



Elemental Analysis of Seeds of Drum Stick (*Moringa Oleifera*) by Using Different Techniques

Sachin S Musale¹, Rajesh B. Jadhav²

¹Assistant. Professor, MIT-ADTU, School of Computing, Dept. of Applied Sciences & Humanities, Pune

²Head of Chemistry, MIT-ADTU, School of Computing, Dept. of Applied Sciences & Humanities, Pune

Abstract: For thousands of years, plants have been a significant source of medicine. According to estimates from the World Health Organization (WHO), up to 80% of people still get their medications mostly from traditional remedies like herbs. These plants' elemental composition and the presence of a range of phytochemicals provide them therapeutic potential.

The ability of medicinal plants to prevent or manage disease has been linked to the antioxidant qualities of their contents, which are typically linked to a variety of amphipathic molecules collectively known as poly-phenolic compounds. The food business and preventative healthcare are becoming more interested in the creation and assessment of natural antioxidants derived from medicinal plant components. *Moringa oleifera* is one of the herbs that shows promise. This tree's leaves, fruits, blossoms, and immature pods are all edible and are consumed as a highly nutritious vegetable in many places, especially Hawaii, India, the Philippines, and several African countries. Because of the chemical components of *M. oleifera*, it is utilized in underdeveloped countries as a substitute for imported food supplements to treat and prevent malnutrition, particularly in new-borns and nursing mothers.

Traditional medical systems have employed drum stick seed extract⁵. As a result, support has been provided for the metal analysis of seeds of *M.oleifera*. To determine the metal content, the finely ground fruit and seeds of *M. oleifera* were examined. Several common methods, including Colorimetry, Kjheldal's Method, Atomic adsorption spectrophotometer, and Flame photometry, were used to analyse a material¹². It was noted that while zinc, copper, iron, and manganese were found in considerable amounts, phosphorus, nitrogen, and calcium were detected in small amounts. It was shown that the fruits of *C. Equisetifolia* have higher concentrations of Zinc, Copper, Iron, Arsenic and Manganese than the seeds of *P. Longifolia*.

According to research currently available, seeds of *M. oleifera* may be a good source of zinc, iron, manganese, & copper.

Keywords: Seed of *M. Oleifera*, Drum sticks, Kjheldal's method, Flame Photometry and Atomic Colorimetry, Absorption Spectrophotometer.

I. INTRODUCTION:

There are a number of insightful evaluations available on *M. oleifera*'s ethno botanical applications. It has been discovered that *Moringa* is a good source of antioxidants and polyphenols. There have been reports of phytochemicals in its roots, flowers, fruits, and seeds, including quercetin, beta-sitosterol, beta-sitosterol, ascorbates, vanillin, and omega fatty acids²¹. Particularly in the leaves, phenolic and flavonoids have been discovered; these substances possess a range of biological properties, such as immunomodulatory, anti-diabetic, anti-carcinogenic, hepato-protective, and thyroid status control¹⁸. Furthermore, trace components necessary for human health can be found in leaves¹⁷. For example, trace elements such as iron, zinc, magnesium, and selenium are critical for metabolism, and claims of a link

between oxidative illnesses and trace element status are driving interest in these elements. On the other hand, a recent study found that dried *M. oleifera* leaves have extremely high levels of lead—352.0 mg/L. Determining the mineral makeup of *M. oleifera* leaves—which are commonly eaten by both people and animals—is therefore crucial⁶.

Furthermore, trace components necessary for human health can be found in leaves. For example, trace elements such as iron, zinc, magnesium, Arsenic and selenium are critical for metabolism, and claims of a link between oxidative illnesses and trace element status are driving interest in these elements¹⁶. On the other hand, a recent study found that dried *M. oleifera* leaves have extremely high levels of lead—352.0 mg/L. Determining the mineral makeup of *M. oleifera* leaves—which are commonly eaten by both people and animals—is therefore crucial.

II. MATERIALS AND METHODS



After washing, the *M. oleifera* seeds were allowed to air dry. For the one-gram sample of *M. oleifera*, ash was created separately in a silica crucible, and it was maintained at 600 degrees Celsius in a muffle furnace until the crucible's weight remained constant. Through qualitative and quantitative examination, the main components of ash were identified. For the purpose of determining the metal content, ash was dissolved in 5.0 millilitres of 10% HCl to turn it into chloride. The treated ash was then filtered through Whatmann filter paper No. 40.

AgNO₃ and then cleaned in hot water. Using gravimetric analysis, the percentage of ash that was acid soluble and insoluble was determined. Using Kjeldahl's approach and a Perkin Elmer 3110 Atomic Absorption Spectrophotometer, a 100 ml filtrate was used for the quantitative measurement of metals. Estimates of nitrogen and phosphorus were made using a photometric colorimeter made by Systronics. The Madiflame model 1278 flame photometer was used to measure metals belonging to the first and second groups, namely the alkali group and the alkaline earth metal. The outcomes were displayed in Tables Nos. 1 and 2.

Sr. No.	Particulars	Seeds	Flowers
1.	Acid soluble ash	4.90 %	3.73%
2.	Acid insoluble ash	0.21 %	0.11%

Table 1: Gravimetric analysis of samples

Sr. No.	Element	Method	Percentage	
			Seed	Flower
1	Mn	Atomic Absorption Spectrophotometer	123.7 ppm	10 ppm
2	Fe	Atomic Absorption Spectrophotometer	223.6 ppm	122 ppm
3	Zn	Atomic Absorption Spectrophotometer	218.1 ppm	45.0 ppm
4	Cu	Atomic Absorption Spectrophotometer	14.10 ppm	121.0 ppm
5	Pb	Atomic Absorption Spectrophotometer	200 ppm	310 ppm
6	Ca	Atomic Absorption Spectrophotometer	0.077%	15 %
7	As	Atomic Absorption Spectrophotometer	0.02%	0.04%
8	K	Flame Photometry	249 ppm	255 ppm
9	P	Colorimetric method	0.05%	0.09%
10.	N	Kjheldal's method	0.40 %	0.34%

Table 2: Elemental analysis by various techniques

III. RESULTS AND CONCLUSION:

Activating substances facilitate a multitude of enzymatic reactions; these agents are often relatively small-concentration elements. Herbs are divided into two categories based on the amount of minerals they contain: sedatives and stimulants. Stimulatory herbs include larger percentages of potassium, iron, and phosphorus, whereas sedative herbs are thought to be those with high calcium and magnesium content. Common elements like calcium, potassium, iron, phosphorus, manganese, zinc, and copper were found in the seeds and flowers of *M. Oleifera*.

It was discovered that while calcium, phosphorus, and nitrogen are minor components, potassium, zinc, manganese, and iron are present in significant amounts. The *M. oleifera* sample is recommended in traditional and folk medicines due to its richness in manganese, iron, zinc, and potassium. Potassium's effect on insulin secretion makes it an essential component of diabetic treatment. A zinc deficiency manifests as disease, stress, elevated thyroid activity, hyper adrenal hyperplasia, skin lesions, stretch marks, delayed wound healing, and nail spots.

The maximum allowable amount of Arsenic is exceeded in the samples, although the presence of dangerous elements like lead and arsenic in *M. oleifera* flower seems to be within acceptable bounds. According to our findings, plants meant for human food require harmful metal monitoring and quality assurance.

IV. REFERENCES:

1. Chopra R. N. Nayar S. L. and Chopra R. C. Glossary of Indian Medicinal Plants (Including the Supplement) Council of Scientific and Industrial Research, p.11, New Delhi, 1986
2. Duke J. A. Handbook of Energy Crops, 269, 1983.
3. Bown D. Encyclopedia of Herbs & their uses., Darling Kinderley, London (1995) ISBN O-7513 - 020-31
4. Duke J.A. and Ayensu E. S., Medicinal Plants of China Reference Publications, Inc. (1985) ISBN O-917256-20-4
5. D.L.Watts, Trace Elements and Other Essential Nutrients, Clinical Application of Tissue Mineral Analysis, Writer's B-L-O-C-K Edition, USA,1997.
6. M.Hidiroglu and J. E. Knipfel, J. Dairy Sci.67, 1141, 1984.
7. N.P. Singh and S. Karthikeyan, Flora of Maharashtra State, Dicotyledones, 1, pp.175, 2000.
8. M. Marthanda Murthy, M.Subramanyam, M. Hima Bindu and Annapurna, Antimicrobial activity of Clerodane Di terpenoids from *Polyalthia longifolia* seeds, Fitoterapia, 76 (3-4), pp.336-339, 2005.
9. Jayaveera K.N.; Sridhar C.; Kumanan R.; Yogananda Reddy K.; Tarakaram K.; Mahesh M., Phytochemical, antibacterial and anthelmintic potential of flowers of *Polyalthia longifolia*, Journal of Pharmacy and Chemistry, 4(2), pp. 66-69, 2010.
10. Sashidhara Koneni V., Singh Suriya P., Shukla P.K., Antimicrobial evaluation of clerodane diterpenes from *P. longifolia* var.pendula, Natural Product Communications, 4(3), pp. 327-330, 2009.
11. Shaheen Khan,; Rashid Ali,; Azher Soobia; Khan, Shajeel A hmed; T auseef, Saima; Ahmad Aqeel, Plant Medica, 69 (4),pp. 350-355, 2003.
12. Dr. C.K.Kokate, textbook of Pharmacognosy, 29 Ed. Nirali Prakashan, Pune, 108-109, 2004
13. Doss, M.Pugalenth, D. R agendrakumaran and V. Vadivel, Phenols, Flavonoids, and Antioxidant activity of underutilized legume seeds, ASIAN J. EXP.BIOL. SCI., 1(3), pp. 700-705, 2010.
14. Dr. C.K.Kokate , Practical Pharmacognosy, 4 Ed. Vallabh prakashan, New Delhi, 107-11, 2008.
15. Evans R and Collins P, The wealth of India-Raw Materials, 5th ed., New Age International Publishers Ltd., New Delhi, 55-57, 2005
16. Dr. S. Ravi Shankar, Text book of Pharmaceutical Analysis, Page no: 15.8, 2010.
17. Vijyalakshmi R, Ravindran R. Preliminary comparative phytochemical screening of root extracts of *Diospyros ferrea* (Wild.) Bakh and *Arva lanata* (L.) Juss. Ex Schultes. Asian J Plant Sci Res; 2:581-587, 2012
18. Doss A. Preliminary phytochemical screening of some Indian medicinal plants. Anc Sci Life 2009;

29:12-16.

19. Pandey P, Mehta R, Upadhyay R. Physico-chemical and preliminary phytochemical screening of *Psoralea corylifolia*. Arch Appl Sci Res 2013; 5:261-265
20. Yadav M, Chatterji S., Gupta S. K. and Watal G., "Preliminary Phytochemical Screening Of Six Medicinal Plants Used In Traditional Medicine" International Journal of Pharmacy and Pharmaceutical Sciences, 2014; Vol 6, Issue 5, ISSN- 0975-1491.
21. Oliveira I, Sousa A, Ferreira I, Bento A, Estevinho L, Pereira JA. Total phenols, antioxidant potential and antimicrobial activity of walnut (*Juglans regia* L.) green husks. Food Chem Toxicol 2008; 46: 2326-2331.
22. Nair VD, Paneerselvam R, Gopi R. Studies on methanolic extract of *Rawolfia* species from Southern Western Ghats of India - In vitro antioxidant properties, characterization of nutrients and phytochemicals. Ind Crop Prod 2012; 39: 17-25.

