



# Evaluation of some botanical plant extracts for their antifungal activity against *Alternaria brassicae* of cabbage

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## Abstract

Black leaf spot disease caused by *Alternaria brassicae* (Berk.) Sacc. It is a serious disease of Indian crucifers plants throughout the country. Extract of 5 plants Sargandha (*Rauvolfia serpentina*), Ashoka (*Saraca asoca*), Amaltash (*Cassia fistula*), Karanj (*Pongomia pinnata*) and Badoh (*Ipomoea carnea*) belonging to 3 families was tested against their efficacy at 25% and 50% concentration in vitro by poisoned food technique. The five extracts found effective in vitro were evaluated on cabbage as pre and post inoculation treatments. All the plant extracts at 50% concentration resulted in significant disease reduction. But maximum reduction was observed with sargandha extract followed by amaltash.

**Keywords:** Cabbage, *Alternaria brassicae*, antifungal activity, medicinal plant extracts, poisoned food technique

## Introduction

Cabbage (*Brassica oleracea* L. Capitata) is leafy green plant, it is an excellent source of vitamin K, vitamin C, vitamin B6, vitamin B1 (thiamine), B2 (riboflavin), B3 (niacin), B5 (pantothenic acid), B9 (folate) vitamin-A creates cooling effect in the body, prevents from constipation, helps in digestion and is useful for diabetes. It is also a very good source of manganese, calcium, iron, manganese, phosphorus, potassium, sodium, zinc (FAO, 2023). India is the second largest producer of cabbage in the world with a production of 9035 thousand tones from an area of 402 thousand hectares. West Bengal is the leading producer of cabbage in India but the productivity is highest in U.P. (Anonymous, 2023). Cruciferous plant are severely affected by fungi, bacteria, nematodes and viruses. Leaf spot disease caused by *Alternaria brassicae* (Berk) Sacc. And *A. brassicicola* has been reported from all the continents of the world. Losses up to 30% and 47% were caused by *Alternaria brassicae* in cauliflower (*Brassica oleracea* var. *botrytis*) (Tomayo *et al.* 2001). Black leaf spot disease in cabbage (*Brassica oleracea* L.) is caused by *Alternaria brassicae* (Berk) Sacc. Although this is a common disease of cruciferous vegetables worldwide. The pathogen is easily spread through the contamination of seeds. The leaves and stems of cabbages infected by *A. brassicae* result in the formation of dark-brown, necrotic lesions with a yellow halo; this damage reduces both yield and economic value. Therefore the different botanical were applied to control *Alternaria brassicae* causing leaf spots in cabbage.

## Materials and methods

### Experimental site and sample collection

Botanical leaf extract used for this experiment were prepared indigenously. Fresh leaves of Sarp Gandha (*Rauwolfia serpentina*), Ashoka (*Saraca asoca*), Amaltash (*Cassia fistula*), Karanj (*Pongamia pinnata*) and Badoh (*Ipomoea carnea*) were collected from the different area of Prayagraj U.P. The experimental site located at 20° 30' North latitude and 85° 49' 60" east longitude city and is about 98m above sea level. The city is situated at the confluence of three rivers- Ganga, Yamuna and the invisible Saraswati. The sample of infected leaf were collected from the experimental field for the study.

### Fungal and Plant material

*Alternaria brassicae* isolated from the leaves of cabbage (*B. oleracea* L.) showing symptoms of black leaf spot disease. Which was isolated from diseased cabbage (*A. brassicae*). The cultures were grown on a potato dextrose agar (PDA) (200gm of potato infusion, 20gm of glucose and 20gm of agar in 1L of distilled water).

### Isolation and morphological study of the pathogen

The leaf tissues of the plant exhibiting typical symptoms of black leaf spot were selected to retrieve the pure culture of the pathogen. Cut the small piece of the infected leaf and surface sterilized with the sodium hypochlorite solution (NaOCl) of the 4.0% concentration for 3min. and followed by three washing with the sterile distilled water. The water was removed with the help of sterile blotting paper and thereafter the tissues were transferred on the sterile PDA plates and kept on incubation at 25+2°C in BOD incubator. The fungal mycelium growth observed after 72h of incubation was transferred on the other PDA plate for pure culture. The morphological and in vitro efficacy of the fungicides was performed from the seven days old culture of the fungus. (Begum *et al.* 2010)

### Preparation of botanical leaf extract-

Fresh botanical leaf extract was prepared by grinding the required quantity of leaves (100gm) before grinding equal quantity of water was added in the respective plant leaves (1:1 weight/ volume basis). The fresh botanical leaf extract was filtered different leaves was sieved through muslin cloth. The filtered extracts were used @ 25% and 50% concentration by adopting poisoned food technique. (Sasode *et al.* 2012)

### Bioassay of plant extracts by poisoned food technique-

The experiment was conducted for measurement of mycelium growth inhibition following the poisoned food technique (Sharvelle 1961). The standardized concentration of the fungicides and botanical extracts were mixed in the sterilized PDA and amended PDA poured in the sterilized petri plates. The petri plates of solidified PDA inoculated with a 7mm mycelium disk cut with the sterile cork borer from the 7 days old fungal culture. The inoculated plates were placed at 25+2°C in BOD incubator for incubation. Each treatment was replicated four and the growth of fungus colony was measured after 7 days of inoculation. The percent inhibition of the mycelium growth over the control was calculated as;

$$PI = \frac{C - T}{C} \times 100$$

where

PI = Per cent inhibition of fungal growth

C = Mycelium growth of the fungus in absent of treatment

T = Mycelium growth of the fungus with treatment.

## Result and discussion

### Effect of plant extracts against *Alternaria brassicae* in vitro

The 5 plants fresh leaf extracts namely sarp Gandha (*Rauvolfia serpentina*), Ashoka (*Saraca asoca*), Amaltash (*Cassia fistula*), Karanj (*Pongomia pinnata*) and Badoh (*Ipomoea carnea*) belonging to 3 families were tested for their efficacy at 25% and 50% concentration against the *A. brassicae*. The experiment was conducted in in-vitro conditions to assay their effect on mycelial radial growth of pathogen.

### Effect on mycelial radial growth

The effect of botanical on the mycelia growth of *Alternaria brassicae* are presented in the (Table -1). All the botanicals tested were effective showing different levels of toxicity against *Alternaria brassicae* inhibiting the growth of pathogen.

At 25% mycelial radial growth of the test pathogen was ranged from T1 sarp Gandha (*Rauvolfia serpentina*)(21.450mm) to T3 Amaltash (*Cassia fistula*)(40.125mm) however, it was maximum with

T3 Amaltash (40.125mm). This was followed by T2 Ashoka (*Saraca asoca*)(38.575mm), two of which were at par and T4 Karanj (*Pongomia pinnata*) (25.80mm), T5 Badoh (*Ipomoea carnea*) (24.325mm) of which were at par. Comparatively least mycelia growth was recorded with the botanical viz., T1 sarp Gandha (*Rauvolfia serpentina*)(21.450mm) and T5 Badoh (*Ipomoea carnea*) (24.325mm)

At 50% radial mycelia growth of the test pathogen was ranged from T1 sarp Gandha (*Rauvolfia serpentina*) (12.175mm) to T3 Amaltash (*Cassia fistula*) (20.675mm) however, significantly highest mycelial growth was maximum with T3 Amaltash (*Cassia fistula*) (20.675mm). This was followed by T2

Ashoka (*Saraca asoca*)(19.725) T4 Karanj (*Pongomia pinnata*) (15.950), T5 Badoh (*Ipomoea carnea*) (13.300). Significantly least mycelia growth was recorded with the botanical T1 sarp Gandha (*Rauvolfia serpentina*) (12.175mm) as compared to untreated control was 90.00mm.

### Mycelial growth inhibition

Results (Table - 1) revealed that all the plant extracts tested (at 25 and 50% each) significantly inhibited mycelial growth of the test pathogen over untreated control (0.00%) further, it was found that percent mycelia growth inhibition of the test pathogen was increased with increase in concentration of the botanicals tested.

At 25% mycelia growth inhibition was ranged from T3 Amaltash (*Cassia fistula*) (45.41) to T1 sarp Gandha (*Rauvolfia serpentina*) (66.16) percent. However, significantly highest mycelial growth inhibition was recorded with T1 sarp Gandha (*Rauvolfia serpentina*) (66.16). This was followed by T5

T5 Badoh (*Ipomoea carnea*)(62.97), T4 Karanj (*Pongomia pinnata*) (61.33), T2 Ashoka (*Saraca asoca*)(47.13) significantly least mycelia growth inhibition was recorded with T3 Amaltash (*Cassia fistula*) (45.41%) over untreated control (0.00%).

At 50% concentration growth inhibition was ranged from T3 Amaltash (*Cassia fistula*) (67.02%) to T1 sarpgandha (*Rauvolfia serpentina*) (76.47%) However significantly highest mycelial growth inhibition was recorded with T1 sarpgandha (*Rauvolfia serpentina*) (76.47%). This was followed by T5 Badoh (*Ipomoea carnea*) (75.22%), T4 Karanj (*Pongomia pinnata*) (72.27%), T2 Ashoka (*Saraca asoca*)(68.08%). Significantly least mycelial growth inhibition was recorded with T3 Amaltash (*Cassia fistula*) (67.02%) over untreated control (0.00%). A lot of work has been reported regarding the botanical extracts against *Alternaria* spp. From different places in different years (Shekhawat and Prasad. 1971; Sheikh and Agnihotri. 1972; Singh *et al.*1993; Tripathi and Shukla. 2002; Bhalona *et al.* 2011; Sitara *et al.* 2008; Sharma *at al.*2007; Kumar *et al.* 2006; Meena *et al.* 2010; Patni and Kolte. 2006; Siva *et al.* 2008; Sundar *et al.* 2010; Yanar *et al.*2011; Singh *et al.* 2013; Singh *et al.*2014; Sasode *et al.* 2012; Zaker , M. 2013; Satish *et al.* 2007; Yadav *et al.* 2019; Meena *et al.* 2020; )

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**Table-1 : In-vitro effect of different plant extracts on mycelia growth and % inhibition of *A. brassicae*.**

Treatment	Colony diameter (mm) at conc.		% inhibition at conc.	
	25%	50%	25%	50%
T <sub>1</sub> Sarpgandha ( <i>Rauvolfia serpentina</i> )	21.450 d	12.175 f	66.16	76.47
T <sub>2</sub> Ashoka ( <i>Saraca asoca</i> )	38.575 b	19.725 c	47.13	68.08
T <sub>3</sub> Amaltash ( <i>Cassia fistula</i> )	40.125 b	20.675 b	45.41	67.02
T <sub>4</sub> Karanj ( <i>Pongomia pinnata</i> )	25.800 c	15.950 d	61.33	72.27
T <sub>5</sub> Badoh ( <i>Ipomoea carnea</i> )	24.325 c	13.300 e	62.97	75.22
T <sub>0</sub> Control	90.00 a	90.00 a		
F- test	S	S		
CD (0.05)	2.15	0.77		

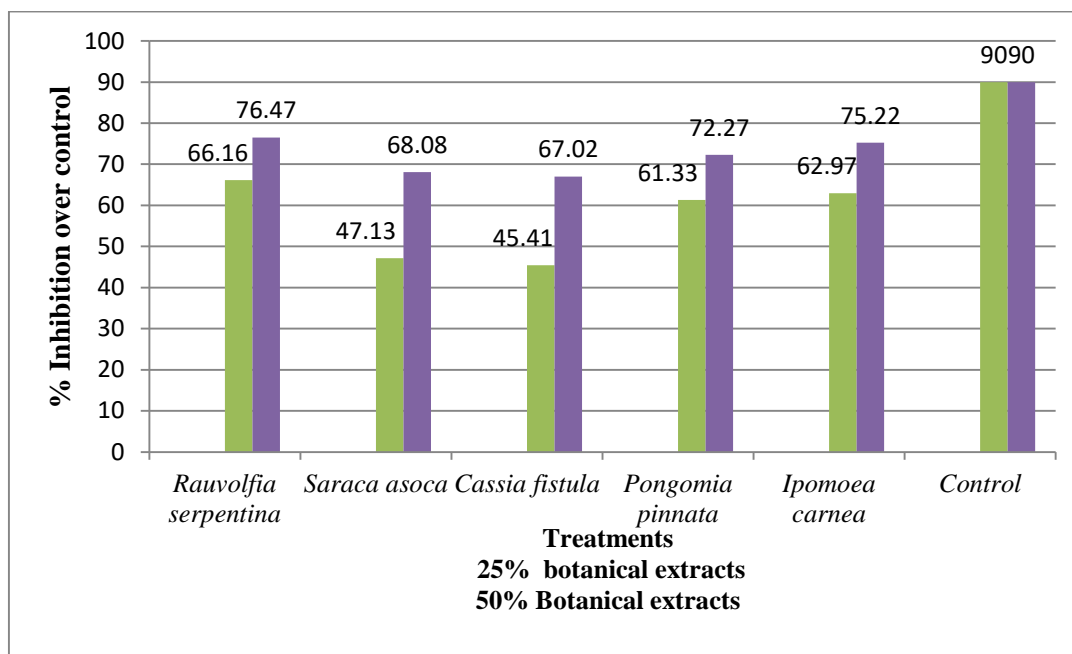


Figure 1: *In- vitro* evaluation of different botanicals against *Alternaria brassicae*

## References

**Anonymous (2023).** Ministry of Agriculture, Government of India.

**Begum, M. F., Mahal, M. F. and Alam, M. S. (2010).** Inhibition of spore germination and mycelial growth of three fruit rot pathogens using some chemical fungicides and botanical extracts. *Journal of Life and Earth Science*, 5: 23-27.

**Bhalodia, N. R., Nariya, P.B. and Shukla, V.J. (2011).** Antibacterial and antifungal activity from flower extracts of *Cassia fistula* L. an ethnomedicinal plant. *International Journal of Pharm. Tech. Research*, 3(1): 160-168.

**FAO (2023).** Statistics Database. Food and Agriculture Organization of the United Nations.

**Kumar, N., Kumar, A. and Sugha, S.K. (2006).** Evaluation of bioagents and plant extracts against *Alternaria* blight of rapeseed mustard. *Plant Disease Research*, 21(1): 48-50.

**Meena, P.D., Awasthi, R.P., Chattopadhyay, C., Kolte, S.J. and Kumar, A. (2010).** *Alternaria* blight: a chronic disease in rapeseed-mustard. *Journal of Oilseed Brassica*, 1(1): 1-11.

**Meena, P.D., Sharma, P. (2012).** Antifungal activity of plant extracts against *Alternaria brassicae* causing blight of *Brassica* spp. *Ann. Pl. Protec. Sci.* 20 : (1) 256-257.

**Meena, R.P., Saran, P.L., Kalariya, K.S. and Manivel, P. (2020).** Efficacy of fungicides and plant extracts against *Alternaria alternate* causing leaf blight of chandrasur (*Lepidium sativum*). *Indian Journal of Agricultural Sciences* 90 (2): 337-40

**Patni, C.S. and Kolte, S.J. (2006).** Effect of some botanicals in management of *Alternaria* blight of rapeseed mustard. *Annals of Plant Protection Sciences*, 14(1):151-156.

**Sasode, R.S., Prakash, S., Gupta, A., Pandya, R.K. and Yadav, A. (2012).** In vitro study of some plant extracts against *Alternaria brassicae* and *Alternaria brassicicola*. *Journal of Phytology.*, 4(1): 44-46

**Satish, S., Mohana, D.C., Raghavendra, M.P. and Raveesha, K.A. (2007).** Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. *Journal of Agricultural Technology*, 3(1): 109-119.

- Sharma, A., Dass, A. and Pau, M.S. (2007).** Antifungal effect of neem extract on some common phytopathogenic fungi. *Adv. Plant Sci.*, 20(2): 357-358.
- Sheikh, R.A. and Agnihotri, J.P. (1972).** Antifungal properties of some plant extracts. *Indian Journal of Mycology and Plant Pathology*, 2: 143-146.
- Shekhawat, P.S, Prasad, R. (1971).** Antifungal properties of some plant extracts in inhibition of spore germination. *Indian Phytopath.* 24:800-802.
- Singh, G., Gupta, S. and Sharma, N. (2014).** *In vitro* screening of selected plant extracts against *Alternaria alternata*. *Journal of Experimental Biology and Agricultural Sciences*, 2(3): 344-351.
- Singh, H.N., Prasad, M.M. and Sinha, K.K. (1993).** Efficacy of leaf extracts of some medicinal plants against disease development in Banana. *Letters in Applied Microbiology*, 17(6): 269-271.
- Singh, S., Godara, S.L. and Gangopadhyay, S. (2013).** Studies on antifungal properties of plant extracts on mustard blight caused by *Alternaria brassicae*. *Indian Phytopathology*, 66(2): 172-176.
- Sitara, U., Niaz, I., Naseem, J. and Sultana, N. (2008).** Antifungal effect of essential oils on *in vitro* growth of pathogenic fungi. *Pakistan Journal of Botany*, 40(1): 409-414.
- Siva, N., Ganesan, S., Banumathy, N. and Muthuchelian (2008).** Antifungal effect of leaf extract of some medicinal plants against *Fusarium oxysporum* causing wilt disease of *Solanum melogena* L. *Ethnobotanical Leaflets*, 12: 156-163.
- Sunder, S., Ram, S. and Dodan, D.S. (2010).** Evaluation of fungicides, botanicals and non – conventional chemicals against brown spot of rice. *Indian Phytopathology*, 63(2): 192-194.
- Tamayo, M.P.J., Becerra, V.D.C. and Jaramillo, N.J.E. (2001).** *Alternaria brassicae*, agente causal de pudricion de la cabeza en coliflor (*Brassica oleracea* L. var. botrytis). *Ascolfi Informa*, 27: 10-11.
- Tripathi, A. K. and Shukla, B. N. (2002).** Antifungal activity of some plant extracts against *Fusarium oxysporum* causing wilt of linseed. *Journal of Mycology and Plant Pathology*, 32: 266-267.
- Yadav, J.K., Singh, H.K., Singh, S.K., Kavita and Singh, S. (2019).** Efficacy of plant extracts against *Alternaria brassicae* under *in-vitro* condition. *Journal of Pharmacognosy and Phytochemistry*, 8(1): 528-532
- Yanar, Y., Gökçe, A., Kadioglu, I., Çam, H. and Whalon, M. (2011).** *In vitro* antifungal evaluation of various plant extracts against early blight disease (*Alternaria solani*) of potato. *African Journal of Biotechnology*, 10(42): 8291-8295.
- Zaker, M. (2013).** Screening some medicinal plant extracts against *Alternaria sesami*, the causal agent of *Alternaria* leaf spot of Sesame. *Journal of Ornamental and Horticultural Plants*, 3(1): 1-8.