

HALO BACTERIA *BREVIBACILLUS* SP. ON BIODEGRADATION OF TEXTILE EFFLUENT CONTAINS DIAZO DYE – CONGO RED

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Abstract : Azo dyes are composed of pollutants such as suspended solids and toxic materials. These dyes are discharged into waterbodies without any treatment. This alters the quality and physico – chemical parameters of water. Therefore, the water is unfit for consumption and the flora and fauna inhabiting such fresh water bodies are severely affected. In order to reduce toxicants in textile azo dye effluent suitable microbes which can tolerate wide environmental conditions are used. The halophilic bacterial strains isolated from solar salt pans have the capacity to degrade the selected diazo dye Congo red. The present study was conducted to investigate the degradation of Congo red dye effluent by using halo bacteria *Brevibacillus* sp. This halo bacterium was inoculated into the flask containing Congo red dye effluent and incubated for 5 days. The *Brevibacillus* sp. treated diazo Congo red dye effluent showed reduction of impurities such as pH $13.33 \pm 0.03\%$, temperature $9.09 \pm 0.00\%$, BOD $61.29 \pm 12.65\%$, COD $49.91 \pm 22.94\%$, TDS $57.43 \pm 48.11\%$, chloride $42.61 \pm 39.73\%$ and total hardness $48.73 \pm 40.64\%$ when compared to raw dye effluent.

Index Terms – Xylanase, *Brevibacillus* sp., Diazo dye- Congo red

1. Introduction

The textile industries require enormous quantity of water for dyeing process which generates substantial amount of effluent containing organic and inorganic matters with high concentration (Soares *et al.*, 2002). Dyes are formed of synthetic aromatic amine compounds which are used as colouring agent in food, drug and textile industries (Aksuet *et al.*, 2005). These are classified based on their origin, physical, chemical properties and characteristics related to the application process (Mishra and Tripathy, 1993). Azodyes are extensively applied in various industrial purposes which having one or more azo bond (-N=N-) that cannot be easily broken down and more over some of them making hazard to animals (Fu and Viraraghavan, 2001). Congo red is a benzidine based anionic diazo dye toxic to many organisms and is suspected carcinogen and mutagen which is difficult to degrade due to its stability (Dafare *et al.*, 2013). Nearly about 10 -15 % of unused dyes with various colours are let out into the fresh water bodies with pollutant concentration of 10-200 ppm (Andra predescu and Avran nicolae, 2012). The biological techniques used for dye degradation that provides prominent benefits over the conventional techniques of waste water management and is more ecologically friendly, economically valuable and produces harmless sludge (Kapdanet *et al.*, 2003). The bacterial strains and their extra cellular enzymes which can able to tolerate wide environmental fluctuations can be used to degrade the azo dye compounds. Biodegradation is a pollution control process that uses microorganisms to catalyze the degradation of various hazardous chemicals into less harmful substances and this process of degradation reduces the contaminants in the natural environment is an important alternative to conventional degradation methods (Vidali, 2001).

2. Materials and methods

2.1. Isolation, identification and screening of bacterial strain

The bacterial strain *Brevibacillus* sp. was isolated from sediment sample of hypersaline environment situated at Kovalam, Kanyakumari District, Tamilnadu. The collected sediment sample was serially diluted

as per standard procedure (Bergey's manual of Systematic Bacteriology, 2000). The isolated colonies were screened for xylanase activity by using birch wood xylan agar (Teather and Wood, 1982). The xylanase activity was determined by DNS method (Miller, 1959). The bacterial biomass was determined as per method followed by Bakri *et al.*, 2009.

2.2. Optimization of the bacterial strain

The *Brevibacillus* sp. strain was optimized by physico – chemical parameters such as temperature, pH, NaCl concentration and incubation time for xylanase activity. The optimum activity of xylanase enzyme was determined by keep the reaction mixture with different temperature conditions (30°C - 50°C), pH range (6 - 10), NaCl concentration (1 - 5%) and different incubation periods (12, 24, 48, 72 and 96 hours).

2.3. Physico-chemical parameters

Each 5 ml (18×10^2 CFU/ml) of the optimized bacterial strain *Brevibacillus* sp. was inoculated in three replicates of flask containing 500 ml of Congo red dye effluent and kept in an incubator for 5 days at 35°C temperature. Then, the parameters like pH, temperature, BOD (Biological Oxygen Demand), COD (Chemical Oxygen Demand), TDS (Total Dissolved Solids), chlorides and hardness were determined in raw Congo red dye effluent and bacterial strain *Brevibacillus* sp. treated effluent after 5 days of incubation. The samples were analyzed as per the standard methods prescribed in APHA, 2005.

2.4. Dye degradation

The degradation in the dye treated with bacterial strains expressed as a percentage was determined by using the formula given below.

$$\text{Dye degradation (\%)} = \frac{\text{Initial reading} - \text{Final reading}}{\text{Initial reading}} \times 100$$

Initial reading – Physico chemical parameters of raw effluent

Final reading – Physico chemical parameters of bacterial strain treated effluent

3. RESULTS AND DISCUSSION

The *Brevibacillus* sp. was optimized by various physico - chemical parameters. The bacterial strain *Brevibacillus* sp. showed minimum xylanase activity 67 U/ml and biomass production 29.15 mg/ml respectively were observed at 50°C temperature and maximum 85 U/ml and 39.43 mg/ml respectively were observed at 35°C temperature. Similarly, at pH 10 lowest xylanase activity 64 U/ml and biomass 27.54 mg/ml were observed and highest production 88 U/ml and 41.08 mg/ml respectively observed at pH 8 (Table 1). The findings of Irfan *et al.*, 2016 in *B. subtilis* and Ghasemi *et al.*, 2014 in *Sphingobacterium* sp. SaH-05 showed that maximum xylanase was produced at 35°C and an alkaline pH 8 were resembled with our study. The *Brevibacillus* sp. strain showed minimum xylanase activity 7U / ml and biomass production 1.5 mg / ml at 5 % NaCl concentration and maximum 34.0 U / ml and 13.0 mg / ml respectively at 1 % NaCl concentration. Similarly, the lowest production of xylanase enzyme 60 U / ml and biomass 23.55 mg / ml respectively at 12 hours of incubation and highest production 93 U / ml and 42.63 mg / ml at 48 hours of incubation (Table 2). The result of Padmavathi Tallapragada and Kavya (2011) in fungal strain *Aspergillus niger* and Sharma *et al.*, 2011 in *Bacillus* sp. showed that maximum xylanase produced at 1% NaCl and 48 hours of incubation were coincide with our investigation in the bacterial strain *Brevibacillus* sp.

Table 1: Effect of temperature and pH on the xylanase activity in *Brevibacillus* sp.

Temperature				pH			
S.NO	(°C)	EA (U/ml)	BM(mg/ml)	S.NO	pH	EA (U/ml)	BM(mg/ml)
1	30	81	36.08	1	6	68	30.17
2	35	85	39.43	2	7	75	34.24
3	40	79	35.65	3	8	88	41.08
4	45	74	33.00	4	9	72	31.91
5	50	67	29.15	5	10	64	27.54

*EA- Enzyme Activity

*BM - Biomass

Table 2: Effect of NaCl concentration and incubation time on the xylanase activity in *Brevibacillus* sp.

NaCl concentration				Incubation time			
S.NO	(%)	EA (U/ml)	BM(mg/ml)	S.NO	(Hours)	EA (U/ml)	BM (mg/ml)
1	1	34	13.0	1	12	60	23.55
2	2	19	7.5	2	24	87	40.59
3	3	11	3.4	3	48	93	42.63
4	4	8	1.9	4	72	84	36.85
5	5	7	1.5	5	96	68	26.31

*EA- Enzyme Activity

*BM – Biomass

The bacterial strain *Brevibacillus* sp. showed an excellent effect on degradation of diazo dye Congo red after 5 days of incubation (Table 3). The bacterial strain *Brevibacillus* sp. treated raw Congo red dye effluent showed reduction of impurities when compared to untreated dye. The optimized strain *Brevibacillus* sp. treated Congo red dye effluent recorded a reduction of pH 13.33 ± 0.03 %, temperature 9.09 ± 0.00 %, BOD 61.29 ± 12.65 %, COD 49.91 ± 22.94 %, TDS 57.43 ± 48.11 %, chlorides 42.61 ± 39.73 % and total hardness 48.73 ± 40.64 % when compared to raw dye effluent. The present results were in accordance with the findings of Amaret *et al.*, 2010. According to them, the reduction of parameters such as BOD, COD and total hardness were observed in the bacterial strain *Pseudomonas* sp. SU-EBT treated azo dye - Congo red effluent. Similarly, the bacterial strain *Micrococcus* sp. treated Congo red and Acid Orange effluent showed the reductions of pH, Temperature and TDS were resembled with our present study (Islam *et al.*, 2017).

Table 3: Biodegradation of azo dye Congo red with the optimized bacterial strain *Brevibacillus* sp.

S. No	Parameters	Congo red		
		Raw	Treated	Reduction (%)
1	pH	9 ± 0.01	7.8 ± 0.05	13.33 ± 0.03
2	Temperature (°C)	33 ± 0.00	30 ± 0.00	9.09 ± 0.00
3	BOD (mg/l)	447 ± 11.26	173 ± 13.92	61.29 ± 12.65
4	COD (mg/l)	613 ± 23.14	307 ± 21.86	49.91 ± 22.94
5	TDS (mg/l)	1109 ± 46.58	472 ± 48.94	57.43 ± 48.11
6	Chlorides (mg/l)	542 ± 40.36	311 ± 39.42	42.61 ± 39.73
7	Total hardness (mg/l)	790 ± 39.59	405 ± 43.50	48.73 ± 40.64

4. CONCLUSION

The conventional method of textile waste water treatment is expensive and becomes uneconomical and causes severe environmental pollution. Therefore, ecofriendly and economically viable techniques using halo bacteria can be applied for reducing toxic ingredients in textilewaste water. The biodegradation procedures are cheaper and effective alternative for removal of pollutants in textile azo dye effluent. Thus, the halo bacteria *Brevibacillus* sp. acts as a good biodegradation agent for textile waste water treatment.

REFERENCES

1. Aksu, Z. (2005). "Application of biosorption for the removal of organic pollutants". *Process Biochemistry*. 40: 997 – 1026.
2. Amar A. Telke., Swati M. Joshi., Sheetal U. Jadhav., Dhawal P. Tamboli., Sanjay P. Govindwar. (2010). Decolorization and detoxification of Congo red and textile industry effluent by an isolated bacterium *Pseudomonas* sp. SU-EBT. *Biodegradation*. 21:283–296.
3. American Public Health Association (APHA) (2005) Standard method for examination of water and wastewater, 21st edn. APHA, AWWA, WPCF, Washington.
4. AndraPredescu., AvramNicolae. (2012). "Adsorption of Zn, Cu And Cd From Waste Waters By Means of Maghemite Nanoparticles, U.P.B". *Sci. Bul., Series. B* 74: 255-264.
5. Bakri, Y., Magali, M., Thonart, P. (2009) Isolation and Identification of a New Fungal Strain for Amylase Biosynthesis. *Polish j. of microbiology* 58(3):269-273.
6. Bergey, D.H.L., Holt, J.G. (2000). Bergey's manual of determinative bacteriology. 9th ed. Philadelphia: Lippincott Williams & Wilkins.
7. Dafare, S., Deshpande, P.S., Bhavsar, R.S. (2013). Photocatalytic degradation of congo red dye on combustion synthesised Fe₂O₃. *Ind. J. Chem. Tech.* 20: 406-410.
8. Fu, Y., Viraraghavan T. (2001). "Fungal decolorization of dye wastewaters: a review". *BioresourTechnol* 79: 251-62.
9. Ghasemi., HajarSadeghi., Ahmad Gholami., MiladMohkam., Mohammad Kargar (2014). "Isolation and Identification of Highly Xylanase Producing Bacterium *Sphingobacterium* sp. 10. 11. SaH-05 from Soil". *Int. J. of Sci & Eng Research*, 5: (3), 2229-5518
10. Irfan, M., Asghar, U., Nadeem, M., Nelofer, R., Syed, Q., Shakir, H.A., Qazi, J.I. (2016). "Statistical optimization of saccharification of alkali pretreated wheat straw for bioethanol production". *Waste Biomass Valor.* 7:1389–1396.
11. IslamTarequl., Md. SaifurRahman., Md. Saddam Hussain. (2017). Heavy Metal Tolerance Pattern of Textile Dye Degrading Native Bacteria: A Bioremediation Viewpoint. *Ann Med Health Sci Res.* 7: 67-73.
12. Kapdan, I.K., Tekol, M., Sengul, F., (2003). "Decolorization of simulated textile wastewater in an anaerobic-aerobic sequential treatment system". *Process Biochem.* 38: (7) 1031–1037.
13. Miller, G.L. (1959). "Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar". *Journal of Analytical Chemistry*, 31: 426-428
14. Mishra, G., Tripathy, M. (1993). "A critical review of the treatments for decolourization of textile effluent". *Colourage*, 40: 35-38.
15. Padmavathi Tallapragada., KavyaVenkatesh. (2011). "Isolation, identification and optimization of xylanase enzyme produced by *Aspergillus niger* under submerged fermentation". *J. Microbiol. Biotech. Res.* 1 (4):137-147
16. Sharma, S. C .D., Shovon, M.S., Asaduzzaman, A.K.M., SarowarJahan, M.G., T Yeasmin., T., Roy, N. (2011), "Optimization of alkali-thermostable and cellulose -free xylanase production from *Bacillus* sp". *J. bio-sci.* 19: 7-14.
17. Soares, G.M., Amorim, R., HardinaFerreire, M.C. (2002). "Studies on the biotransformation of novel diazo dyes by laccase". *Process Biochemistry.* 37(6): 581-587.
18. Teather, R. M., Wood, P. J. (1982). "Use of congo red-polysaccharide interactions in enumeration and characterization of cellulolytic bacteria in the bovine rumen". *Appl Environ Microbiol.* 43: 777–780.
19. Vidali, M. (2001). "Bioremediation – an overview". *Pure Application Chemistry.* 73 (7):1163–1172.