



SYNERGISTIC ANTICANCER ACTIVITY OF CATHARANTHUS *ROSEUS* LINN. AND SOLANUM *LYCOPERSICUM* LINN.

Divya Tiwari, Abhishek Yadav, Prankul Yadav, Shivam Yadav, Ashish Pal

Harishchand Pharmacy College

ABSTRACT

Breast cancer is a condition where malformed breast cells proliferate and develop into tumours. Tumours can become deadly if they proliferate throughout the body and are not treated. So, this study explores the synergistic anti-cancer activity derived from the combination of *Catharanthus roseus* (commonly known as Madagascar periwinkle), a plant source of vinblastine and vincristine, potent anti-cancer alkaloids, with *Solanum lycopersicum* (tomato), a rich reservoir of lycopene, a recognized anti-cancer pigment. The rationale behind this investigation lies in the potential enhancement of anti-cancer effects through the complementary action of these bioactive compounds.

The research involves extraction of *Catharanthus roseus* and *Solanum lycopersicum* in respective solvents. Subsequently, combined ratio of both plant extract are subjected to evaluate for synergistic activity on breast cancer cell lines. Preliminary findings indicate a notable increase in anti-cancer activity when combined chemical constituent from both extract which is more prone for anticancer activity, suggesting a synergistic interaction. This synergy may be attributed to complementary mechanisms of action to achieve hypothesis. Combination therapy, a treatment modality that combines two or more therapeutic agents, is a cornerstone for cancer therapy. The amalgamation of anti-cancer drugs enhances efficacy compared to the mono-therapy approach because it targets key pathways in a characteristically synergistic or an additive manner. This approach potentially reduces drug resistance, while simultaneously providing therapeutic anti-cancer benefits, such as reducing tumour growth and metastatic potential, arresting mitotically active cells, reducing cancer stem cell populations, and inducing apoptosis.

Keywords: *Catharanthus Roseus Linn.*, *Solanum Lycopersicum Linn.*, lycopene, vincristine, vinblastine, synergistic anticancer activity

INTRODUCTION:

Cancer remains a formidable health challenge worldwide, with a growing number of patients seeking effective therapeutic interventions. Increased incidence of cancer in recent years and its impact on different physical, mental, and social dimensions of human life have turned it to a major problem of the century. The incidence of this disease in developed countries varies from 1 to 2 percent, with almost 5% yearly increase in less developed countries. Meanwhile, breast cancer is the most prevalent type of malignant neoplasm among women with more than one million new cases per year. In India, breast cancer accounts for the major type of cancer among women with the incidence of 21.4 or 32%. Breast cancer is the most common type of cancer among women in the US with the incidence rate of 12.5%. The risk of an individual dying from breast cancer is 1-in-35. Regarding the importance of this issue, this study sought to investigate breast cancer and its associated factors [1].

Breast cancer is the most common type of cancer and the second leading cause of death. This disease is the primary cause of mortality among women aged 45–55 years, and is the second leading cause of cancer-induced death. The incidence of breast cancer is almost 1-in-8 women, requiring complete tissue removal,

chemotherapy, radiotherapy, and hormone therapy most of the time. Breast cancer is a type of tissue cancer that mainly involves inner layer of milk glands or lobules, and ducts (tiny tubes that carry the milk) [2]. The primary risk factors of cancer include age, high hormone level, race, economic status, and iodine deficiency in diet. Breast cancer is a multi- stage disease, in which viruses play a role in one stage of this pathogenic process. In general, viruses are involved with different cancer types [3].

Medicinal plants have been traditionally used in folk medicine for centuries as natural healing remedies with significant proven therapeutic effects in many areas including prevention of anticancer activity, cardiovascular diseases, anti-inflammatory, antimicrobial activity. In addition, the emergence of resistance to cancer chemotherapy has forced researchers to turn to natural products of plant and marine origin

[4]. Although many compounds isolated from plants are being strictly tested for their anticancer properties, it is recognized that the beneficial effects of combined plant extract due to a complex interplay of the composite mixture of chemical constituent present in the more than one plant (additive/synergistic and/or antagonistic) rather than single plant extract [5].

In this context, the present research investigates the synergistic anticancer potential of two potent botanical agents, *Catharanthus roseus* Linn. and *Solanum lycopersicum* Linn., on breast cancer cells [6]. The primary objective of this study is to explore the synergistic effects resulting from the combination of these two plant-derived compounds, shedding light on their collective efficacy against breast cancer [7]. Despite the escalating prevalence of cancer, there exists a research gap concerning the simultaneous application of *Catharanthus roseus* and *Solanum lycopersicum* as anticancer agents, making this investigation particularly significant [8].

The study focuses on breast cancer cells, emphasizing the need for novel therapeutic approaches to address this prevalent form of cancer [9]. Through rigorous experimentation and analysis, we aim to contribute valuable insights into the synergistic mechanisms underlying the combined anticancer activity of these botanical agents (*Catharanthus roseus* Linn. and *Solanum lycopersicum* Linn.) [10].

Plant Profile:

a) ***Solanum lycopersicum* Linn.: Taxonomic Classification: [11, 12]**

- Kingdom: Plantae
- Subkingdom: Tracheobionta
- Division: Magnoliophyta
- Class: Magnoliopsida
- Subclass: Asteridae
- Order: Solanales
- Family: Solanaceae
- Genus: Solanum
- Species: *Solanum lycopersicum* Linn.



Figure1:*Solanum lycopersicum* Linn.

Chemical Constituents: [13, 14] Carotenoids such as Lycopene, Phytoene and Beta-carotene, Rich source of Vitamin C, Vitamin E, Luteine.

***Solanum lycopersicum* Linn. as an anticancer: [15]**

Solanum lycopersicum Linn. contains lycopene pigment. Lycopene caused tumor cells to accumulate in the G0/G1 phase and undergo apoptosis. However, most cancer cells presented an increase in G2/M phase as the incubation time was prolonged. Several studies suggested lycopene decreased cell cycle related proteins, such

as cyclin D1, D3 and E, the cyclin dependent kinases 2 and 4, bcl-2, while decreased phospho-Akt levels

b) *Catharanthus roseus* Linn.: Taxonomic Classification: [16, 17]

- Kingdom: Plantae
- Division: Magnoliophyta
- Class: Magnoliopsida
- Order: Gentianales
- Family: Apocynaceae
- Genus: Catharanthus
- Species: *Catharanthus. roseus* Linn.

Chemical Constituents: [18, 19]



Figure 2: *Catharanthus roseus* Linn

because of none separation of chromosomes Vincristine, Vinblastine, Vindoline, Ajmalicine, Vincine, Vincamine, Raubasin, Reserpine, Catharanthine, Vindesine, Vindeline, Tabersonine

***Catharanthus roseus* Linn. as an Anticancer: [20, 21]**

Vinblastine inhibits the cell cycle of cancer cells. It binds with tubulin, and inhibits the formation of microtubules. Due to the inhibition, formation of microtubules takes place and cell cycle arrest in M phase

during anaphase of mitosis. The chemical structure of vinblastine and vincristine are very similar, but their effects are not the same. Vinblastine is used to treat specific types of cancer, such as Hodgkin's disease, and Vincristine is used in the treatment of acute lymphoblastic leukemia.

MATERIALS AND METHOD:

- ***Solanum lycopersicum* Linn. (Sample Code - TL):** Collection: We have collected fruit of *Solanum lycopersicum* Linn. from Deur, Taluka-Koregoan, District- Satara.
- Pulp formation: The pulp is made by crushing fruits into crude tomato juice by using mixure [22].
- Drying: The above pulp was taken in petridish and dried in hot air oven at 50-60°C for two days.
- Powdering: The pulp was reduced into fine size.
- Soxhlation: The fruit powder was subjected for extraction by using methanol at 55°C. (For 8 Hours daily for two days) [23- 26]
- Drying: We took 100 gm activated Magnesium sulphate as adsorbent, and placed into dessicator to dry the extract [27].

B) *Catharanthus roseus* Linn. (Sample Code - VV):

- Collection: Leaves of *Catharanthus roseus* Linn. was collected.
- Drying: Leaves were shed dried for one month.
- Powdering: Dried leaves were finely grinded into powder.
- Soxhlation: The leaves powder was subjected for extraction by using methanol at 55°C. (For 8 Hours daily for five days) [28, 29]
- Drying: We took 100 gm activated Magnesium sulphate as adsorbent, and placed into dessicator to dry the extract.

C) Mixing of both extract (Sample code - VT):

Above Both extract mixed in 1:1 proportion and send for testing.

Table 1

Sample Code	Name of Extract
VV	<i>Catharanthus roseus</i> Linn.
TL	<i>Solanum lycopersicum</i> Linn.
VT	Mixture of both (1:1)



Figure 3: Collection of Plant



Figure 4: Soxhlation (VV)



Figure 5: Soxhlation (TL)



Figure 6: Drying of Sample VV



Figure 7: Drying of Sample TL

Phytochemical Analysis: [30-35]

A) Test for Glycosides:

1. **Keller killani test:** (Cardiac glycosides) the test solution with few drops of glacial acetic acid in 2 ml of ferric chloride solution and concentrated sulphuric acid is added from the sides of the test tube which shows the separation between two layers, lower layer shows reddish brown and upper layer turns bluish green in colour.
2. **Raymond's test:** Test solution treated with dinitrobenzene in hot methanolic alkali gives violet color.
3. **Legal's test:** The test solution treated with 1ml pyridine and 1ml sodium nitroprusside gives pink to red color appears.

B) Test for Alkaloids:

1. **Mayer's test:** Test solution treated with Mayer's reagent (Potassium mercuric iodide) gives cream colored precipitate.
2. **Wagner's test:** The acidic test solution treated with Wagner's reagent (Iodine in potassium iodide) gives brown precipitate.
3. **Hager's reagent:** The acetic test solution treated with Hager's reagent (Saturated picric acid solution) gives yellow precipitate.

C) Test for Flavanoids:

1. **Ferric chloride test:** The test solution with few drops of ferric chloride solution shows intense green color.

2. **Shinoda test:** Test solution with few fragments of magnesium ribbon and concentrated hydrochloric acid shows pink to magenta red color.
3. **Zinc - Hydrochloric acid- reduction test:** Test solution with zinc dust and few drops of hydrochloric acid shows magenta red color.
4. **Alkaline reagent test:** Test solution when treated with sodium hydroxidesolution shows increase in the intensity of yellow color which becomes colorless on addition of few drops of dilute acid.
5. **Lead acetate solution test:** Test solution with few drops of lead acetate solution (10% w/v) gives yellow precipitate.

B) Test for Steroids:

1. **Liebermann burchard test:** The test solution treated with few drop of acetic anhydride and mixed well. When concentrated sulphuric acid is added from the side of the test tube, it shows a brown ring at the junction of the two layer and the upper layer turns green.
2. **Salkowski's Test:** The second portion of solution above was mixed with concentrated sulphuric acid carefully so that the acid formed a lower layer and the interface was observed for a reddish-brown color indicative of steroidring.

C) Test for Phenols:

1. **Ferric Chloride Test:** A small amount of the ethanolic extract was taken with 1 ml of water in a test tube and 1 to 2 drops of Iron III chloride (FeCl₃) was added. A blue, green, red or purple color is a positive test.
2. **Lead acetate Test:** Plant extract is dissolved in 5mL distilled water + 3mL of 10% lead acetate sol. A white precipitate.
3. **Ellagic Acid Test:** Plant extract aqueous solution + 5% glacial acetic acid + 5% sodium nitrite solution A Solution turns muddy / Niger brown precipitate.
4. **Gelatin Test:** Plant extract is dissolved in 5 mL distilled water +1% gelatine solution + 10% NaCl A white precipitate.

D) Test for Caretenoids:

1. **Caretenoid Test:** 1 g of each specimen sample was extracted with 10ml of chloroform in a test tube with vigorous shaking. The resulting mixture was filtered and 85 % sulphuric acid was added. A blue colour at the interface showed the presence of carotenoids.

Pharmacological Activity: [36]

The three different extract was send to Infinite Biotech Laboratory, Sangli for invitro anticancer activity.

a) Cell line:

MCF7 (Breast Cancer cell line)

b) Media:

DMEM with high glucose (Cat No- 11965-092), FBS (Gibco, Invitrogen)

Cat No-10270106 Antibiotic- Antimycotic 100X solution (Thermo fisher Scientific) Cat No-15240062Experimental Procedure (MTT Assay): [37-40]

1. Cells were incubated at a concentration of 1×10^4 cells/ml in culture medium for 24 h at 37°C and 5% CO₂.
2. Cells were seeded at a concentration (100 μ l) 104 cells/well) in 100 μ l culture medium and 20, 40, 60, 80, 100 μ g/ml of Samples into micro plates respectively (tissue culture grade, and 96 wells).
3. Control wells were incubated with DMSO (0.2% in PBS) and cell line. All samples were incubated in triplicate. Controls were maintained to determine the control cell survival and the percentage of live cells after culture.
4. Cell cultures were incubated for 24 h at 37°C and 5% CO₂ in CO₂ incubator.
5. After incubation, the medium was completely removed and Added 20 μ l of MTT reagent. (5mg/min PBS).
6. After addition of MTT, cells incubated for 4 hours at 37°C in CO₂ incubator.
7. Observed the wells for formazan crystal formation under microscope. The yellowish MTT was reduced to dark colored formazan by viable cells only.
8. After removing the medium completely. Added 200 μ l of DMSO (kept for 10 min) and incubate at 37°C (wrapped with aluminum foil).

9.

RESULTS:**i) Results of Phytochemical Test for Sample VV & TL: [41-44]****Table: 2 Results of Phytochemical Test**

Sr. No.	Chemical Constituents	Tests	TL	VV	Observation
1.	Steroids	Salkowski test	++	--	Present in TL,Partially in VV
		Liebermann Burchard test	++	++	
		Sulphur test			
2.	Glycosides	Balget's test			Present in TL Partially in VV
		Keller killiani test	++	--	
		Legal's test	++	++	
3.	Alkaloids	Wagner's test			Present in TL,Partially in VV
		Hager's test			
		Mayer's test	++	--	
4.	Flavonoids	Lead acetate	--	++	Partially present in TL and Moderate in VV
		Zinc hydrochloride	--	--	
		5% Ferric chloride test	++	--	
		Alkaline reagent	--	++	
5.	Phenol	Shinoda Test	--	++	Present in Both
		Ferric chloride test	++	++	
		Lead Acetate test	++	--	
		Ellagic Test	--	--	
6.	Caretinoids	Gelatin test	++	++	Present in VV
		Caretinoid test	--	++	



Figure 8: Alkaloid Test of Sample VV

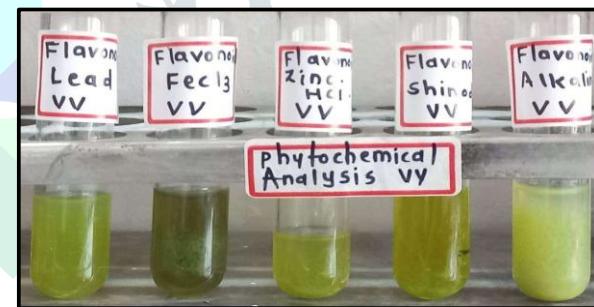


Figure 9: Flavanoid Test of Sample VV

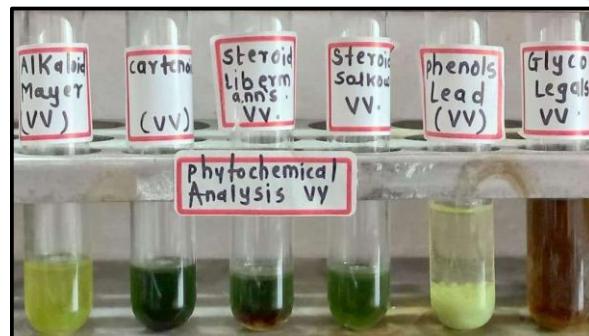


Figure 10: Steroidal Test of Sample VV

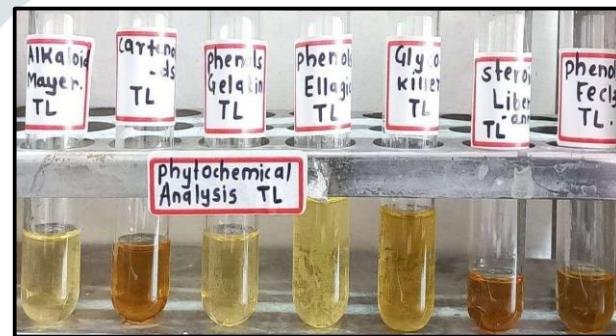


Figure 11: Alkaloid Test of Sample VV

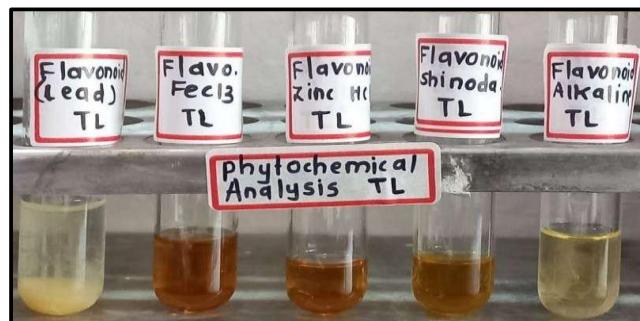


Figure 12: Alkaloid Test of Sample VV

ii) Anticancer Activity:

Table: 3 Anticancer activity of samples on Breast Cancer cell line [45-49]

S. No.	Sample Code	Conc. (ug/ml)	OD			Mean	% of Inhibition	% of Viability	IC50 (uL/ml)
1	Control		1.307			-	-	-	-
2	Standard (%Fluorouracil)	20	1.007	1.008	1.007	1.007	22.95%	77.05%	
		40	0.891	0.890	0.891	0.891	31.82%	68.18%	49.23
		60	0.551	0.550	0.551	0.550	57.91%	42.08%	
		80	0.436	0.436	0.437	0.436	66.64%	33.35%	
		100	0.279	0.278	0.279	0.278	78.72%	21.27%	
3	VV	20	1.168	1.168	1.168	1.168	10.63%	89.36%	NE
		40	1.054	1.054	1.055	1.054	19.35%	80.64%	
		60	0.920	0.920	0.920	0.920	29.60%	70.39%	
		80	0.840	0.842	0.840	0.840	35.73%	64.26%	
		100	0.796	0.796	0.796	0.796	39.09%	60.90%	
4	TL	20	1.206	1.208	1.206	1.206	7.72%	92.27%	NE
		40	1.008	1.007	1.008	1.007	22.95%	77.04%	
		60	0.956	0.954	0.954	0.954	27.00%	72.99%	
		80	0.812	0.810	0.812	0.811	37.94%	62.05%	
		100	0.754	0.754	0.756	0.754	42.31%	57.68%	
5	VT	20	1.005	1.005	1.005	1.005	23.10%	76.89%	54.25
		40	0.920	0.922	0.920	0.920	29.60%	70.39%	
		60	0.825	0.825	0.825	0.825	36.87%	63.12%	
		80	0.710	0.712	0.712	0.711	45.60%	54.39%	
		100	0.626	0.626	0.627	0.626	52.10%	47.89%	

*NE- Not Evaluable

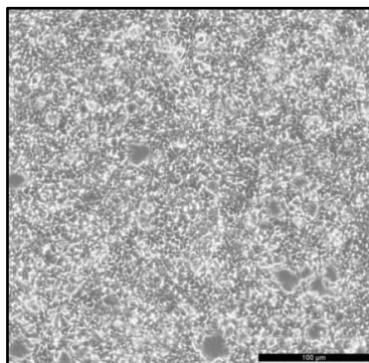


Figure 13: Control

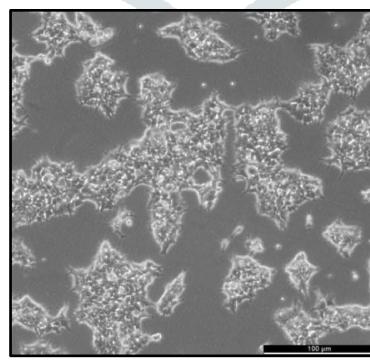


Figure 14: Standard

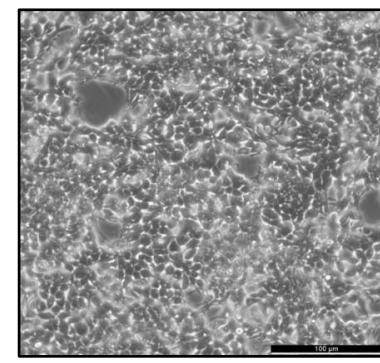


Figure 15: Effect of Sample VV

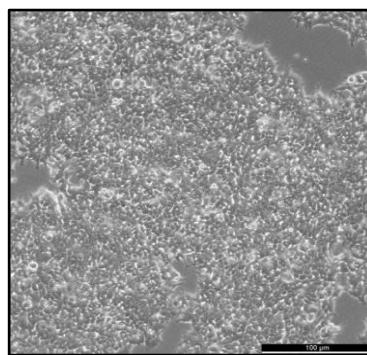


Figure 16: Effect of Sample TL

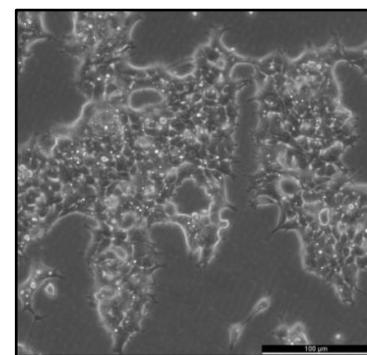


Figure 17: Effect of Sample VT

DISCUSSION:

This research holds the potential to pave the way for innovative and effective strategies in cancer treatment, benefiting the ever-increasing population affected by this devastating disease. At the different concentrations Sample Code VT showed High percent of inhibition and having good anticancer activity against MCF7 Breast Cancer cell line as compared to standard drug 5FU and sample code VV, TL shows the low percent of inhibition and shows the moderate anticancer activity. In this research work, we understood the synergistic potential of these plant-derived compounds opens avenues for developing novel and stable anti-cancer formulations that capitalize on their combined benefits. Further exploration of the underlying molecular mechanisms and *in vivo* studies is warranted to validate and optimize this synergistic approach for potential therapeutic applications in cancer treatment. As well as clinical trials to assess the safety and efficacy of the combined plant extracts in human subjects, moving towards potential therapeutic applications.

ACKNOWLEDGEMENT

Authors are grateful to Tripura University, Faculty of Pharmacy, for providing all the facilities to carry out this research work.

DECLARATION

Conflict of interest

The authors state that there is no conflict of interest with this study.

Funding

None declared.

REFERENCES:

- [1] Ataollahi M, Paknahad M, Paknahad A, 2015, Brest cancer and associated factors: a review, *Journal of Medicine and Life*, 2015, 8(4), pp. 6 – 11.
- [2] Shah R, Rosso K, Nathanson S, 2014, Pathogenesis, prevention, diagnosis and treatment of brest cancer, *World Journal of Clinical Oncology*, 5(3), pp. 283 – 298.
- [3] Hazwani A, Abdul R, Mat I, 2010, *Catharanthus roseus* Aqueous Extract is Cytotoxic to Jurkat Leukaemic T-cells but Induces the Proliferation of Normal Peripheral Blood Mononuclear Cells, *Tropical Life Science Research*, 21(2), pp. 101 – 113.
- [4] Marcelo F, Yoram K, Hinanit K, 2015, Plant derived substances with anti- cancer activity: from folklore to practice, *Frontiers in Plant Science*, 6(799), pp. 1 - 9.
- [5] Murugan P, Salim A, Chenthamara D, *et al*, 2021, Therapeutic and pharmacological efficacy of selective Indian medicinal plants –A review, *Phytomedicine Plus* 1, 100029, pp. 1 – 28.
- [6] Somashekara R, Dondapati R, Mullahalli V, *et al*, 2022, Biological isolation and characterization of *Catharanthus roseus* (L.) G. Don methanolic leaves extracts and their assessment for antimicrobial, cytotoxic, and apoptotic activities, 22, pp. 1 – 18.
- [7] Reza B, Tina S, Narges B, *et al*, 2017, Combination therapy in combating cancer, 2017, 8(23), *Journal of Oncotarget*, pp 38022 – 38043.
- [8] Skok Z, Zidar N, Kikelj J, 2020, Dual Inhibitors of Human DNA Topoisomerase II and other cancer related targets, *Journal Medicinal Chemistry*, 63, pp. 884 – 904.

- [8] Elisha S, Michal L, Sarah S, *et al*, 2014, Evaluating Medicinal Plants for Anticancer Activity, *Journal of Science World*, pp. 1 – 12.
- [9] Greenwell M, Rahman P, 2015, Medicinal Plants: Their use in anticancer treatment, *International Journal Pharmaceutical and Scientific Research*, 6(10), pp. 4103 – 12.
- [10]Shukla P, Bajpai K, Tripathi S, *et al*, 2013, A Review on the Taxonomy, Ethnobotany, Chemistry and Pharmacology of Solanum Lycopersicum Linn, *International Journal of Chemistry and Pharmaceutical Sciences*, 1(8), pp. 521-27.
- [11]Chaudhary P, Sharma A, Singh B, *et al*, 2018, Bioactivities of Phytochemicals Present in Tomato, 55(8), pp 2833-49.
- [12]Wang J, Wang S, Chu P, *et al*, 2021, Investigating into Anti-Cancer Potential of Lycopene: Molecular Targets, *Biomedicine & Pharmacotherapy*, 138, pp 1-11.
- [13]Mishra J, Verma N, 2017, A Brief Study on Catharanthus Roseus: A Review, *International Journal of Research In Pharmacy and Pharmaceutical Sciences*, 2(2), 20-23.
- [14]Chandrasekaran N, Vanitha M, Kaavyalakshmi M, 2014, Extraction of Vindoline From Catharanthus Roseus and Instrumental Model of Automatic Column Chromatography By Using PLC, *International Journal of Chemtech Research*, 6(9), pp 4216-4224.
- [15]Dubey A, Tiwari D, Srivastava K, 2020, A Discussion on Vinca Plant, *Journal of Pharmacognosy and Phytochemistry*, 9(5), pp 27-35.
- [16]Kaushik S, Tomar R, Gupta M, *et al*, 2017, An Overview of Catharanthus Roseus and Medicinal Properties of Their Metabolites Against Important Diseases, *European Academic Research*, 2, pp 1243-44.
- [17]Dajah A, Ahmad M, Khafajah A, 2019, Extraction, Purification, and Characterization of Lycopene From Jordanian Vine Tomato Cultivar, and Study of Its Potential Natural Antioxidant Effect on Samen Baladi, *Food and Nutrition Journal*, 7(2), pp 532-46.
- [18]Ingle K, Deshmukh A, Padole D, *et al*, 2017, Phytochemicals: Extraction Methods, Identification and detection of Bioactive Compounds From Plant Extracts *Journal of Pharmacognosy And Phytochemistry*, 1(6), pp 33.
- [19]Daga A, Tope T, Shukre V, *et al*, 2021, Extraction and Separation of Lycopene from natural sources using Chromatographic technique and its Application, *International Journal of Creative Research Thoughts*, 9(4), pp 723-33.
- [20]Pathak S, Akhade O, Bhojane K, *et al*, 2020, Evidences and prospects of Lycopene as powerful red superfood: Modern approach to food science, *International Journal of Research And Review*, 7(4), pp 487-490.
- [21]Gopalasatheeskumar K, 2018, Significant Role of Soxhlet Extraction Process in Phytochemical Research, *Journal of Pharmaceutical and Medical Sciences*, 7(Suppl 1), pp 43-47.
- [22]Singha R, Mishra V, Das R, 2018, IOP Conference Series: Materials Science And Engineering, 3, pp 1 – 4.
- [23]Halder J, Srivastava C, Dhingra S, *et al*, 2009, Antifeedant Activity of Methanolic Extracts of Vinca Rosea and Callistemon Lanceolatus Against Helicoverpa Armigera, *Sci. Research Gate Publication*, 17(2), pp 459-526.
- [24]Chandrasekaran N, Vanitha M, Kaavyalakshmi M, 2014, Extraction of Vindoline From Catharanthus roseus and instrumental method of Automatic Column Chromatography by using PLC, *International Journal of Chemtech Research*, 6(9), pp 4216 -24.
- [25]Tyler V, Brady L, Robbers J, Pharmacognosy K. M. Varghese Company, 8th Edn.1981, 67.
- [26]Pharmacopoeia of India.4th Edition ministry of health, Govt. of India, 1996, A-114.
- [27]Pulok M, 2002, Quality Control of Herbal Drugs, New Delhi, pp 187-96.
- [28]Khandelwal K, 2003, Practical pharmacognosy techniques and experiments, 10th Edition, pp 149-54.
- [29]Kokate C, Purohit A, Gokhale S, 2004, Pharmacognosy. Nirali Prakashan, Pune. 27th Edition, pp. 106.
- [30]Khandelwal KR, 2002, Practical Pharmacognosy, Nirali Prakashan, Pune. 2nd edition. pp. 149-55.
- [31]Basic Biology and Clinical Assessment, Stephen-2005, pp. 343- 349.
- [32]MTT Assay Protocol For Cell Viability And Proliferation By Sigma Pvt. Ltd.
- [33]Mosmann T, 1983, Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays, *Journal of Immunological Methods*, 65(1-2), pp. 55-63.
- [34]Hansen M, Nielsen S, Berg K, 1989, Re-examination and further development of a precise and rapid dye method for measuring cell growth/cell kill, *Journal of Immunological Methods*, 119(2), pp 203-10.
- [35]Ahuja S, Panchal N, Fidai R, 2024, Fabrication of Carboxymethyl Cellulose/Gelatin Polymeric Composite Film Incorporated With Calcium Nanoparticles and Catharanthus Roseus, *International Journal Biology, Pharmacy, and Allied sciences*, 13(1), pp 403-12.

[36]Ataollahi M, Sharifi J, Paknahad M, 2015, Breast cancer and associated factors: a review, *The Journal of Medicine and Life*, 8(4), pp. 6–11.

[37]Sathelly K, Kalagatur N, Mangamuri U, *et al*, 2023, Anticancer Potential of *Solanum Lycopersicum* L. Extract in Human Lung Epithelial Cancer Cells A549, *Indian Journal of Biochemistry & Biophysics*, 60, pp 76-85.

[38]Islam S, Lucky R, 2019, A Study on Different Plants of Apocynaceae Family and Their Medicinal Uses, *Universal Journal of Pharmaceutical Research*, 4(1), pp. 40-44.

[39]Yuan F, Wang Q, Pan Q, 2011, an Efficient Somatic Embryogenesis Based Plant Regeneration From the Hypocotyls of *Catharanthus Roseus*, *African Journal of Biotechnology*, 10(66), pp. 14786-95.

[40]Jordan M, Thrower D, Wilson L, 1991, Mechanism of inhibition of cell proliferation by Vinca alkaloid, 51(8), pp. 2212-22.

[41]Almagro L, Fernández-Pérez F, Pedreño M, 2015, Indole Alkaloids From *Catharanthus Roseus*: Bioproduction And Their Effect on Human Health, *Molecules*, 20(2), pp. 2973-3000.

[42]Maqsood M, Abdul M, 2017, Yeast Extract Elicitation Increases Vinblastine And Vincristine Yield In Protoplast Derived Tissues And Plantlets In *Catharanthus Roseus*, *Brazilian Journal of Pharmacognosy*, 27(5), pp. 549-56.

[43]Bhuvaneswari V, Nagini S, 2005, Lycopene: a review of its potential as an anticancer agent, *Current Medicinal Chemistry-Anticancer Agents*, 5, pp. 627–35.

[44]Karas M, Amir H, Fishman D, *et al*, 2000, Lycopene interferes with cell cycle progression and insulin-like growth factor I signaling in mammary cancer cells, *Journal of Nutrition and Cancer*, 36, pp. 101–11.

[45]Assar E, Vidalle M, Chopra M, *et al*, 2016, Lycopene acts through inhibition of I_KB kinase to suppress NF B signaling in human prostate and breast cancer cells, *Journal of the International Society for Onco developmental Biology and Medicine Journal*, 37, pp. 9375–85.

[46]Gloria N, Soares N, Brand C, *et al*, 2014, Lycopene and beta-carotene induce cell-cycle arrest and apoptosis in human breast cancer cell lines, *Journal of Anticancer Resarch*, 34, pp 1377–86.

[47]Jain A, Sharma G, Ghoshal G, *et al*, 2018, Lycopene loaded whey protein isolate nanoparticles: an innovative endeavor for enhanced bioavailability of lycopene and anti-cancer activity, *International Journal of Pharmacy*, 546, pp 97–105.

[48]Agarwal S, Jacob S, Chettri N, *et al*, 2011, Evaluation of In-vitro Anthelmintic Activity of *Catharanthus roseus* Extract, *International Journal of Pharmaceutical Sciences and Drug Research*, 3(3), pp 211-13.