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# Phylogenetic Analysis of Vertebrate Lysozyme

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#### **ABSTRACT**

Lysozyme is a naturally occurring enzyme found in bodily secretions such as tears, saliva, and milk. It functions as an antimicrobial agent by cleaving the peptidoglycan component of bacterial cell walls, which leads to cell death. In present study we chose 20 Vertebrate Species including 1 amphibian, 3 actinopterygii, 7 aves and 9 mammals. Amino acid sequences of Lysozyme from 20 vertebrates were derived from National Center for Biotechnology Information database. Phylogenetic analysis and estimation were performed using MEGA program, and neighbor-joining tree was constructed by this software. In this computational study, the phylogenetic tree of Lysozyme has been investigated. Therefore, dendrograms of lysozyme were depicted. From the result of present phylogenetic study of vertebrate Lysozyme, it concluded that the amino acid sequences among different vertebrate species shows slight to moderate differences without affecting its functions.

Key words: Lysozyme, Antimicrobial activity, MEGA, Phylogeny, Vertebrate

# INTRODUCTION

Lysozyme is an antimicrobial enzyme which has been widely found across the animal kingdom as a natural bactericide. Lysozyme was first discovered by Alexander Fleming (1922) in the nasal mucus and subsequently purified from various plant, animal, microbial (bacteria, virus and fungi) materials (Masschalck and Michiels, 2003; Xue *et al.*, 2004; Parisien *et al.*, 2007; Fleming, 1922). Lysozyme (enzyme commission number 3.2.1.17) is also called as N-acetylmuramidase or muramidase, lysozyme is a hydrolyse-type enzyme that catalyses the breakdown of peptidoglycan polymers of bacterial cell wall at the 1-4 bond between N-acetylmuramic (NAM) acid and N-acetylglucosamine (NAG) residues, thereby lysing sensitive bacteria. *In vitro*, lysozyme is generally considered effective against some Gram-positive bacteria, but ineffective against Gram-negative bacteria (Cunningham *et al.*, 1991)

Although lysozyme is important for driving a pro-inflammatory response, lysozyme also plays a role in limiting inflammation systemically, resulting in decreased inflammatory driven pathology (Nash *et al.*, 2006; Ganz *et al.*, 2003) Due to its bacteriostatic, bactericidal, and antiviral properties, lysozyme is currently used as an active pharmaceutical ingredient in medicine and veterinary medicine as well as in the food industry (Leśnierowski and Kijowski, 2007; Aminlari *et al.*,2014). Lysozyme represents one of the most-studied biopreservatives for antimicrobial packaging application. (Silvetti *et al.*,2017)

By applying a specific evolutionary model to explain the amino acid substitutions observed in the multiple sequence alignment, the evolutionary distances between all pairs of proteins can be computed. This evolutionary distance, which reflects the expected mean number of changes per site that have occurred since two sequences diverged from their common ancestor, is used by the so-called distance-methods for phylogenetic inference. One such method is Neighbor Joining (NJ) (Gabaldón, 2007). NJ constitutes a good and fast heuristic algorithm that estimates the "minimal evolution" tree, a phylogenetic tree which minimizes the sum of the lengths (evolutionary distances) of all its branches (Saitou, 1987).

In present study we chose 20 Vertebrate Species including 1 amphibian, 3 actinopterygii, 7 aves and 9 mammals and performed phylogenetic analysis on lysozyme to explore the study of evolutionary relations among groups of organisms. Establishing a relation between structure, function and evolution of this protein is of paramount

importance because it would provide better understanding of further possible mechanisms that the protein involves in.

# MATERIAL AND METHOD

Amino acids sequences for the lysozyme protein of 20 vertebrate species were taken from National Center for Biotechnology Information database (<a href="http://www.ncbi.nlm.nih.gov">http://www.ncbi.nlm.nih.gov</a>). Alignments were applied in order to build a neighbor-joining phylogenetic tree using the Mega program. A phylogenetic tree was saved and analyzed. Constructed phylogenetic tree is depicted in figure 2

table 1: accession numbers of ncbi entries for lysozyme in different vertebrate species

Accession numbers	Name of vertebrate species
CAA32175.1	Homosepian
AAA41551.1	Rattus norvegicus
AIE13825.1	Canis lupus
ACL81750.1	Gallus gallus
AIA09919.1	Xenopus laevis
CAA42084.1	Oncorhynchus mykiss
AAC37312.1	Bos taurus
AAB16862.1	Sus scrofa
AAA73935.1	Opisthocomus hoazin
AHA61788.1	Capra aegagrus hircus
ACE80211.1	Scophthalmus maximus
AIZ00421.1	Anguilla japonica
AIN76235.1	Oryctolagus cuniculus
AHI42590.1	Ovis ammon
ACL81752.1	Ba <mark>mbusi</mark> cola thoracicus
AAB97109.1	Trichosurus vulpecula
AAB31830.1	Syrmaticus soemmerringii
PKK17361.1	Columba livia
ACL81754.1	Francolinus pondicerianus interpositus
AAB20837.1	Lophura leucomelanos

# **RESULTS AND DISCUSSION**

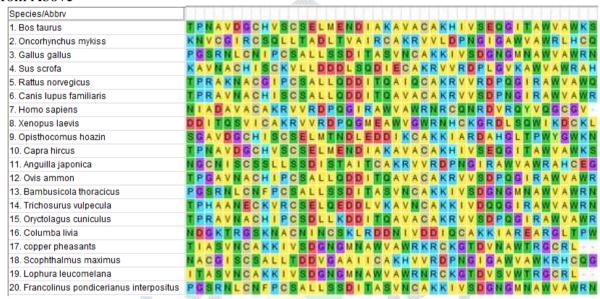
The lysozyme protein sequences of 20 vertebrate species were aligned and analyzed. The phylogenetic analysis was carried out by using distance method and clustering by UPGMA (Unweighted Pair Group Method with Arithmetic Mean). A dendrogram of Lysozyme drawn by using MEGA7multiple sequence alignment of the Lysozyme proteins is depicted in Figure 1 and constructed phylogenetic tree is depicted in Figure 2.

Species/Abbrv				Т			Т							П	1	1.)			- 1,							1		П				1.5
1. Bos taurus	M	K	AL	. 1	1	LE	F	L	FL	S	V.	A١	0	G	K V	F	ER	C	EL	A	R	L	KF	L	GI	D	G	Y	6	V.	3 L	A
2. Oncorhynchus mykiss	M	R	A۱	٧	v	LL	L	٧	A V	A	S	A	٩V	Y	D R	C	EL	A	R A	L	K	3	G N	D	G 1	C A	G	N S	L	PI	Ø W	٧
3. Gallus gallus	M	R	8 1	L	1	LV	L	C	FL	P	L	A	L	G	ΚV	F	G R	C	EL	A	A	M	KF	н	G	D	N	Y F	G	Y	E	G.
4. Sus scrofa	M	ĸ	T L	L	٧	Ľ,	L	L	LL	3	٧	8 1	0	A	KV	Y	D R	C	E:	A	R	L	K	S	<b>G</b> 1	ı D	G	YF	G	V.	L	A
5. Homo sapiens	K	٧	F	R	C	EL	. A	R	TL	K	R	L	M	D	G Y	R	G I	S	L A	M	W I	1 C	LA	K	W	S	G	YN	ľ	R/	X T	N
6. Rattus norvegicus	M	ĸ	A L	L	V	L	F	L	LL	S	A	S	/ Q	A	ΚI	Υ	ER	C	Q F	A	R	L	KF	N	GI	I S	G	Y Y	9	٧	L	A
7. Canis lupus familiaris	M	ĸ	T L	L	F	LE	L	L	LL	3	0	۲	/ Q	G	K I	F	ER	C	E	A	R	L	K	L	GI	. A	G	Y F	G	¥	L	A
8. Xenopus laevis	M	ĸ	L F	Ē	R	C E	L	A	G T	M	K	K١	4 G	Ц	D G	Υ	RG	Y	SL	P	N	٧V	CI	Α	F	E	S	SF	Y	TE	R	
9. Opisthocomus hoazin	M	L	FF	G	F	LL	A	F	L	Α	V.	P	1	E	G E	1	I F	R	CE	L	V)	(1	LF	E	н	F	E	G F	E	G)	ı	1
10. Capra hircus	M	ĸ	A L	. 1	1	LE	F	L	FL	S	V.	A١	0	g	ΚV	F	ER	C	E	A	R	L	K	L	GI	D	G	Y	G	V	L	A
11. Anguilla japonica	M	R	A L	٧	F	LL	L	٧	A V	A	S	A	CV	F	ER	C	Ē٤	A	RI	L	K	A	G N	D	G'	R	G	V S	L	GI	W	٧
12. Ovis ammon	M	K	AL	ı	1	LE	L	L	LL	3	V	A۱	/ 0	G	ΚV	F	ER	C	E۱	A	R	L	KF	F	GI	ı D	G	FF	G	U	L	A
13. Bambusicola thoracicus	M	R	SL	L	1	LV	L	C	FL	P	L	A A	V L	G	KV	F	G R	C	E	A	A	N MA	KF	н	GI	D	N	YF	G	Y	L	G
14. Trichosurus vulpecula	M	ĸ	V L	L	L	LE	F	1	FC	3	M	A A	H	G	KR	М	ER	C	ĒF	A	RF	1 5	K	L	HI	D	G	Y H	Q	I.	L	A
15. Oryctolagus cuniculus	M	K	A L	L	1	LE	L	L	LL	S	T	۲	/ 0	G	K I	Y	ER	C	E۱	A	R	L	K	L	GI	D	G	YE	G	V	L	A
16. Columba livia	M	R	T S	M	L	F۷	G	Υ	LL	. 1	FI	L	L	A	L P	G	T Q	Α	Κt	1	PF	C	EL	V	K	L	R	RH	g	F	G	F
17. copper pheasants	K	٧	Y	R	C	EL	. A	A	A N	IK	RI	L	L	D	N F	R	GY	S	L	N	W	C	A A	K	F	8	14	FN	l T	H/	A T	Ħ
18. Scophthalmus maximus	M	R	C L	L	L	LL	L	٧	A V	A	G	A I	C V	F	ER	C	ĖΝ	A	RL	L	KF	٤N	G N	S	N	R	G		L	A	W	٧
19. Lophura leucomelana	K	٧	Y	R	C	EL	A	A	A N	K	RI	LK	L	D	N Y	R	G Y	3	LG	N.	W	10	A A	K	Y	3	N	FN	ĮΤ	H/	A T	N
20. Francolinus pondicerianus interpositu	M	R	SL	. L	1	L V	L	C	FL	P	L	A A	L	G	ΚV	F	G R	C	Ē١	Α.	A /	M	KF	Н	GI	D	N	Y F	G	Y	L	G

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Species/Abbrv			Т			П	Т	Т	Т	Т	П			П	Т	Т	П		П	Т	Т	h			П	П		T	Т		П	П	Т	Ŧ	Т	Т	
1. Bos taurus	14	WI	C	L	T	K	V E	1	8	Y	N	T	ĸ	A	T I	Y	N	P	S	3	1	Ī	D	Y	G	l F	C	1	N	3	K	M	N	1	IĐ	G	K
2. Oncorhynchus mykiss	C	L	K	W	E		3 1	'n	Ť	Q	A	T	Ñ	R	N 1	D	G	8	Т	١	d	1	E	o	ī	N S	Ē	Y	W	С	D	D	S F	81	P	G	A
3. Gallus gallus	N	W١	/ C	A	A	K	1	ı	Į,	E	N	n	Q	A	TN	IR	N	Т	0		ı	Đ	Y	G	п	Ęĸ	10	N	3	R	W	M	CI	4	G	R	T
4. Sus scrofa	10	W١	/ C	L	A	ΚV	VE		Ü	F	N	T	κ	A	I۱	Н	N	٧	G.		B	Y	G	ī	F	9	h	S	R	Y	W	C	N.	į	K		P
5. Homo sapiens	Y	N/	G	D	R	S	ı	Y	Ġ	I	F	Q	L	П	SF	Y	W	C	N	3 (	K	Г	P	G	A.	1	A	C	Н	L	S	C	S/	4 L	L	Q	D
6. Rattus norvegicus	0	W۱	/ C	L	A	0	ı	1	N	Y	N	T	Q	A	RI	Y	N	P	G (	) c	E	T	D	Y	g	l F	f	1	N	S	R	Y	NE	3	ıg	G	K
7. Canis lupus familiaris	N	W۱	/ C	L	A	ΚV	VE	ı	ı	Y	N	T	R	A	1	Y	N	Ρ	G.	8	\$	T	D	Y	G	I F	¢	1	N	S	R	Y	N C	2 N	IĐ	G	ĸ
8. Xenopus laevis	10	FI	I R	G	D	N	3 1	E	¥Υ	G	1	L	Q	i,	V S	R	W	W	C	V.	Œ	K	Т	P	R	SI	ı	FA	C	N	E	N	CF	2	L	L	S
9. Opisthocomus hoazin	A	DV	V I	C	L	۷	ŀ	E	S	D	Υ	N	T	E	A Y	N	N	N	G	2	R	D	Y	G	1	F	1	N	3	K	Y	N	CI	10	g	K	
10. Capra hircus	N	WL	C	L		Κį	N E	1	ı	Y	N	Ť	ĸ	A	T N	Y	N	Ρ	S	3	1	ī	D	Y	G	I F	ı	1	N	S	K	WV	N	2 N	ľ	G	K
11. Anguilla japonica	C	L/	R	W	E	S	S 1	N	ı	Α	A	I	N	R	V T	D	G	s	Т	3	ď	1	F	Q	i.	V S	E	W	W	C	N	N	3 1	i)	P	S	K
12. Ovis ammon	M	W L	A C	L	Ā	R	V E	ı	H	Ÿ	Ň	T	Q	A	T	Y	N	8	G I	Ē	8	T	D	Y	G.	I F	ï	1	N	S	н	N	NO	21	ı	G	K
13. Bambusicola thoracicus	N	W١	/ C	Α	A	K	1	ı	i	F	K		H	A	T	R	N	T	D		Ī	Н	Y	Ğ	1	Ĺ	Ü	N	8	R	W	M	CI		g	K	T
14. Trichosurus vulpecula	14	W١	/ C	L	A	QÝ	٧Ē		ğ	F	D		ĸ	A	T N	Y	N	Р	G (	9	T	ī	D	Y	G	1.1	ī	1	N	8	н	Y	N C		D	G	K
15. Oryctolagus cuniculus	N	W L	A C	L	A	ΚV	٧Ē	ı	ı	Y	N	T	R	A	TN	Y	N	Ρ	G I	9	ķ,	T	D	Y	G	l F	C	1	N	8	R	Y	NC	2 1	l D	G	K
16. Columba livia	٧	G	Ç T	٧	A	D V	٧N	/ C	L	V	K	Н	E	8	S Y	R	Т	Т	A I	F	48	h	G	P	8	R E	Y	G	1	F	Q	1		×	¢ Y	W	C
17. copper pheasants	R	N	D	G	S	T	3	1	1	L	o	£	N	s	RV	W	C	N	D	F	ı	Р	G	S	R	N L	C	N	Ü	P	C	S	A I	ı	. 8	8	D
18. Scophthalmus maximus	C	L	3 0	W	E	S	9	1	1	R	A	T	N	R	N T	D	Ġ	s	T	2	d	1	F	a	1		F	l V	W	C	D	N	3	1	P	1	3
19. Lophura leucomelana	R	N	0	G	s	T	3		1	L	Q	T	N	S	RV	W	C	N	D (	3	Š	P	G	S	R	N L	C	Н	1	P	С	S	A I	L	S	8	D
20. Francolinus pondicerianus interpositu	I N	W١	/ 0	A	A	K	F	1	N	F	K	Т	H	A	T	R	N	Т	D		1	Y	Y	G	1	2	1	N	S	R	W	N	01	1	G	R	Т

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#### Continued from Above



figure 1: multiple sequence alignment of lysozyme.

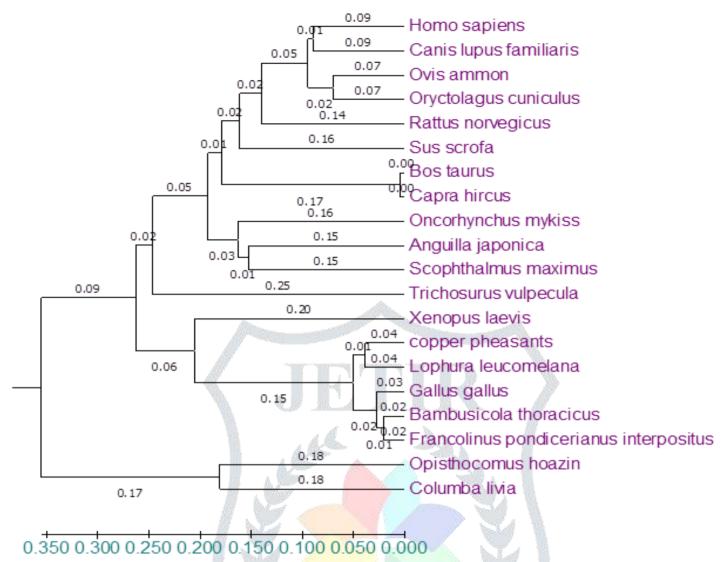


figure 2. phylogenetic tree of lysozyme in vertebrate species.

The phylogenetic tree reveals the formation of an outgroup comprising of *Xenopus laevis* (African clawed frog) and *Trichosurus vulpecula*. The sequences of *Homo sapiens* (Human), *Canis lupus familiaris* (gray wolf), *Ovis ammon* (mountain sheep), *Oryctolagus cuniculus* (European rabbit), *Rattus norvegicus* (brown rat) and *Sus scrofa* (wild boar) bear moderate similarities and also form a cluster.

The sequences of *Bos Taurus (Cattle) and Capra hircus* (domestic goat) are absolutely similar to each and form a cluster. The sequences of *Alligator Oncorhynchus mykiss* (rainbow trout), *Anguilla japonica* (Japanese eel), Scophthalmus *maximus* (turbot) *show* considerable similarity with each other and form a cluster. The sequences of *copper pheasants* (*Syrmaticus soemmerringii*) and *Lophura leucomelana* (kalij pheasant) are absolutely similar to each and form a cluster.

The sequences of *Gallus gallus* (red junglefowl), *Bambusicola thoracicus* (Chinese bamboo partridge), *Francolinus pondicerianus interpositus* shows Considerable similarity and also form a cluster. The sequences of *Opisthocomus hoazin* (hoatzin) and *Columba livia* (rock pigeon) *are* absolutely similar to each and form a cluster. The dendrogram based upon the protein sequences of Lysozyme Protein demonstrate the presence of 6 clusters among the 20 different animals. The similarity in the sequence among all the selected species are shown in the above dendrogram.

## **CONCLUSIONS**

From the results and discussion of present phylogenetic analysis study of vertebrate Lysozyme, the amino acid sequences among different mammalian species show slight to moderate differences without affecting its of antimicrobial activity.

It also concluded from the phylogenetic tree analysis that, Lysozyme in different species also have strong to moderate sequence similarities among studied vertebrates.

# **ACKNOWLEDGEMENT**

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