based indole-3-carboxaldehyde bioactive alkaloids were isolated from *Ruellia species* [19]. A tryptophan derived alkaloid 6-hydroxyluteolin, 6-hydroxyluteolin-7-apioside (crystatin) was isolated from ethanol extract of *Lepidagathis cristata* [20]. Cytotoxic 2- Benzoxazolinone alkaloid and derivatives which were glycosylated in few cases were isolated from *Acanthus ilicifolius*- Acanthicifoline, Trigonellin, 2-benzoxazolinone,

Benzoxazin-3-one, 5,5'-bis-benzoxazoline-2,2'-dione, Benzoxazinoid glucosides, 4-O- β -D-glucopyranosyl-benzoxazolin-2(3H)-one, (2R)-2-D glucopyranosyloxy-2H-1,4-benzoxazine-3(4H)-one, (2R)-2-glucopyranosyloxy-4-hydroxy-1,4-benzoxazine-3-one), 2-hydroxy-2H-1, 4-benzoxazin 3(4H) one[21].

Lignans

Lignans have wide diversity in structure but basic skeleton contain two of more phenylpropanoid units linked through 8-8' bonds [22]. Further investigations of more diverse skeletal units and size the term lignan is used to encompass all linkage types- 8-8', 8-1', 8-5', 8-O-4', 5-5', 3-O-4', 7-1', 8-7', 1-5' and 2-O-3'. Out of various types of lignans, 8-8' linked lignans are commonly found in nature. These are further subdivided as Classical Lignans and Neolignans. In *Acanthaceae* family [14] Classical lignans of type Aryl Naphthalene- Jasmicranthin, Jasmicranthin methyl ether, Neesinoside A, Jasmicranthin methyl ether from *Justicia. Neesii* [23], Helioxanthin, (+)-Isolariciresinol, Podophyllotoxin from *J. flava* [24], Taiwanin E, Taiwanin E methyl ether Justicidin E, Neojusticin A, Diphyllin, Justicidin A, Chinensinaphthol, Justicidinose A, Justicidinose C, Justicidinose B, Procumbenoside A, Diphyllin apioside, Diphyllin apioside-5-acetate from *J. procumbens* [25,26], Justicidin B, Cleistanthin B, Justalakonin, Juspurpurin, Sesamin, Xanthoxylol, Juspurpurin, Sesamin, Xanthoxylol from *J. purpurea* [27], 4''-O-Acetylpatentiflorin B, Patentiflorin A, Patentiflorin B, Carinatone, Justiflorinol from *J. patentiflora*, Tuberculatin, Chinensinaphthol methyl ether, 4'-Dimethyl chinensinaphthol methyl ether, Neojusticin B, Ciliatoside A, Cilinaphthalide A, Cilinaphthalide B, Heliobupthalmin from *J. ciliata* [28,29], (+)-Lyoniresinol 3a-[2-(3,5-dimethoxy-4-hydroxy)-benzoyl]-O- β -D-glucopyranoside, dihydroxymethyl-bis(3,5-dimethoxy-4-hydroxyphenyl)tetrahydrofuran-9(or9')-O- β -D-glucopyranoside, (8R,7'S,8'R)-5,5'-dimethoxyarliciresinol 4-O- β -D-glucopyranoside [30], Acanfolioside, Alangiliginoside C, (+)-syringaresinol-O- β -D-glucopyranoside (+)-lyoniresinol 3-O- β -D-glucopyranoside, (+)-lyoniresinol 2a-O- β -D-galactopyranosyl-3a-O- β -D-glucopyranoside, (+)-lyoniresinol 3a-O- β -D-galactopyranosyl-(1-6)- β -D-glucopyranoside, (-)-lyoniresinol 3-O- β -D-glucopyranoside from the species *Acanthus ilicifolius* [21].

Saponins

The triterpenoid based saponins 3- β -O-[(α -L-rhamnopyranosyl(1 \rightarrow 4)-O- β -D-glucopyranosyl)]16- α -hydroxy-olean-12-en(13)-28-oic acid from *Lepidagathis hyaline* Nees [31], 16,28-dihydroxy 22-acetyl-21-tigloylolean-12-ene-3-O- β -D-glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)-[β -D-glucopyranosyl-(1 \rightarrow 2)]- β -D-glucopyranosyl duronic acid (Cuspidate A), 3-O- α -L-arabinopyranosyl-(1 \rightarrow 3)- α -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside hederagenin (Clemantoside C) from *Lepidagathis cuspidata* [32], α -L-Arabinofuranosyl-(1/4)- β -D-glucuronopyranosyl-(1 \rightarrow 3)-3-hydroxylup-20-ene from *Acanthus ilicifolius* [21].

Steroids, triterpenoids and fatty acid derivatives- Pentacyclic triterpenes- β -amyirin, α -amyirin [33], lupeol, oleanolic acid and ursolic acid [34]. Steroids- Cholesterol, campesterol, stigmasterol [50], β -sitosterol [35], stigmast-7-en-3-ol, stigmasteryl-D-glucopyranoside [36], 28-isofucosterol, octacosyl alcohol [36], β -sitosterol-3-O- β -D-glucopyranoside, stigmasterol-3-O- β -D-glucopyranoside [37] from *Acanthus ilicifolius* [21]. Oleanolic acid, β -Sitosterol, 9-Hydroxydodecanoic acid from *B. scindica* [38], (2S,3S,4R)-2[(2'R)-2'-(Hydroxyeicosanoyl amino) octadecane-1,3,4-triol], β -Sitosterol-3-O- β -D-glucopyranose, Stigmasterol, Stigmasterol-3-O- β -D-glucopyranose, Stigmasterol tetracosanoate from *B. ciliaris* [39]. From *Justicia flava* steroids campesterol, stigmasterol, sitosterol, and sitosterol-D-glucoside were isolated [17]. Sterols- β -Sitosterol, β -sitosterol glucoside, stigmasterol, campesterol and stigmat-6-en-3- β -ol, Triterpenes-Lupeol, Betulin, β -Amyrin, 1-Methyl dammar-22-en-3 β ,18,27-triol, Coumarins - 7-Hydroxy-4-Methyl Coumarin, Dicoumarol have been isolated from the genus [17]. Fatty acid derivatives Palmitic acid, octadecanoic acid, stigmasterol octadecanoate, sitosterol octadecanoate, tetracosanol, octacosanol [37] from *Acanthus ilicifolius*.

Miscellaneous (2R)-2-O- β -D-glucopyranosyl-4-hydroxy-2H-1,4-benzoxazin-3(4H)-one, (2R)-2-O- β -D-glucopyranosyl-7-hydroxy-2H-1,4-benzoxazin-3(4H)-one, 7-chloro-(2R)-2-O- β -D-glucopyranosyl-2H-1,4-benzoxazin-3(4H)-one [41], betaine, vanillic acid [42], luteolin-7-O- β -D-glucuronide, uridine, uracil [40].

Pharmacological activities of family Acanthaceae

The methanol extract of the species of Acanthaceae have expressed strong antioxidant potential under standard pharmacological conditions using DPPH/ in vitro. The crude extracts from the genera *Andrographis*, *Asteracantha*, *Rhinacanthus* and *Hypoestes* have exhibited strong antibacterial and antifungal activities against respective bacterial and fungal strains in disc diffusion method/ in vitro. The chemical compounds Justifortinol, Justicidinol, Patentiflorin, 4-O-acetyl patentiflorin, Andrographolide, 14-deoxy andrographolide, 14-deoxy-11, 12-di hydroandrographolide isolated from *Justicia petentifolia* and Rhinacanthin-A, B, C, D from *Rhinacanthus nasutus* have expressed strong cytotoxic potential against human cell lines culture in MTT and LDH Cytotoxic Assay. The β -sitosterol-3-O- β -glucoside, Palmitic acid, Linaroside, Acetoside isolated from *Acanthus montanus* have shown remarkable efficacy against *Aedes aegypti*. The pharmacological effects examined from Acanthaceae species are summarized as

- In most of the pharmacological activities crude extract was utilized. However, Elanoside, Justifortinol, Justicidinol, Patentiflorin, 4-O-acetyl patentiflorin, Andrographolide, 14-deoxy andrographolide, 14-deoxy-11, 12-di hydroandrographolide, Rhinacanthin-A, B, C, D isolated compounds from the species *Justicia hyssopifolia*, *Justicia petentifolia* [47] were recorded for their cytotoxic activities.
- Rhinacanthin-E, Rhinacanthin-F compounds from *Rhinacanthus nasutus* [48] were used to test Antiviral effects.
- Cuspidate A, Clemantoside C compounds from *Lepidagathis cuspidata* were tested positively for their antifungal activity [32].

REFERENCES:

- Umadevi U and Kamalam M (2014) Phytochemical and Antioxidant studies on important indigenous medicinal plant *Andrographis Paniculata* (Burm.F) Nees. *Int. J. Pharm. Sci. Res.*, 5(12), 5240-5244.
- Bader GN (2013) Antimicrobial studies of rhizome of *Swertia petiolata*. *J App Pharm Sci* 4: 128-130.
- Krishnamoorthy, K., & Subramaniam, P. (2014). Phytochemical profiling of leaf, stem, and tuber Parts of *Solena amplexicaulis* (Lam.) Gandhi using GC-MS. *International Scholarly Research Notices*, 2014, 1-13.

- [4] De-Fatima A, Modolo L.V, Conegero L.S, Pilli R.A, Ferreira C.V, Kohn L.K and de-Carvalho J.E. (2006) Lactones and their derivatives: biological activities, mechanisms of action and potential leads for drug design. *Curr. Med. Chem.*, 13, 3371-3384.
- [5] Royle, J.F. (1970) Illustrations of the Botany and other branches of the natural history of the Himalayan Mountains and of the flora of cashmere. Today's and Tomorrow's Printers & Publishers, New Delhi. I: 296-298.
- [6] Shetty, B.V. and Singh, V. (1993) Flora of Rajasthan. Botanical Survey of India, Calcutta.
- [7] Sharma BD, Balakrishnan NP, Rao RR, Hajra PK. (1993) Flora of India, Botanical Survey of India Calcutta, I:52-80.
- [8] Muthu C, Ayyanar M, Raja N, Ignacimuthu S. (2006) Medicinal plants used by traditional healers in Kancheepuram District of Tamil Nadu, India. *J Ethnobiology Ethnomedicine* 2:1-10.
- [9] Jeruto P, Lukhoba C, Ouma G, Otieno D, Mutai C. (2008) An ethnobotanical study of medicinal plants used by the Nandi people in Kenya. *J Ethnopharmacol* 116:370-6.
- [10] Sandhya B, Thomas S, Isabel W, Shenbagarathai R. (2006) Ethnomedicinal plants used by valaiyan community of piranmalia hills (reserved forest), Tamilnadu, India. a pilot study. *Afr J Tradit Complementary Altern Med.* 3:101-14.
- [11] Mia MMK, Kadir MF, Hossan MS, Rahmatullah M. (2009) Medicinal plants of Garo tribe inhabiting the Madhupur forest region of Bangladesh. *Am-Eur Jour of Sustainable Agriculture.* 3:165-71.
- [12] Husain SZ, Malik RN, Javaid M, Bibi S. (2008) Ethnobotanical properties and uses of medicinal plants of Morgah biodiversity park, rawalpindi. *Par J Boany* 40:1897-911.
- [13] Ignacimuthu S, Ayyanar M, Sankara sivaraman K. (2008) Ethnobotanical study of medicinal plants used by paliyar tribals in theni district of Tamil Nadu, India. *Fitoterapia* 79:562-8.
- [14] Krishnaraju AV, Rao TVN, Sundara raju D, Vanisree M, Tsay HS, Subbaraju GV. (2005) Assessment of bioactivity of indian medicinal plants using brine shrimp (*artemiasalina*) lethality assay. *Int J Appl Sci Eng.* 3:125-34.
- [14] Geone M. Corrêa and Antônio F. de C. Alcântara. (2012) Chemical constituents and biological activities of species of *Justicia* - a review. *Brazilian Journal of Pharmacognosy.* 22(1): 220-238.
- [15] Awan AJ, Ahmed CB, Uzair M, Aslam MS (2014) Family Acanthaceae and Genus *Aphelandra*: Ethnopharmacological and Phytochemical review, *Int J Pharm Pharm Sci*, 6, 10, 44-55.
- [16] Samy MN, Sugimoto S, Matsunami K, Otsuka H, Kamel MS. (2015) CHEMICAL CONSTITUENTS AND BIOLOGICAL ACTIVITIES OF GENUS *RUELLIA*, *International Journal of Pharmacognosy.* 2(6): 270-279.
- [17] Dirar, A.I., Devkota, A.A., Kunwar, R.M., Paudel, K.R., Belwal, T., Gupta, G., Chellappan, D.K., Hansbro, P.M., Dua, K., Devkota, H.P., Genus *Blepharis* (Acanthaceae): A review of ethnomedicinally used species, and their phytochemistry and pharmacological activities, *Journal of Ethnopharmacology*, <https://doi.org/10.1016/j.jep.2020.113255>.
- [18] Subbaraju GV, Kavitha JDR, Rajasekhar D, Jimenez JI (2004) Jusbetonin, the first indolo [3,2-b] quinoline alkaloid glycoside, from *Justicia betonica*. *J Nat Prod* 67:461-462.
- [19] Lin C, Huang Y, Cheng L, Sheu S, Chen C (2006) Bioactive flavonoids from *Ruellia tuberosa*. *Journal of Chinese Medicine* 17:103-109.
- [20] Abubacker MN, Devi PK. (2014) In vitro antifungal potentials of bioactive compound oleic acid, 3-(octadecyloxy) propyl ester isolated from *Lepidagathis cristata* Willd. (Acanthaceae) inflorescence. *Asian Pac J Trop Med.* 7 (Suppl 1). 190-193.
- [21] Saranya A., Ramanathan T., Kesavanarayanan K., Adam A. (2015) Traditional Medicinal Uses, Chemical Constituents and Biological Activities of a Mangrove Plant, *Acanthus ilicifolius* Linn. : A Brief Review. *American-Eurasian J. Agric. & Environ. Sci.*, 15 (2): 243-250.
- [22] Lewis NG, Davin LB. Lignans: Biosynthesis and Function, Washington State University, Pullman, WA, USA ;640-707.
- [23] Rajasekhar D, Subbaraju GV (2000) Jusicranthin, a new aryl naphthalide lignan from *Justicia neesii*. *Fitoterapia* 71: 598-599.
- [24] Küpeli E, Erdemoglu N, Yesilada E, Sener B (2003) Antiinflammatory and antinociceptive activity of taxoids and lignans from the heartwood of *Taxus baccata* L. *J Ethnopharmacol* 89: 265-270.
- [25] Chen CC, Hsin WW, Ko FN, Huang YL, Ou JC, Teng CM (1996) Antiplatelet aryl naphthalide lignans from *Justicia procumbens*. *J Nat Prod* 59: 1149-1150.
- [26] Fukamiya N, Lee K (1986) Antitumor agents, 81. Justicidin-A and diphyllin, two cytotoxic principles from *Justicia procumbens*. *J Nat Prod* 49: 348-350.
- [27] Kavitha J, Gopalaiah K, Rajasekhar DG, Subbaraju V (2003) Juspurpurin, an unusual secolignan glycoside from *Justicia purpurea*. *J Nat Prod* 66: 1113-1115.
- [28] Day SH, Lin YC, Tsai ML, Tsao LT, Ko HH, Chung MI, Lee JC, Wang JP, Won SJ, Lin CN (2002) Potent cytotoxic lignans from *Justicia procumbens* and their effects on nitric oxide and tumor necrosis factor-production in mouse macrophages. *J Nat Prod* 65: 379-381.
- [29] Wu CM, Wu SC, Chung WJ, Lin HC, Chen KT, Chen YC, Hsu MF, Yang JM, Wang JP, Lin CN (2007) Antiplatelet effect and selective binding to cyclooxygenase (COX) by molecular docking analysis of flavonoids and lignans. *Int J Mol Sci* 8: 830-841.
- [30] Samy MN, Khalil HE, Wanas AS, Kamel MS, Sugimoto S, Matsunami K, Otsuka H (2013) Chemical constituents from the leaves of *Ruellia tuberosa*. *Chemistry of Natural Compounds*; 49:175-176.
- [31] R. Yadava. (2001) a new biologically active triterpenoid saponin from the leaves of *lepidagathis hyalina* nees. *Natural product letters.* 15,5,315-322.
- [32] Rattan R, Fozdar BI, Gautam V, Sharma R, Kumar D, Sharma U (2016) Cuspidate A, new antifungal triterpenoid saponin from *lepidagathis cuspidata*. *Natural Product Research*, 1-6.
- [33] Van Kiem, P., T.H. Quang, T.T. Huong, T.H. Nhung le, N.X. Cuong, C. Van Minh, E.M. Choi and Y.H. Kim (2008) Chemical constituents of *Acanthus ilicifolius* L. and effect on osteoblastic MC3T3E1 cells. *Arch. Pharmacol Res.*, 31: 823-829.
- [34] Minocha, P.K. and K.P. Tiwari (1980) Chemical constituents of *Acanthus ilicifolius* Linn. *Pol. J. Chem.*, 54: 2089-2090.

- [35] Zhang, L., Z. Wang, J. Chen, P. Lin, Z. Yang and Y. Lin (2007) Studies on chemical constituents in ethanol extract from *Acanthus ilicifolius* as a pharmaceutical mangrove. *Zhongguo Haiyang Yaowu.*, 26: 5-9.
- [36] Kokpol, U., V. Chittawong and D.H. Miles (1986) Chemical constituents of the roots of *Acanthus ilicifolius*. *J. Nat. Prod.*, 49: 354.
- [37] Zhong, L.J., M.Y. Huang, J.G. Zhang, G.W. Li and Y.H. Zhang (2012) Study on the chemical constituents from *Acanthus ilicifolius* Linn. var. *xiamenensis*. *Zhongguo Haiyang Yaowu.*, 31: 23-28.
- [38] Ahmad, V.U., Burki, Akhtar Mahammad, Mahmood, I., Smith, D.L. (1984) Chemical Constituents of *Blepharis indica* seeds. *J. Chem. Soc. Pakistan* 6, 217–223.
- [39] El-Shanawany, M.A., Sayed, H.M., Ibrahim, S.R.M., Fayed, M.A.A. (2014) Stigmasterol Tetracosanoate, a New Stigmasterol Ester from the Egyptian *Blepharis ciliaris*. *Drug Res. (Stuttg)*. 65, 347–353.
- [40] Huo, C., Y. Zhao, H. Liang and W. Lin (2005) Studies on chemical constituents in herbs of *Acanthus ilicifolius*. *Zhongguo Zhongyao Zazhi.*, 30: 763-765.
- [41] Kanchanapoom, T., M.S. Kamel, R. Kasai, K. Yamasaki, C. Picheansoonthon and Y. Hiraga (2001) Lignan glucosides from *Acanthus ilicifolius*. *Phytochemistry.*, 56: 369-372.
- [42] Huo, C., B. Wang, H. Liang, Y. Zhao and W. Lin (2006) Study on chemical constituents of mangrove *Acanthus ilicifolius*. *Zhongguo Zhongyao Zazhi.*, 31: 2052-2054.
- [43] Charoenchai P, Vajrodaya S, Somprasong W, Mahidol C, Ruchirawat S, Kittakoop P. (2010) Antiplasmodial, acytotoxic, radical scavenging and antioxidant activities of thai plants in the family acanthaceae. *Planta Med.* 76:1940-43.
- [44] Bukke S, Raghu PS, Sailaja G, Kedam TR. (2011) The study on morphological, phytochemical and pharmacological aspects of *rhinacanthus nasutus*. (L) Kurz (A Review). *J Appl Pharm Sci*;1:26-32.
- [45] Singha PK, Roy S, Dey S (2003) Antimicrobial activity of *Andrographis paniculata*. *Fitoterapia* ;74:692-4.
- [46] Samy RP (2005) Antimicrobial activity of some medicinal plants from India. *Fitoterapia*; 76:697-9.
- [47] Rasoamiaranjanahary L, Marstan A, Guilet D, Schenk K, Randimbivololona F, Hostettmann K (2003) Antifungal diterpenes from *hypoesteserpens* (Acanthaceae). *Phytochem*; 62: 15, 333-7..
- [48] Navarro E, Alonso SJ, Trujillo J, Jorge E, Perez C (2001) General Behavior, Txicity, and Cytotoxic Activity of Elenoside, a lignin from *Justiciahyssopifolia*. *J Nat Pro* ;64:134-5.
- [49] Susplugas S, Hung NV, Bignon J, Thoison O, Kruczynsk A, Sevenet T, Gueritte F (2005) Cytotoxic aryl naphthalene lignans from a vietnamese acanthaceae, *justiciapatentiflora*. *J Nat Pro* ;68:737-8.
- [50] Patra A, Jha S, Murthy PN, Vaibhav A, Chattopadhyay P, Panigrahi G, et al. Anti-inflammatory and antipyretic activities of *Hygrophilaspinososa* T. (2009) *Anders leaves* (Acanthaceae). *Trop J Pharm Res*; 8:133-7.
- [51] Bhargual DD, Kumar N, Garg VK, Sharma PK. (2010) Review on plants having Hepatoprotective activity. *J Pharm Res*; 3:2077-82.
- [52] Amin E, Radwan MM, Hawary SSE, Fathy MM, Mohammad R, Becnel JJ (2012) Potent insecticidal secondary metabolites from medicinal plant *acanthus montanus*. *Record Nat Pro* 6:301-5.

