



Harmony in Agriculture: Exploring Diverse Soil Types and Harnessing Rhizobacteria for Sustainable Crop Growth

Megha Kumre^{a}, Satish Mohabe^a, Nidhi Pateria Mishra^b*

^aDepartment of Biotechnology, Madhyanchal Professional University, Bhopal, India,462044

^bDepartment of Botany, Madhyanchal Professional University, Bhopal, India,462044

Abstract

This comprehensive study explores the intricate tapestry of agricultural soils, investigating various soil types and their distinctive properties, distribution, and significance within agricultural ecosystems. The examination encompasses loamy, sandy, clay, peaty, silt, chalky, saline, alkaline, laterite, black (vertisols), alluvial, mountain (andosols), and volcanic soils, shedding light on their roles in sustainable and efficient farming practices.

In response to contemporary challenges in agriculture, such as soil erosion, chemical inputs, and limited arable land, the study advocates for the integration of Plant Growth-Promoting Rhizobacteria (PGPR) to stimulate plant growth. The rhizosphere, the soil surrounding plant roots, is identified as a critical environment fostering a diverse population of PGPR. The mechanisms by which PGPR positively influence plant growth, including biological nitrogen fixation, synthesis of phytohormones, phosphorus solubilization, and enzyme production, are elucidated.

The study underscores the potential of PGPR to address current agricultural issues, presenting it as a sustainable alternative to chemical fertilizers and a means to mitigate environmental stressors. The discussion delves into the benefits and drawbacks associated with the use of rhizobacteria in modern agriculture, emphasizing the creation of stable ecosystems, reduced environmental impact, and long-term food security. In conclusion, the study provides valuable insights into the integration of PGPR for enhanced crop production, offering a holistic approach to sustainable agriculture amidst evolving challenges.

Keywords: Soil types, Agricultural ecosystems, Sustainable farming, Rhizobacteria, Plant growth promotion, Environmental sustainability and Crop productivity

Introduction

Agricultural soils are the foundation of global food production, providing the essential nutrients and support for the growth of crops. The diverse range of soils around the world contributes to the rich tapestry of agricultural practices. Understanding the different types of agricultural soils is crucial for sustainable and efficient farming. In this comprehensive exploration, we will delve into various soil types, examining their properties, distribution, and significance in agricultural ecosystems.

1. Loamy Soil:

Loamy soil is often considered the ideal type for agriculture due to its balanced composition of sand, silt, and clay. This soil type provides excellent drainage, retains moisture well, and offers aeration for plant roots. Loamy soils are fertile and support the growth of a wide variety of crops. They are commonly found in regions with moderate climates and are highly prized by farmers for their versatility.

2. Sandy Soil:

Sandy soil is characterized by its coarse texture, allowing for quick drainage and good aeration. However, its drawback lies in poor water and nutrient retention. Sandy soils are prevalent in arid and coastal regions. While they are not as fertile as loamy soils, they can be managed effectively with proper irrigation and fertilization. Crops like carrots, potatoes, and radishes thrive in sandy soils.

3. Clay Soil:

Clay soil is rich in nutrients and has excellent water retention properties. However, it tends to compact easily, restricting root growth and drainage. Proper management practices, such as adding organic matter, can improve its structure. Clay soils are often found in areas with high rainfall and can support a variety of crops, including wheat, barley, and rice.

4. **Peaty Soil:**

Peaty soil, also known as histosol, is characterized by its high organic matter content, primarily derived from partially decomposed plant material. It is usually found in wetlands and marshy areas. While peaty soils are rich in nutrients, they can be acidic and have poor drainage. These soils are commonly used for growing crops like cranberries and blueberries, which thrive in acidic conditions.

5. **Silt Soil:**

Silt soil has fine particles, smaller than those in sandy soil but larger than clay particles. It offers good fertility, retains water well, and has better drainage than clay soil. Silt soils are commonly found in river valleys and are suitable for growing crops like corn, alfalfa, and soybeans. However, they can be prone to erosion, and proper conservation measures are essential for sustainable agriculture.

6. **Chalky Soil:**

Chalky or alkaline soil is characterized by its high pH level due to the presence of calcium carbonate. This type of soil is often found in areas with limestone bedrock. While it can limit the growth of acid-sensitive crops, chalky soils are suitable for crops like grapes and certain vegetables. Soil amendments, such as adding organic matter, can help neutralize the pH and improve fertility.

7. **Saline Soil:**

Saline soils have high concentrations of soluble salts, making them unsuitable for most crops. They are often found in arid and semi-arid regions where evaporation exceeds precipitation, leaving behind salt deposits. Salinity can hinder water uptake by plants, leading to reduced yields. Soil reclamation techniques, such as leaching and the addition of gypsum, are employed to make saline soils more conducive to agriculture.

8. **Alkaline Soil:**

Alkaline soils have a high pH level due to the presence of basic minerals. They can limit nutrient availability to plants, leading to deficiencies. Certain crops, such as beets and broccoli, are more tolerant of alkaline conditions. Soil amendments like sulfur or organic matter can be added to lower pH and improve the fertility of alkaline soils.

9. **Laterite Soil:**

Laterite soils are commonly found in tropical regions and are characterized by their rich iron and aluminum content. These soils can be poor in nutrients, making them less fertile. However, they are suitable for crops like cashew nuts and tea. Laterite soils are often red in color due to the oxidation of iron. Effective soil management, including the addition of organic matter, is crucial for sustainable agriculture on laterite soils.

10. **Black Soil (Vertisols):**

Black soils, also known as vertisols, are clay-rich soils with high expansibility. They are known for their dark color, indicating high organic matter content. These soils can undergo significant volume changes with wetting and drying cycles, which can create challenges for farming. However, black soils are highly fertile and support the cultivation of crops like cotton, soybeans, and sorghum.

11. **Alluvial Soil:**

Alluvial soils are formed by the deposition of sediment carried by rivers. They are typically fertile and well-drained, making them ideal for agriculture. Alluvial plains are among the most agriculturally productive areas globally. The Ganges-Brahmaputra delta in South Asia and the Nile delta in Egypt are prime examples of regions with highly fertile alluvial soils.

12. **Mountain (Andosols) and Volcanic Soil:**

Mountain soils, also known as andosols, are formed in volcanic regions. They are characterized by their high fertility and excellent water retention properties. Volcanic soils often contain essential minerals like phosphorus and potassium. These soils support the cultivation of crops such as coffee, bananas, and various fruits. However, volcanic soils can be prone to erosion, and sustainable farming practices are essential

Stimulation of Plant Growth through the Action of Rhizobacteria that Promote Plant Growth (PGPR)

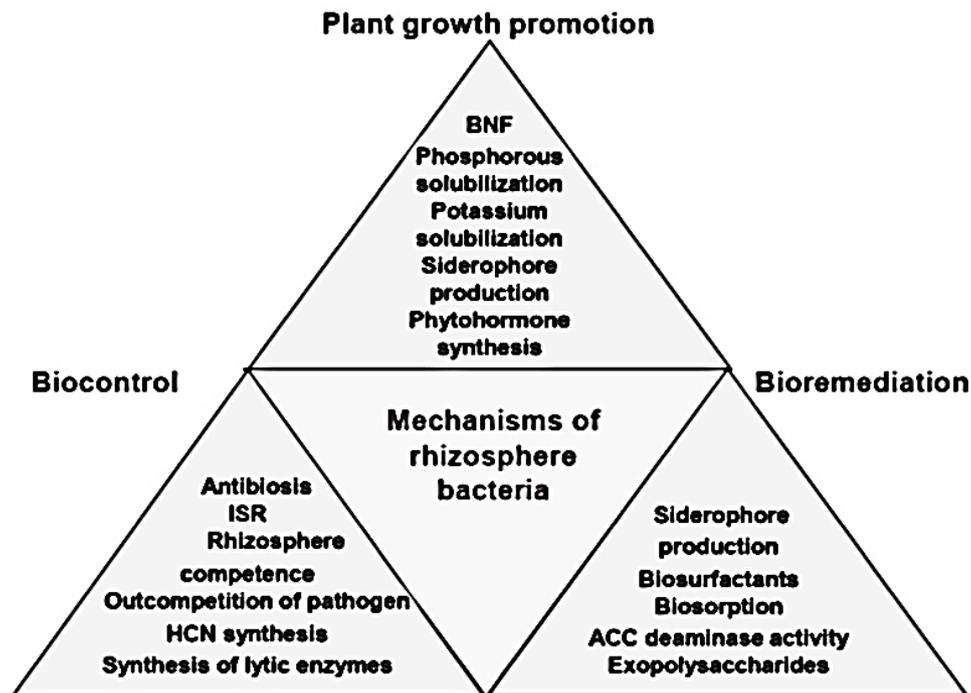
In recent times, the agricultural sector has been confronted with a multitude of issues that have had a significant influence on crop output. Scholars and farmers in today's globe have tremendous challenges when it comes to the discovery of strategies that may increase agricultural production on a limited land area while also providing food for a population that is continually growing [1]. Current agricultural methods that contribute to soil erosion and environmental problems include the use of pesticides, fertilisers, herbicides, and

untreated wastewater for irrigation [2]. Other examples of these activities include the use of synthetic fertilisers. Even though the quantity of land that is accessible for planting is decreasing as a result of urbanisation and industrialization, researchers are actively looking for long-term solutions to increase agricultural yields [3]. In light of the difficulties associated with increasing the amount of land that can be used for food production, it is of the utmost importance to discover methods that may increase agricultural yields per unit area without further depleting natural resources. On account of this, it is of the utmost importance that contemporary agricultural practices embrace novel ways in order to guarantee the continued existence of food security and environmental sustainability in the future.

There have been ideas made in the past for technical techniques that have the potential to increase agricultural output while simultaneously lowering the frequency with which fertilisers and pesticides are required. Within the context of this particular circumstance, there is a rising movement to advocate for the use of rhizobacteria. In the context of plant development, rhizobacteria that promote plant growth and have favourable effects are referred to as rhizobacteria with beneficial effects [5, 6]. The presence of these rhizobacteria in a variety of agricultural settings has the ability to stimulate plant development via a wide variety of direct and indirect processes. The administration of plant growth-promoting rhizobacteria (PGPR) by spraying has been shown to be an efficient strategy for enhancing plant development in field crops that are grown using organic methods, according to previous study. In order to promote the development of plants, a vast variety of bacteria in the rhizosphere adopt a variety of different techniques. Among them include modifications to the metabolism, shifts in the levels of phytohormones, root colonisation, increased nutritional availability, and metabolic engineering [7,8,9]. In order to promote plant development and provide protection against numerous stressors, such as heavy metal pollution and pathogen assaults, these rhizobacteria make use of a combination of different processes. Some of these processes include the manufacture of antibiotics, the generation of induced systemic resistance, the competence of the rhizosphere, and the production of antagonistic substances for the purpose of biocontrol [7,10,11,12,13]. Biosorption, the formation of biosurfactants and siderophores, the activity of ACC deaminase, and the synthesis of polymeric materials are some of the other strategies that rhizobacteria use to clean up contaminated soils [14,15,16].

This chapter is a detailed review of the ways in which bacteria found in the rhizosphere may support the development of plants in a variety of agricultural environments. Rhizobacteria have the ability to successfully

clean up contaminated environments, boost plant development, and provide biocontrol, and we present a thorough explanation of how this might be accomplished. In addition to this, we provide a straightforward summary of the most important results (Figure 1). In the context of modern agriculture, let us now investigate the benefits and drawbacks associated with the use of rhizobacteria for the purpose of improved plant development. A comprehensive scientific analysis of suggestions for the future use of rhizobacteria in agricultural settings is presented at the conclusion of our study.



By providing a setting for a varied population of bacteria in the soil that surrounds the roots of plants, the rhizosphere is an essential component in the process of fostering the development of plants. It is usual practice to refer to microbes that live in the rhizosphere of diverse plants as plant growth-promoting rhizobacteria (PGPR) [17, 18]. These microbes have the ability to have beneficial effects on the host plant via a variety of distinct biological pathways. Exudates from the roots have a substantial influence on the rhizosphere because they incite the presence of bacteria and improve the physicochemical characteristics of the soil. On the other hand, these secretions are very important in ensuring that the microbial populations that are connected with the roots continue to function properly and retain their integrity [9, 19]. The establishment of symbiotic relationships between plants and bacteria facilitates the alleviation of abiotic stressors [20,21,22, 23]. PGPR is involved in a number of processes that are essential to the growth of plants. These processes include nitrogen fixation, the generation of siderophores, the manufacture of phytohormones (including auxins, gibberellins, and cytokinins), the solubilization of phosphorus, and the synthesis of enzymes that alleviate stress [24]. It has

been shown that microorganisms have the capability to stimulate root development, promote the accessibility of nutrients, and reduce the damage that is caused by stress in plants [25, 26]. In addition, PGPR is an essential component in the process of strengthening the resistance of plant symbionts to pathogens, displaying antibiosis effects, and maybe promoting the synthesis of metabolites in plant cells [25, 27]. The capacity of PGPR to withstand a wide range of demanding circumstances, such as heavy metal contamination, weed invasion, salt stress, water scarcity, and nutritional insufficiency, is shown in [28]. The implementation of PGPR may result in a number of desired results, including the creation of a more stable ecosystem and the adoption of agricultural techniques that are less harmful to the environment. Even in the absence of infections, root-associated PGPR have the capacity to enhance the development of plants. They also play a significant part in reducing the detrimental impact that pathogens have on crop productivity via a variety of methods, such as antibiosis, competition, induced systemic resistance, and the generation of siderophores [29,30,31]. In the next part, we will talk about the numerous ways in which PGPR encourages the development of plants.

Exploration of PGPR Mechanisms for Enhancing Plant Growth

Biological Nitrogen Fixation Mechanism

Nitrogen is an essential nutrient that is involved in a wide range of metabolic processes and has a significant role in the development of plants. Despite the fact that nitrogen (N_2) makes up 78% of the gases in the atmosphere, plants are unable to make use of it. Biological nitrogen fixation (BNF) is a process that requires the utilisation of a nitrogenase protein complex by certain archaea and bacteria in order to convert nitrogen into ammonia (NH_3) [7,33,32]. There are a great number of nitrogen fixers that may be found in nature, and it is likely that in the future, microorganisms that are capable of fixing nitrogen dioxide (N_2) might possibly replace chemical fertilisers [34, 35]. It has been proven via research that nitrogen-fixing bacteria are capable of forming either symbiotic connections with leguminous plants or non-leguminous trees, or non-symbiotic partnerships with free-living species or endophytic organisms [7,36,37,38]. There are, in addition to *Rhizobium* and *Frankia*, additional microorganisms that are capable of fixing nitrogen that may be found in soil. These include nitrogen fixers that do not need symbiotic relationships, such as *Cyanobacteria*, *Azospirillum*, *Pseudomonas*, *Azotobacter*, *Acetobacter*, and *Nostoc*. Diazotrophic PGPR is a name that is used to characterise these bacteria that fix nitrogen [18,36]. In spite of the fact that they fix a lower quantity of

nitrogen, non-symbiotic bacteria nevertheless play an important role in the natural environment. This is because nitrogen is essential for the plants that they are dependent on as hosts. It is the complex process of infection and establishment that occurs between symbionts and the roots of leguminous plants that is responsible for the creation of root nodules [39]. The process of oxidative phosphorylation is responsible for the conversion of carbon resources in bacterial metabolism. This process enables the storage of energy in the form of glycogen reserves. Furthermore, the generation of ATP (adenosine triphosphate) and the process of nitrogen fixation are both dependent on this critical activity [40]. In symbionts as well as in free-living organisms that are not symbionts, the nitrogenase complex (nif) genes are responsible for the process of nitrogen fixation [41]. Enzymes are produced by the nif genes, which are responsible for converting atmospheric nitrogen into a form that plants are able to utilise. The nitrogenase complex, which is what the nif genes are responsible for encoding, is the primary enzyme. The Nif genes, in addition to encoding enzymes, also encode regulatory proteins that are involved in the process of nitrogen fixation. A low quantity of nitrogen and a high concentration of oxygen are present in the root environment of the host plant, which results in the production of the nif gene [40]. Rhizobium and bacteroid species are the only ones that solely use oxygen for respiration. On the other hand, the presence of this gas may inhibit the activity of nitrogenase enzymes and have a detrimental effect on the regulation of genes that are involved in the nif pathway. Bacterial haemoglobin is an essential component of root nodules, since it facilitates the process of nitrogen fixation and ensures that sufficient oxygen (O₂) is available for bacteroid respiration. The efficient binding of free oxygen radicals is what allows this to be accomplished [36]. When a haemoglobin gene from the Gram-negative bacteria *Vitreoscilla* sp. was introduced into *Rhizobium etli* by genetic modification, the respiration rates of Rhizobial cells exhibited a considerable rise [38].

Intricate processes that are regulated and include the nitrogenase complex (nif) genes are the activation of the Fe-molybdenum cofactor and the creation of the Fe protein. Both of these processes are considered to be complicated. It is common for the nif genes to be located in close proximity to one another in non-symbiotic bacteria. These genes encompass a range of around 20 to 24 kilobase pairs. This cluster is comprised of seven operons, each of which is accountable for the processing of twenty different proteins. 44 is an important number to consider. Rhizobium species are responsible for the infection and nodulation that may occur in the roots of legume crops [7]. This can occur even with a minor rise in the levels of ethylene. In contrast, legumes

have a reduced degree of nodulation as the amounts of ethylene in the environment increase [43]. In certain instances, some strains of rhizobacterium have the capacity to block the activity of the ACC synthase enzyme, which is involved in the synthesis of ethylene. This enzyme is responsible for the creation of ethylene. The creation of a substance known as rhizobiotoxine in the roots of the legumes that they infect is the means by which this is accomplished [7, 14]. Some rhizobacteria are responsible for the production of ACC deaminase, which is an enzyme that plays a significant part in the elimination of ethylene, a chemical that is essential for the development of green plants [43]. There is a possibility that the nodulation efficiency of these strains might be improved by transferring isolated ACC deaminase genes to *Rhizobium* species that are lacking in ACC deaminase. For instance, a research found that the introduction of a *nif* gene from *R. leguminosarum* *biovar viciae* into the DNA of *Sinorhizobium meliloti* led to a significant increase in the biomass and nodulation of lucerne plants [44]. This was the outcome of the insertion of the gene. This table provides an overview of the numerous studies that have been conducted to study the influence that PGPR has on boosting crop growth via the use of BNF.

Conclusion

In conclusion, this study provides a comprehensive exploration of diverse soil types and their critical roles in agricultural ecosystems. By delving into the unique properties, distribution, and significance of soil varieties such as loamy, sandy, clay, and others, the research contributes valuable insights for sustainable and efficient farming practices. Understanding the complexities of different soil types is crucial for optimizing agricultural strategies and ensuring long-term food security.

The second part of the study highlights the pressing challenges faced by the agricultural sector, including soil erosion, environmental issues from chemical inputs, and the diminishing availability of arable land. In response to these challenges, the study advocates for a sustainable solution – the integration of Plant Growth-Promoting Rhizobacteria (PGPR) to stimulate plant growth. The rhizosphere, as a dynamic environment hosting diverse PGPR populations, emerges as a key player in fostering healthier and more resilient crops.

Exploring the mechanisms by which PGPR positively influences plant growth, such as biological nitrogen fixation, phytohormone synthesis, and phosphorus solubilization, underscores the potential of rhizobacteria to enhance agricultural productivity while minimizing environmental impact. The study emphasizes the need for

a shift towards environmentally friendly and sustainable agricultural practices, highlighting the benefits of PGPR in creating stable ecosystems and reducing reliance on chemical inputs.

The careful consideration of benefits and drawbacks associated with the use of rhizobacteria in modern agriculture informs a balanced perspective. As agriculture faces evolving challenges, adopting PGPR emerges as a promising avenue for promoting sustainable crop production.

In summary, this study contributes valuable knowledge on soil diversity, sustainable farming practices, and the potential of rhizobacteria to revolutionize agriculture. By embracing these insights, the agricultural community can work towards resilient and environmentally conscious practices, ensuring the continued prosperity of global food production systems.

References

1. Bharti N., Barnawal D. Amelioration of salinity stress by PGPR: ACC deaminase and ROS scavenging enzymes activity. In: Singh A.K., Kumar A., Singh P.K., editors. *PGPR Amelioration in Sustainable Agriculture*. Woodhead Publishing; Cambridge, UK: 2019. pp. 85–106.
2. Barrow C. Biochar: Potential for countering land degradation and for improving agriculture. *Appl. Geogr.* 2012;34:21–28. doi: 10.1016/j.apgeog.2011.09.008.
3. Niamat B., Naveed M., Ahmad Z., Yaseen M., Ditta A., Mustafa A., Rafique M., Bibi R., Sun N., Xu M. Calcium-Enriched Animal Manure Alleviates the Adverse Effects of Salt Stress on Growth, Physiology and Nutrients Homeostasis of *Zea mays* L. *Plants*. 2019;8:480. doi: 10.3390/plants8110480.
4. Godfray H.C.J., Beddington J.R., Crute I.R., Haddad L., Lawrence D., Muir J.F., Pretty J., Robinson S., Thomas S.M., Toulmin C. Food security: The challenge of feeding 9 billion people. *Science*. 2010;327:812–818. doi: 10.1126/science.1185383.
5. Saharan B.S., Nehra V. Plant growth promoting rhizobacteria: A critical review. *Life Sci. Med. Res.* 2011;21:1–30.
6. Danish S., Zafar-Ul-Hye M., Mohsin F., Hussain M. ACC-deaminase producing plant growth promoting rhizobacteria and biochar mitigate adverse effects of drought stress on maize growth. *PLoS ONE*. 2020;15:e0230615. doi: 10.1371/journal.pone.0230615. Retracted

7. Ali M.A., Naveed M., Mustafa A., Abbas A. Probiotics and Plant Health. Springer; Singapore: 2017. The good, the bad, and the ugly of rhizosphere microbiome; pp. 253–290.
8. Ismail M.A., Amin M.A., Eid A.M., Hassan S.E.D., Mahgoub H.A., Lashin I., Abdelwahab A.T., Azab E., Gobouri A.A., Elkelish A. Comparative Study between Exogenously Applied Plant Growth Hormones versus Metabolites of Microbial Endophytes as Plant Growth-Promoting for *Phaseolus vulgaris* L. *Cells*. 2021;10:1059. doi: 10.3390/cells10051059.
9. Khan N., Ali S., Shahid M.A., Mustafa A., Sayyed R.Z., Curá J.A. Insights into the Interactions among Roots, Rhizosphere, and Rhizobacteria for Improving Plant Growth and Tolerance to Abiotic Stresses: A Review. *Cells*. 2021;10:1551. doi: 10.3390/cells10061551.
10. Mustafa A., Naveed M., Saeed Q., Ashraf M.N., Hussain A., Abbas T., Kamran M., Minggang X. Sustainable Crop Production. IntechOpen; London, UK: 2019. Application potentials of plant growth promoting rhizobacteria and fungi as an alternative to conventional weed control methods.
11. Mustafa A., Naveed M., Abbas T., Saeed Q., Hussain A., Ashraf M.N., Minggang X. Growth response of wheat and associated weeds to plant antagonistic rhizobacteria and fungi. *Ital. J. Agron*. 2019;14:191–198. doi: 10.4081/ija.2019.1449.
12. Abbas T., Zahir Z.A., Naveed M., Abbas S., Alwahibi M.S., Elshikh M.S., Mustafa A. Large scale screening of rhizospheric allelopathic bacteria and their potential for the biocontrol of wheat-associated weeds. *Agronomy*. 2020;10:1469. doi: 10.3390/agronomy10101469.
13. Ray P., Lakshmanan V., Labbé J.L., Craven K.D. Microbe to microbiome: A paradigm shift in the application of microorganisms for sustainable agriculture. *Front. Microbiol*. 2020;11:3323. doi: 10.3389/fmicb.2020.622926.
14. Ahmad M., Naseer I., Hussain A., Zahid Mumtaz M., Mustafa A., Hilger T.H., Ahmad Zahir Z., Minggang X. Appraising endophyte–plant symbiosis for improved growth, nodulation, nitrogen fixation and abiotic stress tolerance: An experimental investigation with chickpea (*Cicer arietinum* L.) *Agronomy*. 2019;9:621. doi: 10.3390/agronomy9100621.

15. Nazli F., Mustafa A., Ahmad M., Hussain A., Jamil M., Wang X., Shakeel Q., Imtiaz M., El-Esawi M.A. A Review on Practical Application and Potentials of Phytohormone-Producing Plant Growth-Promoting Rhizobacteria for Inducing Heavy Metal Tolerance in Crops. *Sustainability*. 2020;12:9056. doi: 10.3390/su12219056.
16. Haider F.U., Ejaz M., Cheema S.A., Khan M.I., Zhao B., Cai L., Salim M.A., Naveed M., Khan N., Núñez-Delgado A., et al. Phytotoxicity of petroleum hydrocarbons: Sources, impacts and remediation strategies. *Environ. Res.* 2021;197:111031. doi: 10.1016/j.envres.2021.111031.
17. Goudaa S., Kerryb R.G., Dasc G., Paramithiotisd S., Shine H.S., Patra J.K. Revitalization of plant growth promoting rhizobacteria for sustainable development in agriculture. *Microbiol. Res.* 2018;206:131–140. doi: 10.1016/j.micres.2017.08.016.
18. Umar W., Ayub M.A., urRehman M.Z., Ahmad H.R., Farooqi Z.U.R., Shahzad A., Rehman U., Mustafa A., Nadeem M. *Resources Use Efficiency in Agriculture*. Springer; Singapore: 2020. Nitrogen and phosphorus use efficiency in agroecosystems; pp. 213–257.
19. Pandey P., Irulappan V., Bagavathiannan M.V., Senthil-Kumar M. Impact of combined abiotic and biotic stresses on plant growth and avenues for crop improvement by exploiting physio-morphological traits. *Front. Plant Sci.* 2017;8:537. doi: 10.3389/fpls.2017.00537.
20. Raza W., Ling N., Yang L., Huang Q., Shen Q. Response of tomato wilt pathogen *Ralstoniasolanacearum* to the volatile organic compounds produced by a biocontrol strain *Bacillus amyloliquefaciens* SQR-9. *Sci. Rep.* 2016;6:24856. doi: 10.1038/srep24856.
21. Raza W., Yousaf S., Rajer F.U. Plant growth promoting activity of volatile organic compounds produced by Bio-control strains. *Sci. Lett.* 2016;4:40–43.
22. Bharti N., Pandey S.S., Barnawal D., Patel V.K., Kalra A. Plant growth promoting rhizobacteria *Dietzianatronolimnaea* modulates the expression of stress responsive genes providing protection of wheat from salinity stress. *Sci. Rep.* 2016;6:34768. doi: 10.1038/srep34768.

23. Ramakrishna W., Rathore P., Kumari R., Yadav R. Brown gold of marginal soil: Plant growth promoting bacteria to overcome plant abiotic stress for agriculture, biofuels and carbon sequestration. *Sci. Total Environ.* 2020;711:135062. doi: 10.1016/j.scitotenv.2019.135062.
24. Olanrewaju O.S., Glick B.R., Babalola O.O. Mechanisms of action of plant growth promoting bacteria. *World J. Microbiol. Biotechnol.* 2017;33:197. doi: 10.1007/s11274-017-2364-9.
25. Khan N., Bano A., Rahman M.A., Guo J., Kang Z., Babar M.A. Comparative physiological and metabolic analysis reveals a complex mechanism involved in drought tolerance in Chickpea (*Cicer arietinum* L.) induced by PGPR and PGRs. *Sci. Rep.* 2019;9:2097. doi: 10.1038/s41598-019-38702-8.
26. Gupta A., Gopal M., Thomas G.V., Manikandan V., Gajewski J., Thomas G., Seshagiri S., Schuster S.C., Rajesh P., Gupta R. Whole genome sequencing and analysis of plant growth promoting bacteria isolated from the rhizosphere of plantation crops coconut, cocoa and arecanut. *PLoS ONE.* 2014;9:e104259. doi: 10.1371/journal.pone.0104259.
27. Verma P.P., Shelake R.M., Das S., Sharma P., Kim J.Y. *Microbial Interventions in Agriculture and Environment.* Springer; Singapore: 2019. Plant growth-promoting rhizobacteria (PGPR) and fungi (PGPF): Potential biological control agents of diseases and pests; pp. 281–311.
28. Kumari A., Kumar R. Exploring phyllosphere bacteria for growth promotion and yield of potato (*Solanum tuberosum* L.) *Int. J. Curr. Microbiol. Appl. Sci.* 2018;7:1065–1071. doi: 10.20546/ijemas.2018.704.117.
29. Shivakumar B. Ph.D. Dissertation. UAS; Dharwad, India: 2007. Biocontrol Potential and Plant Growth Promotional Activity of Fluorescent Pseudomonads of Western Ghats.
30. Mohammed A.F., Oloyede A.R., Odeseye A.O. Biological control of bacterial wilt of tomato caused by *Ralstoniasolanacearum* using *Pseudomonas* species isolated from the rhizosphere of tomato plants. *Arch. Phytopathol. Plant Prot.* 2020;53:1–16. doi: 10.1080/03235408.2020.1715756.
31. Larkin R.P. Biological control of soil borne diseases in organic potato production using hypovirulent strains of *Rhizoctoniasolani*. *Biol. Agric. Hortic.* 2020;36:1–11. doi: 10.1080/01448765.2019.1706636.
32. Dixon R., Kahn D. Genetic regulation of biological nitrogen fixation. *Nat. Rev. Microbiol.* 2004;2:621–631. doi: 10.1038/nrmicro954.

33. Kim K.Y.D., Jordan D., McDonald G.A. Solubilization of hydroxyapatite. Enterobacter agglomerans and cloned Escherichia coli in culture medium. *Biol. Fertil. Soils.* 1997;24:347–352. doi: 10.1007/s003740050256.
34. Ladha J.K., de Bruijn F.J., Malik K.A. Introduction: Assessing opportunities for nitrogen fixation in rice-a frontier project. *Plant Soil.* 1997;124:1–10. doi: 10.1023/A:1004264423436.
35. Raymond J., Siefert J.L., Staples C.R., Blankenship R.E. The natural history of nitrogen fixation. *Mol. Biol. Evol.* 2004;21:541–554. doi: 10.1093/molbev/msh047.
36. Bhattacharyya P.N., Jha D.K. Plant growth-promoting rhizobacteria (PGPR): Emergence in agriculture. *World J. Microbiol. Biotechnol.* 2012;28:1327–1350. doi: 10.1007/s11274-011-0979-9.
37. Ahmad M., Khan M.S. Effects of pesticides on plant growth promoting traits of Mesorhizobium strain MRC4. *J. Saudi Soc. Agric. Sci.* 2012;11:63–71.
38. Zahran H.H. Rhizobia from wild legumes: Diversity, taxonomy, ecology, nitrogen fixation and biotechnology. *J. Biotechnol.* 2001;91:143–153. doi: 10.1016/S0168-1656(01)00342-X.
39. Giordano W., Hirsch A.M. The expression of MaEXP1, a Melilotus alba expansin gene, is upregulated during the sweetclover-Sinorhizobium meliloti interaction. *Mol. Plant Microbe Interact.* 2004;17:613–622. doi: 10.1094/MPMI.2004.17.6.613.
40. Marroquí S., Zorreguieta A., Santamaría C. Enhanced symbiotic performance by Rhizobium tropici glycogen synthase mutants. *J. Bacteriol.* 2001;183:854–864. doi: 10.1128/JB.183.3.854-864.2001.
41. Glick B.R. The enhancement of plant growth by free living bacteria. *Can. J. Microbiol.* 1995;41:1091-114. doi: 10.1139/m95-015.
42. Parray J.A., Jan S., Kamili A.N., Qadri R.A., Egamberdieva D., Ahmad P. Current perspectives on plant growth-promoting rhizobacteria. *J. Plant Growth Regul.* 2016;35:877–902. doi: 10.1007/s00344-016-9583-4.
43. Ma W., Penrose D.M., Glick B.R. Strategies used by rhizobia to lower plant ethylene levels and increase nodulation. *Can. J. Microbiol.* 2002;48:947–954. doi: 10.1139/w02-100.

44. Ma W., Charles T.C., Glick B.R. Expression of an exogenous 1 aminocyclopropane-1-carboxylate deaminase gene in *Sinorhizobium meliloti* increases its ability to nodulate alfalfa. *Appl. Environ. Microbiol.* 2004;100:5891–5897. doi: 10.1128/AEM.70.10.5891-5897.2004.

